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2
3 THE USE OF PHARMACOKINETICS TO
4 OPTIMISE DOSING AND REFORMULATION OF
5 ANTIRETROVIRALS FOR TREATMENT AND
6 PREVENTION.

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9 *THESIS SUBMITTED IN ACCORDANCE WITH THE REQUIREMENTS OF THE*
10 *UNIVERSITY OF LIVERPOOL FOR THE DEGREE OF DOCTOR IN PHILOSOPHY BY*

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16 *SEPTEMBER 2017 [CORRECTIONS APRIL 2018]*

1 TABLE OF CONTENTS

2 1 INTRODUCTION..... 2-8

3 1.1 HIV: NATURAL HISTORY OF THE DISEASE. EVOLUTION FROM DISCOVERY TO DATE .. 2-8

4 1.1.1 *Discovery of Disease epidemic* 2-8

5 1.1.2 *A Syndrome; Search for a Cause* 2-10

6 1.2 THE SCOPE OF THE CHALLENGE..... 2-13

7 1.3 LONG-ACTING INJECTABLE HIV PREVENTION: PHARMACOKINETIC PROOF-OF-CONCEPT 8 2-18

9 1.4 INITIATION OF DUAL THERAPY IN ANTIRETROVIRAL NAÏVE PATIENTS? 2-21

10 1.5 A NOVEL METHOD FOR PRODUCING SOLID-DOSE NANOPARTICLE FORMULATIONS; 11 EXPLORATION OF THEIR POTENTIAL FOR DOSE REDUCTION OF LICENSED ORAL THERAPIES. 2-25

12 2 SSAT040: LONG-ACTING RILPIVIRINE 3-27

13 2.1 BACKGROUND 3-27

14 2.2 METHODS 3-29

15 2.2.1 *Protocol development*..... 3-30

16 2.2.2 *Study population and randomization*..... 3-33

17 2.2.2.1 Inclusion Criteria 3-34

18 2.2.2.2 Exclusion Criteria..... 3-36

19 2.2.3 *Study design* 3-38

20 2.2.4 *Administered product*..... 3-38

21 2.2.5 *Sample collection*..... 3-39

22 2.2.5.1 Plasma..... 3-39

23 2.2.5.2 CVF..... 3-40

24 2.2.5.3 Rectal Fluid 3-41

1	2.2.5.4	Tissue	3-41
2	2.2.6	<i>Analytical Methods</i>	3-42
3	2.2.6.1	Pharmacokinetics.....	3-42
4	2.2.6.2	Pharmacodynamics	3-44
5	2.2.7	<i>Data Analysis</i>	3-45
6	2.3	RESULTS.....	3-47
7	2.3.1	<i>Participant Demographics, Disposition and Safety</i>	3-47
8	2.3.2	<i>Rilpivirine Plasma Pharmacokinetics</i>	3-50
9	2.3.3	<i>Cervico-Vaginal Fluid Pharmacokinetics</i>	3-60
10	2.3.4	<i>Rectal Fluid and Tissue concentrations</i>	3-60
11	2.3.5	<i>Vaginal Tissue</i>	3-67
12	2.3.6	<i>Covariate Analysis</i>	3-67
13	2.3.7	<i>Viral inhibition in cervicovaginal lavage</i>	3-70
14	2.4	DISCUSSION	3-75
15	3	A CASE OF HIV SEROCONVERSION IN SSAT040	4-81
16	3.1	MATERIALS AND METHODS.....	4-84
17	3.1.1	<i>Resistance analysis</i>	4-84
18	3.1.2	<i>Phenotypic analysis</i>	4-86
19	3.2	RESULTS.....	4-86
20	3.2.1	<i>HIV-1 Infection and RPV Levels</i>	4-86
21	3.2.2	<i>Selection of HIV-1 Drug Resistance</i>	4-92
22	3.2.3	<i>Cross-resistance to NNRTIs</i>	4-92
23	3.2.4	<i>Discussion</i>	4-93
24	4	SSAT049: RILPIVIRINE AND BOOSTED-DARUNAVIR DUAL THERAPY.....	5-99

1	4.1	BACKGROUND	5-99
2	4.2	METHODS	5-104
3	4.2.1	<i>Participants</i>	5-104
4	4.2.2	<i>Study design</i>	5-105
5	4.2.3	<i>Bioanalysis (drug plasma concentration measurement)</i>	5-106
6	4.2.4	<i>Data analysis</i>	5-107
7	4.3	RESULTS	5-108
8	4.3.1	<i>Viral load dynamics</i>	5-110
9	4.3.2	<i>Pharmacokinetics</i>	5-115
10	4.3.3	<i>Safety and tolerability</i>	5-119
11	4.4	DISCUSSION	5-119
12	5	SSAT055: NANO-EFAVIRENZ AND NANO-LOPINAVIR.....	6-123
13	5.1	BACKGROUND	6-123
14	5.2	METHODS	6-128
15	5.2.1	<i>Participants</i>	6-128
16	5.2.2	<i>Pharmacogenomics</i>	6-129
17	5.2.3	<i>Bioanalysis (Drug plasma concentration measurement)</i>	6-129
18	5.2.4	<i>Bioanalysis (Pharmacogenetic Analysis)</i>	6-130
19	5.2.5	<i>Study Enrolment</i>	6-131
20	5.2.6	<i>(NANO-EFV) Group A: Primary stage Pharmacokinetic Protocol.....</i>	<i>6-</i>
21		<i>133</i>	
22	5.2.7	<i>(NANO-LPV) Group B Primary stage: Pharmacokinetic Protocol</i>	<i>6-134</i>
23	5.3	PRIMARY STAGE RESULTS	6-136

1	5.3.1	Group A Primary Stage (NANO-EFV)	6-136
2	5.3.2	Group B Primary Stage (NANO-LPV)	6-140
3	5.4	MODELLING THE PRIMARY STAGE RESULTS TO INFORM PROTOCOL STEERING	
4	COMMITTEE		6-144
5	5.4.1	Protocol Steering Committee	6-147
6	5.5	SECONDARY STAGE: NANO-LOPINAVIR	6-149
7	5.5.1	Results.....	6-150
8	5.6	DISCUSSION	6-153
9	6	SUMMARY	7-156
10	7	ACKNOWLEDGEMENTS	8-161
11	8	REFERENCES	9-165
12			
13			

1 GLOSSARY

3TC	lamivudine
Ag	antigen
AIDS	acquired immunodeficiency syndrome
AUC	area under the concentration/time curve
AZT	azidothymidine (see also ZDV)
BMI	body mass index
cART	combination antiretroviral therapy
CDC	US Centers for Disease Control
Cmax	maximum concentration after dosing
Ctrough	concentration at the end of dosing interval
CV	coefficient of variation
CVF	cervicovaginal fluid
CVL	cervicovaginal lavage
d4T	stavudine
DNA	deoxyribose nucleic acid
DRV	darunavir
ECG	electrocardiograph
EFV	efavirenz
ETR	etravirine
FTC	emtricitabine
GM	geometric mean
GMR	geometric mean ratio
HIV	human immunodeficiency virus
IM	intramuscular
LLQ	lower limit of quantification
LPV	lopinavir
MSM	men who have sex with men
NNIBP	non-nucleoside inhibitor binding pocket
NNRTI	non-nucleoside reverse transcriptase inhibitor
NRTI	nucleoside reverse transcriptase inhibitor
NVP	nevirapine
PEP	post-exposure prophylaxis
PrEP	pre-exposure prophylaxis
PSC	Protocol Steering Committee
QC	quality control
RAL	raltegravir
RF	rectal fluid
RLU	relative luminometer units
RPV	rilpivirine

RPV-LA	long-acting rilpivirine
RT	reverse transcriptase
rtv	ritonavir
SDN	solid-drug nanoparticle
SSAT	St. Stephens AIDS Trust
TDF	tenofovir disoproxil fumarate
Tmax	time to maximum concentration after dosing
TZM-bl	cell type used in HIV-1 endpoint neutralisation assay. TZM-bl cell line is derived from a HeLa cell clone that was engineered to express CD4, CCR5 and CXCR4
VL	viral load
WHO	World Health Organisation
ZDV	zidovudine

1 2 INTRODUCTION

2

3 2.1 HIV: NATURAL HISTORY OF THE DISEASE. EVOLUTION FROM DISCOVERY TO DATE

4

5 2.1.1 DISCOVERY OF DISEASE EPIDEMIC

6

7 This work addresses the development of pharmacological therapeutic agents to
8 enable better treatment of people infected with the Human Immunodeficiency Virus
9 (HIV). The specific challenge in both the prevention and treatment of this condition
10 marks one of the great successes of the last fifty years, but yet remains an
11 incomplete solution to the problem with many people still living with the
12 manifestation of the infection, Acquired Immunodeficiency Syndrome (AIDS), or
13 dying from its complications.

14 It is almost four decades since the attention of the public and the media was
15 drawn to a seeming new disease syndrome with alarming lethality and the ability to
16 spread amongst the young population. The emergence of HIV/AIDS to become a
17 global pandemic represented a most significant social and biological challenge, a
18 disorder of uncertain aetiology, but embedded in interpersonal behaviour. Once
19 the existence of the condition and the uncertainty surrounding its nature was
20 acknowledged it rapidly caught the attention of the public and media, clinicians and
21 scientists. Even before the discovery of the causative agent, there was a high index

1 of suspicion that the clinical syndrome which came to be known as AIDS (Acquired
2 Immunodeficiency Syndrome), was caused by a highly infectious agent. Classical
3 epidemiology indicated the likely infectious nature, based upon its pattern of spread
4 and concurrent reports of outbreaks in geographically divergent locations. While it
5 remained possible that the underlying cause of the syndrome might be related to
6 environmental exposure to chemical toxins, recreational drug use or radioactive
7 material, there was high suspicion of the causative agent being a zoonotic virus, and
8 therefore in principle being amenable to therapeutic intervention with a suitable
9 pharmacological compound. (Gazzard 1988; Dalglish et al. 1995)

10

11 The aggressive nature of the natural of natural history of HIV was vividly evident
12 during the first two decades when uncontrolled it became prevalent amongst
13 infected patient populations. The considerable challenge was of a disease with
14 protean and unusual manifestations system, in which no organ or tissue was
15 seemingly unaffected. The obvious cachectic loss of weight, together with severe
16 constitutional symptoms could be related directly to a variety of causes of
17 gastrointestinal disintegration. However, differentiating the secondary
18 opportunistic infections and malignancies from the primary cause associated with a
19 profoundly immune-depleted state required careful clinical observation and
20 investigation. In time, these gave indication that any putative causative pathogen
21 must display a negative lymphotropic action, wherein the very orchestrators of the
22 immune system was disrupted. Thus, the defensive surveillance against invading
23 pathogens depended were potential the target for destruction; CD4 cell-membrane

1 bearing T-lymphocytes and some macrophages. This would come to define the
2 fundamental nature of the challenges faced in all efforts that have been taken to
3 date to combat the disease. This demonstration of this fact represented a major
4 step forward as it provided an accessible, and readily available early biomarker for
5 the presence and progress of the disorder within the routine clinical context.. By
6 indicating the extent of depletion of the circulating pool of helper T cells, it allowed
7 both a presumptive diagnosis of the presence of this syndrome as well as a
8 statement about the extent or severity to which the immune system had already
9 been damaged. It allowed an immediate assessment and also provided a means
10 through which the variable progression of the disorder could be followed over time

11

12 **2.1.2 A SYNDROME; SEARCH FOR A CAUSE**

13

14 The early epidemiology identified clusters of people at high risk of the disease,
15 including men who have sex with men, heroin users, haemophilia patients and US
16 immigrants from Haiti. A focus on these groups led to the identification of the
17 causative agent as a slow-incubating *Lentivirus*, a genus of the family of *Retroviridae*
18 in 1983. This was in the year after the US Centers for Disease Control (CDC) had
19 named the syndrome AIDS, confirming that different appellations given to each
20 observed cluster, were indications of a common disease process.

21 In common with all retroviruses, a *lentivirus* is a single-stranded positive sense
22 RNA virus contained within an envelope, which in order to replicate by integration

1 of a nuclear DNA intermediate into host chromosomes, first requires transcription of
2 the positive RNA strand into double-stranded DNA by a virally-encoded reverse
3 transcriptase (RT) enzyme in the cytoplasm. This RT enzyme, which accompanies the
4 viral genome within the infectious particle both reverse transcribes the
5 complementary DNA (cDNA) strand from the viral RNA, before acting as a DNA-
6 dependent DNA polymerase to transcribe the positive sense strand from this cDNA
7 to form the double stranded pair. Simultaneously, its ribonuclease activity due to a
8 domain RNAase H, degrades the RNA strand whilst the cDNA strand is being reverse
9 transcribed. The lack of error-correction in this mechanism, underpins the
10 continuous generation of all possible mutations in infected cells in a host, the vast
11 majority of which result in defective “junk” gene transcripts which results in self-
12 termination of the replication cycle. Where mutations occur, either singly or in
13 combination, which are compatible with replication their proportionate “success” in
14 efficiency of replication relative to un-mutated wild-type virus transcripts prevents
15 their domination of the heterogenous pool of virions, without the selective pressure
16 of drug therapy, capable of interrupting the cycle.

17 Trials in the early 1990s, illustrated the initial success of early nucleotide reverse
18 transcriptase inhibitors, which proved to be transient as the selective pressure
19 provided by just one therapeutic molecule, allowed viral escape with emergence of
20 mutations which retained replicative capacity in the presence of that drug. Even
21 though early studies were conducted prior to the general availability of
22 biotechnologic assays to detect and quantify the presence of virus in clinical
23 samples, this viral escape could was indirectly reflected in recrudescence of prior

1 AIDS-defining clinical syndromes and laboratory indication of loss of circulating
2 lymphocyte subsets.

3

4 It is possible to target each step of viral replication, from its first extracellular
5 interaction with host cell membrane glycoproteins, through cell-wall fusion and
6 internalisation of the viral particle in an uncoating process, at cytosolic reverse
7 transcription followed by intra-nuclear chromosomal integration utilising virally
8 encoded enzymes, in the post-translational assembly of an immature virion, with
9 protease cleavage and steps of budding from the cytosol into extracellular space
10 and the subsequent maturation steps for the viral particle to acquire full infectivity.

11 The ability to use small molecules interrupt this step of reverse transcription has
12 been of importance since the very first earliest efforts to attenuate viral replication;
13 subsequently through their combination with other agents to achieve effective
14 suppression of replication to the extent that circulating viraemia could not be
15 detected in circulating plasma by standard clinically available assays.

16

17 Inhibition of the reverse transcription step has been accomplished by two
18 mechanisms of action.

19 The first, by direct covalent binding to the active site of the RT enzyme, by
20 synthetic nucleoside or nucleotide analogues resulting in highly effective
21 termination of elongation of the transcribed DNA chain. As a class, these are

1 collectively termed NRTI, reverse transcriptase inhibitors which are either
2 nucleoside or mono-phosphorylated nucleotide analogues. These require
3 intracellular activation, through 3 or 2 phosphorylation steps to form the active
4 moiety.

5 The second mechanism of RT inhibition involved the development of molecules
6 with the ability to bind to a region of the RT protein, adjacent to the active site in a
7 hydrophobic pocket in what is described as the palm region, which interacts
8 allosterically with the catalytic site. This region is common to all drugs of this class,
9 termed non-nucleoside reverse transcriptase inhibitors (NNRTI), which on binding
10 cause a conformational change at the binding site to form a pocket (the NNIBP –
11 non-nucleoside inhibitor binding pocket). Further, the adjacent structure of the RT
12 enzyme subunit also undergoes a destabilising structural adjustment, which alters
13 the shape and thus binding affinity of the catalytic site. This destabilisation of the
14 enzyme on the nucleic acid chain template, from which it transcribes, alters its
15 catalytic site affinity for nucleotide binding, thus attenuating its ability to transcribe
16 from viral RNA and effectively. This synergistic activity between NNRTI and NRTI is
17 the underlying reason why a combination of a dual NRTI backbone, with a NNRTI
18 third agent has proven to be an effective and durable mainstay of combination
19 antiretroviral therapy, despite all three agents targeting a single replicative step.

20

21 2.2 THE SCOPE OF THE CHALLENGE

22

1 Globally, the availability of such NNRTI based combinations have been an
2 important enabling factor of the ambitious goals, set by those working in the field of
3 HIV, to be achieved. These stakeholders, including policy makers, programmers,
4 national governments and community-based organisations, operate within the
5 framework of global HIV targets to systematically set a series of goals to combat the
6 epidemic, which have driven a successful agenda for global health.

7 In 2003, the first step towards a goal of universal access to HIV/AIDS prevention
8 and treatment was the "3 by 5" initiative. This was launched by UNAIDS and WHO
9 with a global TARGET to provide three million people living with HIV/AIDS in low-
10 and middle-income countries with antiretroviral treatment by the end of 2005. This
11 established the consensus principle of treatment as a human right, accessible to all
12 who needed it, regardless of their location in the world or their individual ability or
13 that of their country's health system to afford the purchase of life-preserving
14 medication.

15 Whilst the target of 3 million on therapy was missed by a wide margin, 1.3 million
16 people were able to start therapy, which itself tripled the number of people being
17 treated and is estimated to have prevented 250,000 to 350,000 AIDS-related deaths
18 worldwide. Even with this relative success, the scale and relative distribution of the
19 challenge of low coverage of treatment at the time is shown in Figure 2:A with vast
20 swathes of low and middle income African and Eurasian countries having less than
21 10% coverage.

22

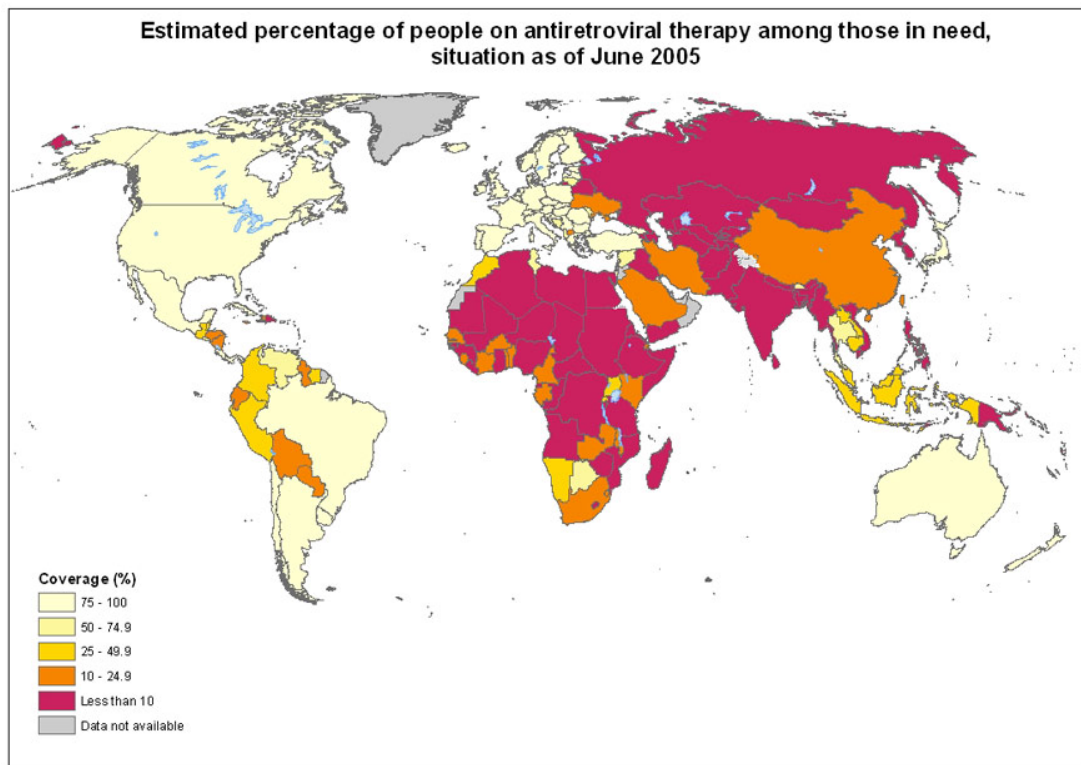


Figure 2:A WHO report on the global distribution of people living with HIV/AIDS on therapy in 2005.
(<http://www.who.int/hiv/facts/cov0605map/en/>) accessed September 2017

In the decade which followed, global stakeholders continued to work together to develop successive statements of agreed actions, with high-level signatories committing to coordinated efforts which acknowledged the iniquitous variability of the starting position, but resolved common goals across all territories (Table 2:A)

1

2

Table 2:A Progressive evolution of global framework for HIV/AIDS

2000	Millenium Development Goals target for 2015 included: <ul style="list-style-type: none">• Goal 6: Combat HIV/AIDS, malaria and other diseases
2006	UN General Assembly High-Level Meeting: <u>Political Declaration on HIV</u> <ul style="list-style-type: none">• review of the targets of the 2001 Declaration of Commitment on HIV/AIDS• countries signed the declaration to pledge their commitment to tackling the global HIV epidemic
2011	UN General Assembly: <u>60/262. Political Declaration on HIV/AIDS</u> <ul style="list-style-type: none">• reaffirmed the commitment to 2006 declaration• Set out the '15 by 15' target for 15 million people to be reached with HIV treatment by 2015
2011	UNAIDS developed ten interlinking targets in their <u>'Getting to Zero' strategy 2011-2015.</u> <ul style="list-style-type: none">• zero new HIV infections• zero AIDS-related deaths• zero discrimination.
2014	UNAIDS Fast track strategy; "90-90-90" 90 % targets for each step of: <ul style="list-style-type: none">• testing and linkage to care• initiation of therapy• effective therapy resulting in viral suppression

3

4 Progress against these stated goals has been impressive within a relatively short

5 timescale.

1 In 2015, of an estimated worldwide HIV prevalence of 36.7million the “15 by 15”
2 treatment target was reached nine months before the end of the year, with 17
3 million on treatment by the end of 2015.

4 However, beyond these successes, the ongoing magnitude of burden of disease
5 which saw an annual incidence of 2.1 million new HIV infections and 1.1 million
6 AIDS-related deaths, remained stark and sobering. In July of 2011, published
7 evidence from the HPTN053 confirmed the effectiveness of Treatment as
8 Prevention (TasP) in preventing onward sexual transmission between partners of
9 different serologic status. (Cohen et al. 2011) This evidence shows that both HIV
10 incidence and AIDS deaths reported in 2015 are unnecessarily high statistics which
11 would be sensitive to a further successes in the efforts to reduce the treatment gap
12 between those infected those on treatment.

13 Thus, the public health challenge in 2017 has shifted from the situation of
14 equipoise which existed in the early days of antiretroviral therapies which yielded
15 intolerable and sometimes life-limiting toxicities, which meant that healthcare
16 policies could cautiously to consider treatment.

17 Current evidence demonstrates that the balance, between the benefit of
18 effective viral suppression against any potential deleterious effects due to that
19 therapy, irrefutably supports immediate and ongoing treatment with suppressive
20 antiretroviral therapy. (Strategies for Management of Antiretroviral Therapy SMART
21 Study Group et al. 2006; Lodi et al. 2016) The current pharmacopeia provides varied

1 options to achieve this, with highly effective, well tolerated, simple therapies which
2 are resilient and tolerating of suboptimal dosing adherence.

3

4 Within the context of this global situation, as stated, the work in this manuscript
5 documents the exploration of three aspects of the existing topical challenges related
6 to HIV treatment and prevention. The tools and techniques available within the
7 study of clinical pharmacology are employed to address pragmatic clinical problems
8 which are common to a wide spectrum of healthcare delivery scenarios where
9 people living with HIV are managed.

10

11 **2.3 LONG-ACTING INJECTABLE HIV PREVENTION: PHARMACOKINETIC PROOF-OF-** 12 **CONCEPT**

13

14 As described above, despite the advances in diagnosis and treatment of HIV
15 infection, on a daily basis, new infections continue to occur in countries regardless
16 of their wealth and status. In the US and in Europe the licensure of
17 tenofovir/emtricitabine for HIV prevention provides a new and effective option for
18 HIV prevention. In spite of this development, it is clear that daily oral prophylaxis is
19 not suited for all populations in whom the risk of HIV acquisition may be elevated.
20 An example of this is the VOICE study, conducted in Sub-Saharan Africa, in which
21 over five thousand women were randomised to receive oral or topical tenofovir as

1 PrEP. In this study, neither of the products were demonstrated to be effective. It
2 became clear, through measurement of tenofovir in the plasma samples of
3 participants that it was detectable in fewer than a third of them, indicating low
4 levels of adherence. (Marrazzo et al. 2015) This effect was compounded by the fact
5 that those participants at highest risk of infection were also found to be at highest
6 risk of non-adherence to the products; women who were young and unmarried.
7 However, this demographic group is routinely adherent to intermittent injectable
8 progestins such as depot-medroxyprogesterone acetate, administered as a long-
9 acting reversible contraceptive. (Morrison et al. 2015) Qualitative surveys on the
10 attitudes and beliefs amongst populations at high risk of HIV infection indicate a
11 potential interest in a long-acting injectable, including other risk groups such as men
12 who have sex with men. (Eisingerich et al. 2012; Meyers et al. 2014)

13 Janssen Infectious Diseases BVBA, Belgium, the developers of the licensed oral
14 formulation of rilpivirine are in the process of conducting a development
15 programme of a nano-formulated suspension of rilpivirine as a long-acting
16 therapeutic antiretroviral. At the early stages of its development, Janssen formed a
17 collaborative agreement with the Bill and Melinda Gates Foundation (BMGF), in
18 which they transferred the rights to develop this formulation as an HIV pre-exposure
19 prophylactic to the Gates Foundation. In accordance with this agreement they
20 would have a supportive role, providing technical expertise and a transfer of data
21 and technology, but would not retain any rights to the commercialisation of long-
22 acting rilpivirine for any future prophylactic indication.

1 Prior to the eventual study plan, named SSAT040, the initial planned proposal to
2 study TMC-278 LA was a multi-centre study, with a single arm design which was
3 planned to be conducted at 4 clinical research sites in the United Kingdom; 3 sites in
4 London {St. Stephen's AIDS Trust as study sponsor, Imperial College NHS Trust and
5 St. Thomas' Hospital} as well as the Royal Sussex County Hospital in Brighton. This
6 initial study, given the code SSAT037, was developed as concept in 2009, receiving
7 regulatory and ethics approvals at the start of 2010. The study - registered
8 ClinicalTrials.gov Identifier NCT01049932, EudraCT: 2009-017631-17 – was designed
9 to explore the primary endpoint of safety of administration of intramuscular 600mg
10 TMC-278LA, after single dose with a second loading dose at 2 weeks and follow-up
11 to presumed steady state following monthly doses. The aim was to accumulate data
12 on 100 enrolled and evaluable subjects, with approximately 50 of African ancestry,
13 and 50 females. This would have provided 50-subject-years of safety data in order to
14 support and inform the design of a later large phase III global efficacy study.

15 As mentioned earlier, the parallel development paths of TMC-278LA for both HIV
16 treatment (Tibotec) and HIV prevention, in this first exploratory pilot study were
17 conducted simultaneously. In the first quarter of 2010, results from the ongoing
18 small pharmacokinetic study TMC278-TiDP15-C150 ClinicalTrials.gov Identifier:
19 NCT00741741 conducted by Tibotec Pharmaceuticals, indicated that the plasma
20 pharmacokinetic exposure obtained with a single 600mg dose, afforded plasma drug
21 exposure profiles which though in the desired therapeutic range, were lower than
22 expected based on preclinical modelling simulations.

1 After extensive discussion with the SSAT037 study steering group stakeholders,
2 the decision was made to terminate the study at a point after regulatory and ethical
3 approvals had been obtained, and volunteers had been screened for participation
4 but prior to any being enrolled.

5 Over the ensuing months in 2010, the study protocol was reviewed and
6 abandoned to be replaced by a new study design, SSAT040 which was to be an
7 adaptive design, dose-ranging exploratory PK study in healthy volunteers.

8 SSAT040 was the protocol designed to provide the first-in-man investigation of
9 the long-acting rilpivirine, defining its plasma pharmacokinetics at a range of doses,
10 whilst simultaneously exploring the compartmental pharmacokinetic exposures at
11 tissue sites relevant to the sexual acquisition of HIV; vagina, cervix and rectum.
12 Chapters 3 and 4 detail the results and conduct of this study.

13 **2.4 INITIATION OF DUAL THERAPY IN ANTIRETROVIRAL NAÏVE PATIENTS?**

14

15 Chapter 5 explores the potential utility of a conceptual approach to
16 rationalisation of therapy which challenges the prevailing body of evidence and
17 guidelines; is it feasible to use a two-drug combination of potent current-generation
18 therapies, normally used as the third agent alongside a nucleos(t)ide backbone, to
19 safely achieve viral suppression in HIV patients initiating treatment for the first time.
20 At the time of conception of this study ClinicalTrials.gov Identifier: NCT01736761 in
21 late 2012, the body of available evidence on this strategy was relatively limited and

1 the concept was unproven; Table 2 summarises the current published evidence on
2 two-drug regimens used in initiation of antiretroviral therapies.

3 Specifically, the SSAT049 study, was an investigator-led study conducted under
4 the regulatory sponsorship of the St. Stephen's AIDS Trust, in which the once-daily
5 oral combination of ritonavir-boosted darunavir and rilpivirine was studied in an
6 adaptive two-stage design. Funding for the study was provided by a grant from the
7 manufacturer, Janssen. The study aimed to carefully investigate this combination in
8 a controlled manner by utilising both sparse and intensive pharmacokinetics in the
9 first month of therapy, combined with real-time assessment of pharmacodynamic
10 outcome of viral decay to firstly assess the combination in patients whose circulating
11 viraemic load was below 100,000 copies/mL in an initial cohort of ten patients. The
12 guidance of a protocol steering committee, convened from experts with extensive
13 experience of both clinical and pharmacokinetic, was utilised to provide monitoring
14 oversight of the study and to make a "GO / NO GO" decision after the first ten
15 patients, to allow further enrolment of patients with viraemic burdens above the
16 threshold.

1

2

Table 2 Studies which evaluated nucleotide reverse transcriptase inhibitor-sparing regimens for initiation of therapy in naive patients..

Study name & design	Study size (N)	Dual therapy	Reference
<i>Boosted protease inhibitor plus integrase strand-transfer inhibitor</i>			
RADAR (open-label)	85	DRV/rtv/RAL Vs Control (DRV/rtv/TDF/FTC)	(Bedimo et al. 2014)
ACTG 5262 (open label)	112	DRV/rtv/RAL	(Taiwo et al. 2011)
NEAT001 (open-label)	805	DRV/rtv/RAL Vs DRV/rtv/TDF/FTC	{LambertNiclot:2016dl} {Bernardino:2015el}
SPARTAN (open-label)	94	Unboosted protease inhibitor plus integrase strand-transfer inhibitor ATV +RAL BiD Vs ATV/rtv/TDF/FTC	(Kozal et al. 2012)
PROGRESS (open-label)	206	LPV/rtv + RAL BiD Vs LPV/rtv/TDF/FTC	{Reynes:2011iv}{Reynes:2013ca}
<i>Boosted protease inhibitor plus non-nucleoside reverse transcriptase inhibitor</i>			
ACTG5142 (open-label)	753	LPV/rtv BiD + EFV Vs EFV/3TC/ {d4T or TDF or ZDV}	{Riddler:2008fv}

Study name & design	Study size (N)	Dual therapy	Reference
Or LPV/rtv/3TC/ {d4T or TDF or ZDV}			
<i>Boosted protease inhibitor plus CCR5 co-receptor antagonist</i>			
MODERN (Double Blind)	797	DRV/rtv/MVC Vs DRV/rtv/TDF/FTC	{Stellbrink:2016ee}
A4001078 (open-label)	121	ATV/rtv/MVC Vs ATV/rtv/TDF/FTC	{Mills:2013cs}
MIDAS (open-label)	64	MVC/DRV/rtv	{Taiwo:2013eh}
<i>Protease inhibitor plus 3TC</i>			
GARDEL (open-label)	373	LPV/rtv/3TC Vs LPV/rtv/2 NRTIs	{Cahn:2014hc}

1

2

3

4 **2.5 A NOVEL METHOD FOR PRODUCING SOLID-DOSE NANOPARTICLE FORMULATIONS;**
5 **EXPLORATION OF THEIR POTENTIAL FOR DOSE REDUCTION OF LICENSED ORAL**
6 **THERAPIES.**

7

8 In SSAT040 (Chapter 3), the potential of a solid-dose nanoparticle formulation of
9 rilpivirine was studied for its ability to provide extended pharmacokinetic coverage
10 when administered as a single-intramuscular injection.

11 Solid dose nano-formulations may also offer enhanced characteristics when
12 applied to orally administered drugs. The Chemistry and Clinical Pharmacology
13 groups at the University of Liverpool have developed a novel method of
14 manufacture of solid-dose nano-particle formulations which have been chemically
15 optimised and characterised using in-vitro tissue models of the interaction between
16 these cell based systems and these nano-particles. By integrating this in a whole
17 body mathematical model, they have been able to make a completely pre-clinical
18 prediction of the dose kinetics of the studied drugs as a starting point for clinical
19 phase studies. Chapter 6 details the ongoing study SSAT055, which explores in HIV-
20 negative healthy volunteers, the pharmacokinetic behaviour of nano-formulations of
21 the licensed antiretrovirals efavirenz and lopinavir (±ritonavir), comparing to the
22 standard oral formulations.

3 SSAT040: LONG-ACTING RILPIVIRINE

The candidate, AGAJ, acted as Chief Investigator for this study and was responsible for designing the concept, writing the protocol and lead the submissions for ethical and regulatory approvals. AGAJ established the local methodology for collection of samples and wrote standardisation manual and developed template proforma to ensure consistent collection of study samples and data. AGAJ was responsible for the clinical care of all participants on study, including informed consent and obtaining clinical history and assessing physical examination and determining eligibility and was responsible for administration of investigational product and collection of invasive samples. After bioanalysis was performed at the University of Liverpool bioanalysis facility - under GLP conditions - AGAJ was involved in compilation of the final results, presented the findings as an oral at the 2012 Conference on Retroviruses and Opportunistic Infection and wrote the manuscript as first author, which was published in Clinical Pharmacology and Therapeutics in June 2014.

3.1 BACKGROUND

Human Immunodeficiency virus (HIV) pre-exposure prophylaxis (PrEP) refers to a strategy involving the use of antiretroviral drugs to decrease the risk of HIV infection in uninfected individuals whose behaviour would combine with local HIV prevalence to place them at a high risk of infection. In 2012, the use of tenofovir disoproxil fumarate and emtricitabine (TDF/FTC) in combination was approved by the US Food and Drug Administration for use as PrEP, based on the results of the iPrEx (Grant et

1 al. 2010) (van der Straten et al. 2012) study and Partners PrEP, (Baeten et al. 2012)
2 with the former showing a 44% reduction in the incidence of HIV transmission in
3 men who have sex with men as compared with placebo treatment, when combined
4 with a comprehensive package of prevention. Partners PrEP showed a 67–75%
5 relative reduction in the incidence of HIV infection using TDF/FTC among
6 heterosexual couples in sexual partnerships containing one seronegative partner.

7 Although there is conceptual proof of PrEP in these specific contexts, recent
8 negative results of two studies in women, FEMPrEP (Van Damme et al. 2012) and
9 VOICE, (Marrazzo et al. 2013) showed no evidence of benefit of daily oral TDF/FTC.
10 These negative outcomes were later ascribed to suboptimal adherence to the
11 dosing regimen, thus indicating the need for high motivation in order to attain
12 prevention success. Therefore, durable adherence is critical for a successful long-
13 term prevention strategy. (Grant et al. 2010; Baeten et al. 2012; Marrazzo et al.
14 2013) In addition, the potential for side effects and toxicities associated with the use
15 of TDF/ FTC (Grant et al. 2010; Thigpen et al. 2012) remains a concern due to its
16 widespread administration as HIV PrEP.

17 An optimal PrEP therapy should be safe to administer and be readily distributed
18 to the relevant target tissues in concentrations that are sufficient to provide
19 protection against HIV infection. Ideally, PrEP agents should be characterized by
20 convenient dosing and by routes of administration that do not depend on the
21 recipient maintaining daily adherence to dosing. The nonnucleoside reverse-
22 transcriptase inhibitor RPV is a diarylpyrimidine derivative that was approved by the
23 Food and Drug Administration in 2011 for oral administration for the treatment of

1 HIV infection in combination with other ARV drugs.(Azijn et al. 2010; Ford et al.
2 2011) A parenteral formulation of rilpivirine (RPV-LA) with prolonged
3 pharmacokinetic (PK) exposure is being developed, enabling improved adherence to
4 ARV treatment over prolonged periods and having potential as an agent for HIV
5 PrEP.(Baert et al. 2009; Grant et al. 2010; van't Klooster et al. 2010) The potential
6 advantages of a long-acting formulation include infrequent parenteral
7 administration and a low potential for gastrointestinal side effects associated with
8 lifelong oral ARV intake.

9 For HIV prevention, it is important that the traditional sequence of the drug
10 development phases be followed as closely as possible to address whether a PrEP
11 agent is safe and effective for use in humans. However, dose optimization studies
12 are challenging because protective concentration targets in both plasma and the
13 genital/rectal compartments are unknown. Therefore, they must be inferred from
14 treatment efficacy studies, animal models, and/or ex vivo pharmacodynamic (PD)
15 experiments. We performed an adaptive design study to determine the plasma PK
16 of RPV-LA and to measure, for the first time, RPV concentrations in cervicovaginal
17 fluid (CVF), rectal fluid (RF), and tissue from the female genital tract and male
18 rectum, after i.m. administration of a range of doses to HIV-negative volunteers. The
19 study also aimed to assess the safety and tolerability of i.m. injections at 300, 600,
20 and 1,200 mg of RPV-LA and to determine the effect of RPV in genital fluid on HIV
21 replication ex vivo.

22 3.2 METHODS

1 **3.2.1 PROTOCOL DEVELOPMENT.**

2 This study was a phase I, prospective, openlabel, exploratory dose-ranging study
3 conducted at a single center (St. Stephen’s Centre clinical trial unit, Chelsea and
4 Westminster Hospital, London) with development of the protocol under the
5 regulatory sponsorship of the St. Stephen’s AIDS Trust. Funding for the study was
6 provided by a grant from the Bill & Melinda Gates Foundation with the engagement
7 of Tibotec (now Janssen R&D Infectious Diseases), which provided the
8 investigational agent, provided protocol oversight, and formed part of the protocol
9 steering committee. The protocol was approved by the Medicines Healthcare
10 Regulatory Agency, UK, and ethical approval was obtained, before commencement
11 of the study and after each protocol amendment, from the National Research Ethics
12 Service, UK. The protocol concept was developed as an adaptive exploratory design
13 with the aim of investigating RPV exposure in plasma, fluids, and tissues from the
14 female genital tract and male rectum at up to four different doses of RPV-LA
15 administered i.m.: 300 or 600 mg, with the option to explore either a 1,200 mg or a
16 150 mg dose, dependent on pharmacokinetic drug exposure attained with the
17 former doses relative to historic data with the licensed oral therapeutic dose (Figure
18 3:A SSAT040; Protocol Steering Committee algorithm).

SSAT040; guidance algorithm for Protocol Steering Committee

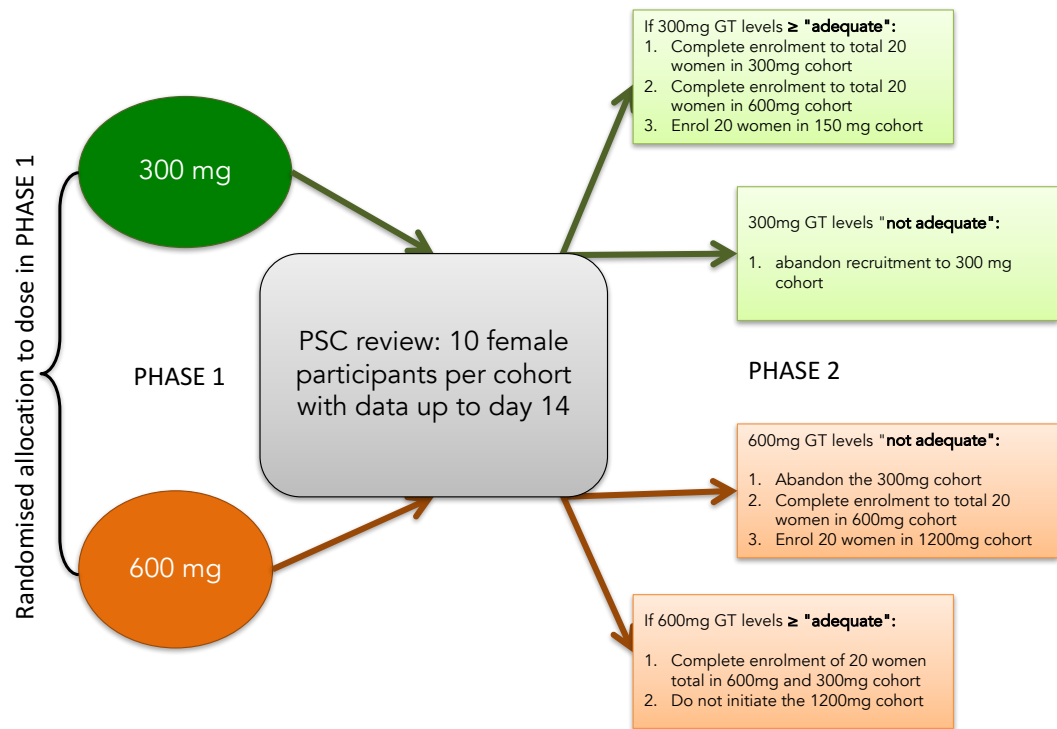


Figure 3:A SSAT040; Protocol Steering Committee algorithm

•The above figure was stated as guidance in the original protocol, approved by UK regulatory and ethics authorities, for the first Protocol Steering Committee (PSC) meeting.

•As approved, the protocol allowed the inclusion of 6 male volunteers, and 60 female volunteers total; in three dose groups of 20 each.

•PHASE 1 The first phase of study explored the plasma pharmacokinetics of 300mg (n=10) and 600mg (n=10), and the PSC was convened to compare the plasma exposure to day 28, comparing this for "adequacy" with a reference concentration from phase 3 treatment with oral rilpivirine of 50 ng/mL. Given that the data showed that 300mg was below this reference, with mean exposure for 600mg matching the reference range with no safety concerns the PSC deemed that the

1 second phase should proceed to enrol 10 volunteers to receive 1200mg and to
2 complete enrolment to the 600mg dose.

3 •PHASE 2.

4 •The second PSC meeting reviewed plasma data from the phase 2 - 1200mg and
5 600mg cohorts - in addition to complete plasma exposure from the first phase in
6 addition to genital compartment exposure in CV fluid. There were no further
7 concerns regarding safety data with 600mg or 1200mg and plasma exposure at
8 600mg matching the reference at day 28 whilst 1200mg dose exceeded this
9 concentration at days 28 and 56 following dose. The ratio of CV:plasma
10 concentrations was greater than 1.0 at all timepoints. Viral inhibition studies on
11 phases 1 & 2 were however unsuccessful, with directly aspirated vaginal fluid; the
12 viscosity of the fluid causing technical failure of the assay.

13 •The decision of the second PSC therefore was to complete enrolment to the
14 1200mg dose, with an amendment to the collection method for CV fluid at days 28
15 and 56 using a lavage method rather than direct aspiration, in order to facilitate the
16 viral inhibition assay.

17 •Given that there would be limited data available on viral inhibition at lower
18 concentrations than those achieved with 1200mg, the decision was taken to also
19 complete enrolment to the 300mg cohort with this amended collection method.

20

1 The study was conducted in three phases, with a review of data by the protocol
2 steering committee after each of the first two phases to determine the subsequent
3 protocol amendments. In the first phase, 6 male participants were recruited to
4 receive a single dose of 600 mg and 10 female participants in each cohort received a
5 single dose of 300 or 600 mg. After the first protocol steering committee review, the
6 second phase enrolled two cohorts of 10 women to receive either a 600 or a 1,200
7 mg single dose, and the third phase completed the study with a further two cohorts
8 of 10 women receiving either a 300 or a 1,200 mg single dose. In addition, on the
9 basis of the protocol steering committee meetings, changes were made to the
10 timing of vaginal tissue sampling for PK analysis (between phases) and to the timing
11 and method of collection of CVF samples taken for exploratory PD assessments.

12 **3.2.2 STUDY POPULATION AND RANDOMIZATION.**

13 Male and non-pregnant, non-lactating female participants were eligible for
14 enrolment if they provided written informed consent and met the eligibility criteria
15 which were designed and reviewed by regulatory and research ethics authorities
16 with the aim of recruiting healthy volunteers between the ages of 18 to 50 years
17 who were HIV negative at the screening visit and whose and whose behavioural
18 practices pertaining to risk, were likely to place them in the lowest category for HIV
19 acquisition, both in the 6 months prior to enrolment and for at least the duration of
20 study follow-up

21 Participants agreed to undergo regular HIV testing within the study and to
22 abstain from sexual intercourse for the first 28 days of the trial and for 48 h before
23 each subsequent study visit, before collection of genital or rectal samples.

1 The specific protocol defined eligibility criteria are detailed below.

2 3.2.2.1 ***Inclusion Criteria***

3

4 Participants must satisfy all of the following criteria within 42 days prior to the
5 baseline visit:

6 1. The ability to understand and sign a written informed consent form, prior to
7 participation in any screening procedures and must be willing to comply with all trial
8 requirements.

9 2. Non-pregnant, non-lactating females (at least 40% will be of self-identified
10 African ancestry)

11 3. Age between 18 to 50 years, inclusive.

12 4. Body Mass Index (BMI) of 16 to 35 kg/m², inclusive.

13 5. Negative antibody/antigen combined test for HIV1 and HIV2.

14 6. Absence of any significant health problems (in the opinion of the investigator)
15 on the basis of the screening procedures; including medical history, physical
16 examination, vital signs, ECG.

17 7. Willing to undergo HIV testing, HIV discussion and receive HIV test results
18 throughout the trial (according to the “UK National Guidelines for HIV Testing 2008”,
19 www.bhiva.org).

1 8. Women of childbearing potential (WOCBP) must be using an adequate method
2 of contraception (intrauterine device, condoms, anatomical sterility in self or
3 partner) to avoid pregnancy throughout the trial and for a period of at least four
4 months after the trial follow up visit (oral hormonal methods and implant
5 contraceptives are allowed but only in combination with the additional protection of
6 a barrier method). Males participating in sexual intercourse that could result in
7 pregnancy must use condoms during the duration of the study and for up to four
8 months following the follow up visit.

9 9. Willing to abstain from sexual intercourse (vaginal for females and receptive
10 anal for males) for 48 hours prior to each trial visit (with complete abstinence in the
11 first 28 days post-dose).

12 10. Females willing to refrain from the use of vaginal products or objects
13 including, tampons, female condoms, cotton wool, rags, diaphragms, cervical caps
14 (or any other vaginal barrier method), douches, lubricants, vibrators/dildos, and
15 drying agents for 14 days prior to enrolment and for the duration of the trial. Males
16 willing to refrain from the use of anal products or objects including douches,
17 lubricants and vibrators/dildos for 14 days prior to enrolment and for the duration
18 of the trial.

19 11. Likely to remain resident in the UK for the duration of the trial period.

20 12. Willing to consent to their personal details being entered onto The Over
21 volunteering Prevention Scheme (TOPS) database.

1 13. Willing to provide photographic identification at each visit.

2 14. Registered with a GP in the UK

3

4 3.2.2.2 *Exclusion Criteria*

5

6 1. Any significant acute or chronic medical illness.

7 2. Evidence of organ dysfunction or any clinically significant deviation from
8 normal in physical examination, vital signs, ECG or clinical laboratory
9 determinations.

10 3. Positive blood screen for syphilis, hepatitis A (IgM) B (HBs Ag) and/or C
11 antibodies.

12 4. Positive blood screen for HIV-1 and/or HIV-2 antibodies.

13 5. Positive screen for sexually transmitted infections at screening visit (if bacterial
14 vaginosis or candidiasis detected at screen, these may be treated with test-of cure
15 prior to enrolment).

16 6. Prolonged QT interval on screening ECG, or clinically significant change as
17 judged by investigator.

18 7. High-risk behaviour for HIV infection which is defined as having one of the
19 following within six months before trial day 0 (first dose):

- 1 i. had unprotected vaginal or anal sex with a known HIV infected person or a
- 2 casual partner.
- 3 ii. engaged in sex work for money or drugs.
- 4 iii. acquired a sexually transmitted disease.
- 5 iv. having a high-risk partner either currently or in the previous six months
- 6 8. Clinically relevant alcohol or drug use (positive urine drug screen) or history of
- 7 alcohol or drug use considered by the Investigator to be sufficient to hinder
- 8 compliance with treatment, follow-up procedures or evaluation of adverse events.
- 9 9. Exposure to any investigational drug or placebo within 30 days of first dose of
- 10 trial drug (additional check to be made on TOPS www.tops.org.uk).
- 11 10. History of severe drug allergy that in the opinion of the Investigator may
- 12 increase the risk of developing an allergic reaction to the trial drug.
- 13 11. Use of any drug, including over-the-counter medications and herbal
- 14 preparations, within two weeks prior to first dose of trial drug (unless approved or
- 15 prescribed by the Investigator (for exceptions see section 5.2).
- 16 12. Females who are pregnant or breast-feeding.
- 17 13. Clinically significant laboratory abnormalities (according to normal range
- 18 as defined by central laboratory).

19

1 Six male participants were recruited, and all received a single 600 mg dose.
2 Female participants were randomly assigned (within each study phase) to receive
3 either of the two doses being studied. A statistician created randomization lists,
4 stratified by phase, with blinded allocation maintained by means of sealed
5 envelopes kept in a restricted-access pharmacy. Once allocation was performed,
6 doses were administered on an open-label basis.

7 **3.2.3 STUDY DESIGN**

8 Eligible participants attended on the morning of day 0, when they received a
9 single i.m. dose of RPV-LA between 8 and 10 am. Plasma (predose and 4- and 8 h
10 postdose on day 0) and female genital or male RFs at 8 h postdose were collected
11 for RPV PK analysis. Paired plasma and genital or RF samples were then collected on
12 days 1, 3, 7, 11 (plasma only), 14, 21, 28, 42, 56, and 84 postdose. Paired tissue
13 biopsies were taken from the pericervical vaginal fornix in women or the rectal
14 mucosa in men at days 7 and 14, 14 and 28, or 28 and 56, depending on the study
15 phase (Table 1).

16 **3.2.4 ADMINISTERED PRODUCT**

17 RPV-LA (formulation G001), available as a nanoparticle suspension with a
18 concentration of 300 mg/ml, was given by i.m. injection into either buttock (300
19 mg—1 ml and 600 mg—2 ml) or both buttocks (1,200 mg—two 2 ml injections) on
20 day 0.

21 **3.2.5 ADVERSE EVENTS**

1 Adverse events were collected from the time of signing consent until each
2 participant ended their involvement in the study. Adverse events were graded in
3 accordance with the prevailing Division of AIDS Table for Grading of the severity of
4 Adult and Paediatric Adverse Events (Version 1.0 December 2004 – clarification
5 dated August 2009). Detail of the grading applied to events occurring at the
6 injection site are shown in Table 3.

7 ***Table 3 Excerpted grading scale from version 1.0 of DAIDS grading scale***
8 ***for adverse events, with specific detail of adverse events related to injection***
9 ***site reactions***

PARAMETER	GRADE 1 MILD	GRADE 2 MODERATE	GRADE 3 SEVERE	GRADE 4 POTENTIALLY LIFE- THREATENING
INJECTION SITE REACTIONS				
Injection site pain (pain without touching) or Tenderness (pain when area is touched)	Pain/ tenderness causing no or minimal limitation of use of limb	Pain/ tenderness limiting use of limb OR Pain/ tenderness causing greater than minimal interference with usual social and functional activities	Pain/ tenderness causing inability to perform usual social and functional activities	Pain/ tenderness causing inability to perform basic self- care function OR Hospitalisation (other than emergency room visit) indicated for management of pain/ tenderness
Adult > 15 years	Erythema OR Induration of 5x5 cm – 9x9 cm (or 25 cm ² – 81cm ²)	Erythema OR Induration OR Edema > 9 cm any diameter (or > 81 cm ²)	Ulceration OR Secondary infection OR Phlebitis OR Sterile abscess OR Drainage	Necrosis (involving dermis and deeper tissue)

10

11

12 **3.2.6 SAMPLE COLLECTION**

13 **3.2.6.1 Plasma**

1 At each scheduled time point, 6 ml whole blood was collected into lithium
2 heparin Vacutainer vials (BD Biosciences, San Jose CA) and immediately placed on
3 wet ice inside a light-protective container to prevent photo-degeneration before
4 centrifugation at 1,000g for 10 min at 4 °C (within 2 h). Plasma aliquots were stored
5 in light-protective polypropylene tubes at –20 °C until analysis.

6 3.2.6.2 *CVF*

7 Female participants collected samples by aspiration of vaginal secretions using a
8 self-inserted disposable, sterile plastic volumetric device (Rovumeter, University of
9 North Carolina School of Pharmacy)— a syringe-like device 135 mm long, with a
10 constant outer diameter of 8 mm and a blunt, rounded distal end at which a 5-mm
11 opening enables sample aspiration on applying suction to the plunger.

12 The undiluted aspirate was weighed, then chilled on wet ice in a light-protective
13 polypropylene cryovial, before storage at –80 °C until analysis. During study phase III
14 on days 0 (before dose), 28, and 56, CVL samples were collected, by aspiration after
15 lavage of the cervix and vaginal vault with 10 ml of normal saline, in order to
16 determine antiviral activity (PD analysis).

17 Therefore, at these study visits, direct aspiration of fluid was not performed in
18 order to maximize the sample yield collected during the lavage process; instead,
19 undiluted CVF was collected using Schirmer Tear-Test blotting paper strips (Intervet,
20 Roseland, NJ) applied to the high vaginal mucosa with a vaginal speculum in place.
21 The strips were weighed both before and after collection of the sample to enable

1 calculation of the volume of adsorbed fluid, and the strips were placed in a light-
2 protective cryovial for storage at –80 °C until analysis.

3

4 3.2.6.3 *Rectal Fluid*

5 Samples for RF PK analysis were collected from male participants, after
6 evacuating bowels, by adsorption onto Weck Cel cellulose spears (EYETEC; Network
7 Medical Products, North Yorkshire, UK) that were placed in contact with apposed
8 rectal mucosal surfaces (using a proctoscope) for at least 120 s; dry and wet weights
9 were used to calculate the volume of fluid collected, and spears were placed inside
10 light-protective cryovials for storage at –80 °C until analysis.

11

12 3.2.6.4 *Tissue*

13

14 Vaginal tissue was collected by biopsy; after insertion of the speculum, a 3 × 3 × 1
15 mm specimen was obtained by Sarratt biopsy forceps (Stericom, Chesham, UK), and
16 samples were stored within 30 min in a light-protective cryovial at –80 °C until
17 analysis. Rectal biopsies were obtained by the same methodology described above
18 (following the insertion of a proctoscope) from a site in the rectum proximal to the
19 dentate line to avoid highly innervated tissue.

20 The timing of

21

1 3.2.7 ANALYTICAL METHODS

2

3 3.2.7.1 *Pharmacokinetics*

4

5 RPV concentrations in all matrices were quantified by validated high-pressure
6 liquid chromatography–mass spectrometry using a Thermo triple quadrupole TSQ
7 Ultra mass spectrometer (Thermo Electron Corporation, Hemel Hempstead, UK)
8 operating in the positive ionization mode (selected reaction monitoring). (Azijn et al.
9 2010; Else et al. 2014)

10 In brief, a stable isotope–labeled internal standard (¹³C-d4-Rilpivirine; 20 µl, 80
11 ng/ml) was added to plasma, CVF, and tissue (VT and RT) samples (100 µl per
12 sample), followed by extraction with protein precipitation (acetonitrile/water; 5:1
13 vol/vol) and was quantified using an RPV-spiked plasma calibration curve (0.5–400
14 ng/ml; 100 µl per calibrator level; in duplicate).

15 Due to its acidic (pH 4–5) and viscous nature, CVF was diluted 1:4 with a known
16 volume of phosphate-buffered saline (1 mmol/l; adjusted to pH 4.5 with
17 orthophosphoric acid) in order to create a homogeneous matrix and to improve
18 pipetting accuracy. A 100-µl aliquot was then transferred to glass tubes, and the
19 relevant dilution factor was recorded and these imputed values were entered into
20 the analytical software.

1 For tissue biopsies, the weight of tissue (in milligrams) was recorded before
2 extraction. Tissue biopsies were transferred to a MINILYS tissue homogenizer (Bertin
3 Technologies, Bordeaux, France) and Precellys– Keramik kit (Bertin Technologies)
4 containing 0.5-ml tubes prefilled with 14-mm ceramic beads, and made up to a
5 volume of 100 µl with blank plasma. Inter- and intra-assay precision and accuracy
6 for quality control (QC) samples at low (LQC), medium (MQC), and high (HQC)
7 concentrations in plasma were <15%. The percentage recovery (internal standard
8 normalized) of RPV from plasma (≥96%) was shown to be consistent, precise, and
9 reproducible. Furthermore, the percentage recovery of RPV from direct CVF
10 aspirates (~90%) and rectal tissue (RT) (~96%) using protein precipitation was
11 equivalent to that of plasma, thus demonstrating that drug-free plasma serves as a
12 suitable pseudo-matrix for quantification of RPV in these matrices.

13 RPV concentrations in RF were quantified using a plasma calibration curve spiked
14 (50 µl; in duplicate) onto Weck Cel or PVA polyvinyl alcohol-based spears and
15 extracted by liquid–liquid extraction (hexane/ ethyl acetate; 80:20 vol/vol). The
16 calibration curve was linear over the 0.025–20 ng/sample. Inter- and intra-assay
17 precision and accuracy for all QC concentrations were between 3 and 11%. The
18 percentage recovery (internal standard normalized) of RPV absorbed onto Weck Cel
19 or PVA-based spears, after liquid–liquid extraction, was ≥80%, and the effect of the
20 sample matrix was minimal (<5% interference) when evaluated by spiking and
21 postcolumn infusion experiments.

22 RPV concentrations in CVL were quantified using an RPV-spiked CVL calibration
23 curve (100 µl per calibrator level) and extracted by protein precipitation

1 (acetonitrile/water; 5:1 vol/vol). RPV-free CVL (for spiking purposes) was obtained at
2 baseline from the subjects undergoing CVL sampling and, subsequently, pooled. The
3 CVL calibration curve was linear over 0.05–20 ng/ml.

4 RPV concentrations in all matrices were expressed as nanograms/milliliter. Tissue
5 homogenate and rectal/vaginal fluid samples absorbed onto ophthalmic sponges
6 were quantified using a nanogram/sample calibration curve in order to account for
7 variations in tissue weight and fluid volumes.

8 RPV concentrations in tissue (expressed as nanograms/milliliter) were calculated
9 by converting x mg of tissue to a volume assuming a tissue density of 1.05 g/ml.

10 3.2.7.2 *Pharmacodynamics*

11 To determine the effect of RPV genital-tract concentrations on HIV replication in
12 vitro, CVL samples were collected at baseline and on days 28 and 56 from women
13 who received either 300 or 1,200 mg doses, during the third study phase. The
14 antiviral activity of CVL samples was assessed against HIV-1BaL challenge of TZM-bl
15 cells as previously described.^{7,10}

16 In brief, TZM-bl cells were plated at 3×10^4 /well and incubated overnight before
17 exposure to approximately 10^3 TCID₅₀ HIV-1BaL in the presence of undiluted CVL or
18 control buffer (normal saline containing 200 µg/ml bovine serum albumin) in
19 triplicate wells. At 48 h postinfection, the inoculum was removed by washing once
20 with 200 µl phosphate-buffered saline; cells were lysed in 100 µl luciferase cell
21 culture lysis reagent (Promega, Madison, WI), and cell lysates were stored at –80 °C

1 until they were assessed for luciferase activity using a luciferase assay buffer
2 (Promega).

3

4 **3.2.8 DATA ANALYSIS**

5

6 The calculated parameters for plasma and genital-tract RPV were maximum
7 observed concentration (C_{\max}), the area under the concentration–time curve from
8 day 0 to 84 (AUC_{84d}), and the concentration measured at 84 days after the observed
9 i.m. dose (C_{84d}).

10 All PK parameters were calculated using actual blood sampling times and non-
11 compartmental modeling techniques (WinNonlin Phoenix (version 6.1; Pharsight,
12 Mountain View, CA).

13 Dose proportionality was assessed by comparing dose-normalized (to 300 mg)
14 and log-transformed PK data (AUC and C_{\max}) using an analysis of variance and
15 pairwise comparisons. In addition, a regression analysis of individual data based on
16 the model

17 $y = \alpha \cdot \text{dose}^{\beta}$ was applied, where y is either AUC or C_{\max} , and β is the slope. A value
18 of 1 for β indicates perfect dose proportionality.

19 Descriptive statistics, including GMs and 90% CIs, were calculated for all
20 parameters.

1 Ratios of compartmental-to-systemic drug concentrations were calculated for
2 each PK parameter (C_{\max} , AUC_{84d}, C₈₄) and at all time points over the course of 84
3 days. The effects of gender, body weight, BMI, age, and ethnicity on systemic
4 (plasma) and compartmental (female genital-tract) PK were evaluated using
5 univariate and multivariate linear regression analyses. PK parameters were log
6 transformed and dose normalized (to a dose of 300 mg). Variables were included in
7 a full multivariate regression model if $P < 0.1$, and backward elimination ($P < 0.1$)
8 was used to identify the most important predictors. Colinearity diagnostics were
9 undertaken for expected interacting variables. All statistical analyses were
10 performed using SPSS (version 20.0; IBM, New York, NY).

11 For the PK/PD correlation (CVL RPV concentration vs. HIV inhibition *in vitro*), a
12 nonparametric Spearman's correlation was calculated using GraphPad Prism
13 software (version. 6; GraphPad Prism, La Jolla, CA).

14

1

2

3.3 RESULTS

3

4 3.3.1 PARTICIPANT DEMOGRAPHICS, DISPOSITION AND SAFETY

5

6 Of the 89 women screened for this study, 60 were enrolled; in three phases of
7 20, they received a single dose of RPV-LA and provided blood, fluid, and tissue
8 samples for analysis over the ensuing 84 days in each phase of the adaptive study
9 design (Table 3:D Study Phases and sample schedule according to adaptive design
10 modifications).

11

Table 3:D Study Phases and sample schedule according to adaptive design modifications

Study phase	Gender	Dose received (mg)	Number recruited	Pharmacokinetic			Viral Inhibition
				Plasma samples	Cervical fluid Rectal fluid (male)	Tissue: Cervical Rectal (Male)	Cervico-vaginal fluid lavage
				Sample schedule (study day)			
Phase 1	Male	600	6	Day 0 (pre-dose, 4 and 8 hour post dose) Days 1 to 84		7 and 14	—
	Female	300	10			7 and 14 or 14 and 28	—
		600	10				—
Phase 2	Female	600	10		Day 0 (8 hour post-dose) Days 1 to 84*	14 and 28	—
		1200	10				—
Phase 3	Female	300	10			28 and 56	0 (predose), 28 and 56
		1200	10				

* Days 1, 1, 3, 7, 11 (plasma only), 14, 21, 28, 42, 56 and 84

All female participants who received the single dose completed the study with no withdrawals due to adverse events. Six men were screened and found eligible to enrol, completing the study after receiving the 600 mg i.m. dose. Demographic information is presented in (Table 3:E Study Population Demographics and Dose Distribution); the three groups of women receiving 300, 600, and 1,200 mg doses had similar demographic characteristics.

1

Table 3:E Study Population Demographics and Dose Distribution

			Female		Male
		300 mg	600 mg	1200 mg	600 mg
Age (year)	Mean (± SD)	34 (±9)	35 (±8)	36 (±9)	36 (±15)
Height (m)	Median (range)	1·67 (1·51 – 1·79)	1·68 (1·58 – 1·76)	1·64 (1·52 – 1·76)	1·76 (1·69 – 1·81)
Weight (kg)		74·3 (51·2 – 96·7)	74·5 (55·6 – 100·4)	66·5 (47·8 – 90·6)	74·5 (61·0 – 100·4)
BMI (kg·m ⁻²)		26·6 (19·5 - 34·7)	26·6 (20·1 – 34·7)	24·7 (17·1 – 34·1)	24·5 (20·4 – 30·0)
Ethnicity					
Black		9 (45%)	14 (70%)	9 (45%)	1 (17%)
White		9 (45%)	5 (25%)	11 (55%)	5 (83%)
Asian		2 (10%)	1 (5%)	0	0

2

3 Participants tolerated the medication well. The majority of the adverse events
4 experienced were mild in severity, and of those defined by the investigator as
5 definitely or probably related to treatment, the most common were transient, self-
6 limiting discomfort at the injection site and temporary presence of a palpable non-
7 tender nodule. This was considered to be the deposit of the administered product;

1 no cases of this were complicated by any local signs of infection, and all cases
2 resolved completely over the course of study involvement.

3 In the third phase of the study, one female participant receiving the 300 mg dose
4 experienced a significant medical event of HIV infection after nonadherence to the
5 use of barrier contraception with a new male sexual partner (subsequently found to
6 be newly HIV seropositive) at approximately 6 weeks after receiving study
7 medication. This will be discussed further in section 1 below.

8 **3.3.2 RILPIVIRINE PLASMA PHARMACOKINETICS**

9 At the lowest dose of 300 mg, 19 of the 20 women had detectable plasma RPV
10 above the lower limit of quantification (LLQ) at 4 h postadministration.

11 In women, the geometric mean (GM) and 90% confidence intervals (CIs) for RPV
12 concentrations in blood plasma for all participants over 84 days after dosing are
13 depicted in Figure 3:B, Figure 3:D,

14 Figure 3:F , and pharmacokinetic parameters are presented in Table 3:F.

1 *Table 3:F RPV PK in female plasma and genital tract (CVF); (geometric mean, 90% confidence intervals)*

	Plasma			CVF			[RPV] _{CVF} /[RPV] _{PLASMA}		
Dose	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg
N	20	20	20	20	20	20	20	20	20
PK parameter	GM 90%CI								
C _{max} (ng/mL)	34	82	160	67	99	200	2.0	1.2	1.3
	28, 40	69, 95	137, 183	42, 93	67, 132	155, 245	1.6, 2.5	0.8, 1.6	1.0, 1.5
T _{max} (days)	7.9	6.0	6.2	5.3	7.2	8.5	0.7	1.2	1.4
	4.3, 11.5	3.4, 8.6	4.3, 8.1	2.5, 8.2	3.1, 11.3	5.1, 11.9	0.1, 1.5	0.3, 2.1	0.1, 2.6
t _½ (days)	43	39	38	34	31	43	0.8	0.8	1.2
	28, 58	33, 45	30, 47	23, 45	25, 37	31, 56	0.6, 1.0	0.7, 1.0	0.6, 1.8

2 C_{max} = maximum concentration, T_{max} = time to reach C_{max}, t_½ = half-life, C₂₈ = drug concentration 28 days post dose, C₅₆ = drug concentration

3 56 days post dose, C₈₄ = drug concentration 84 days post dose, AUC_{84d} = area under the curve from 0 to 84 days.

1

Table 3:C contd

	Plasma			CVF			[RPV] _{CVF} /[RPV] _{PLASMA}		
Dose	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg
C ₂₈	19	44	83	25	39	85	1.2	0.9	1.0
(ng/mL)	16, 23	34, 55	67, 99	14, 36	18, 61	64, 106	0.8, 1.6	0.3, 1.5	0.8, 1.3
C ₅₆	9	23	45	12	15	36	1.3	0.8	0.8
(ng/mL)	8, 11	19, 26	36, 55	7, 17	7, 22	26, 46	1.0, 1.7	0.5, 1.2	0.5, 1.1
C ₈₄	6	16	30	10	12	36	1.7	0.9	1.2
(ng/mL)	6, 7	13, 19	24, 37	6, 15	5, 20	26, 46	1.1, 2.2	0.6, 1.3	0.7, 1.7
AUC _{84d}	1231	2934	5982	2027	3207	6500	1.7	1.1	1.1
(ng/day/mL)	1054,1408	2569,3300	5156,6807	1409,2645	2262,4152	5264,7735	1.3, 2.0	0.8, 1.4	0.9, 1.3

2 C_{max} = maximum concentration, T_{max} = time to reach C_{max}, t_½ = half-life, C₂₈ = drug concentration 28 days post dose, C₅₆ = drug concentration

3 56 days post dose, C₈₄ = drug concentration 84 days post dose, AUC_{84d} = area under the curve from 0 to 84 days.

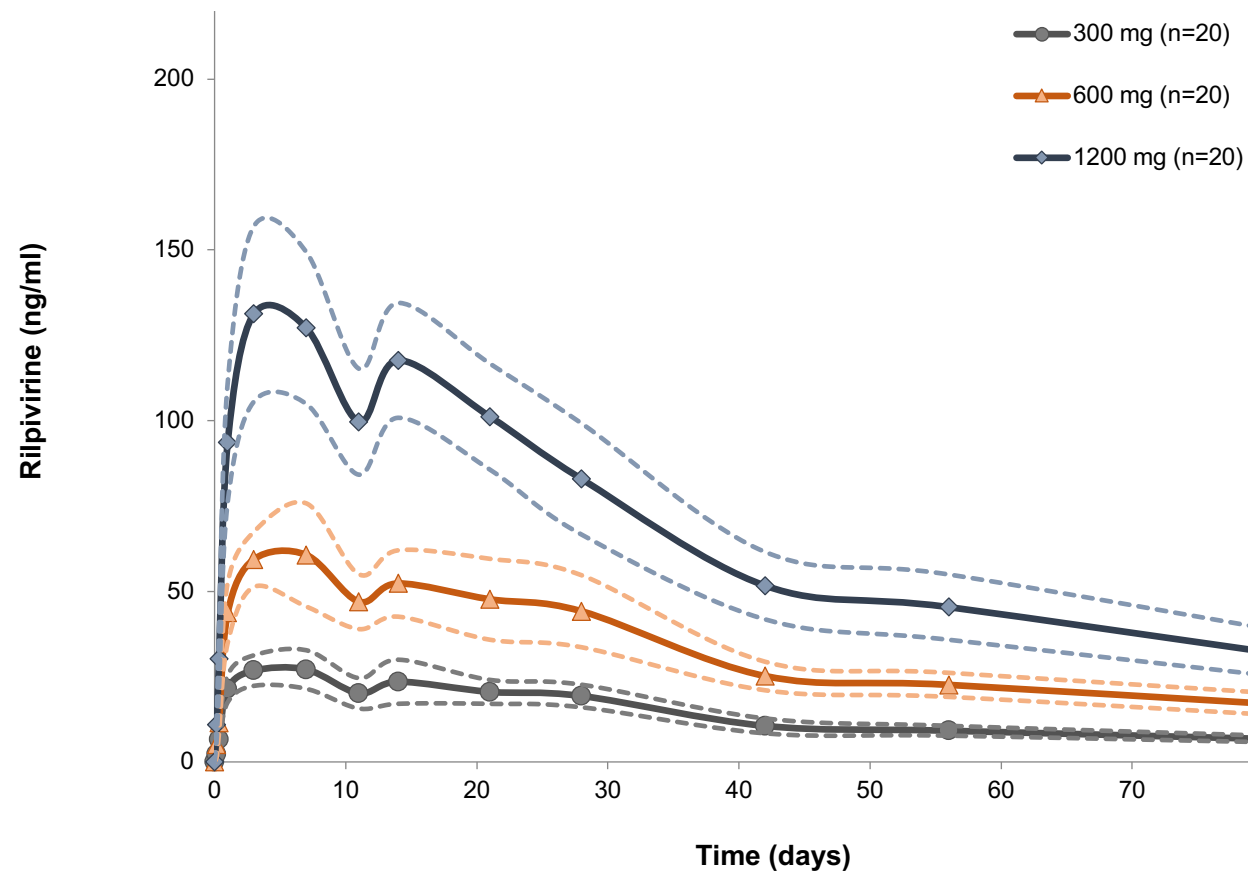


Figure 3:B Rilpivirine concentrations in women in plasma over 84 days post dose, when receiving LA-RPV i/m at 300, 600 or 1,200mg.

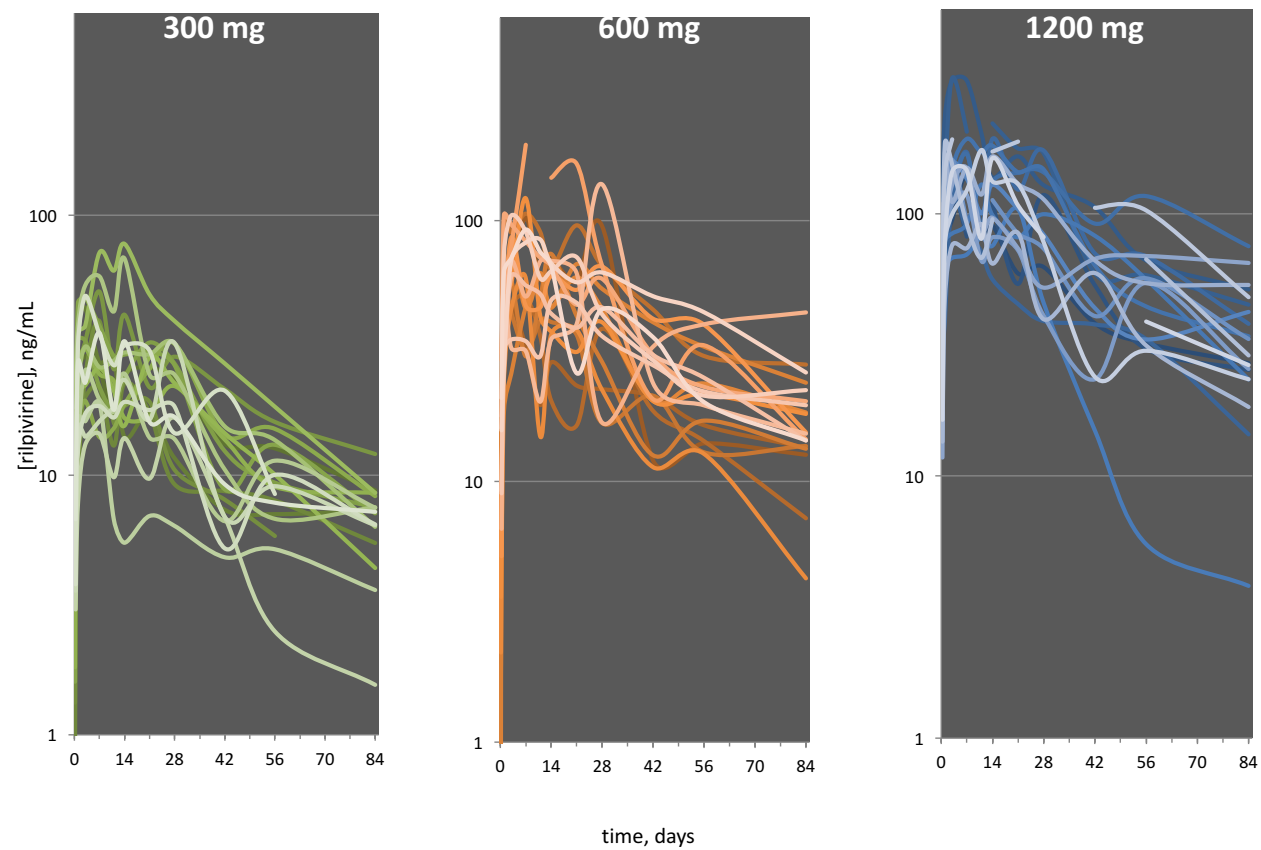
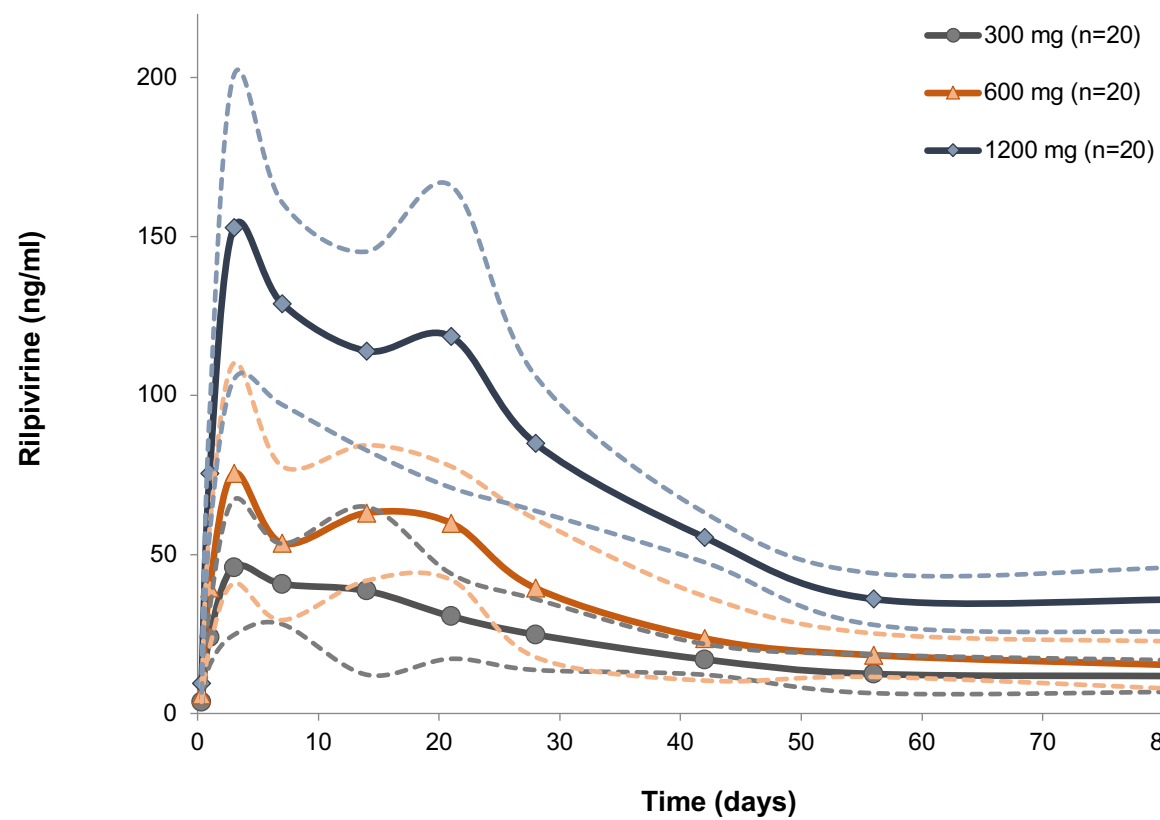


Figure 3:C Individual PLASMA rilpivirine concentration/time curves in female participants after receiving a single a) 300mg, b) 600mg or c) 1200mg dose.



1

2 Figure 3:D Rilpivirine concentrations in women in cervico-vaginal fluid over 84 days post dose, when receiving LA-RPV i/m at 300, 600 or

3 1,200mg.

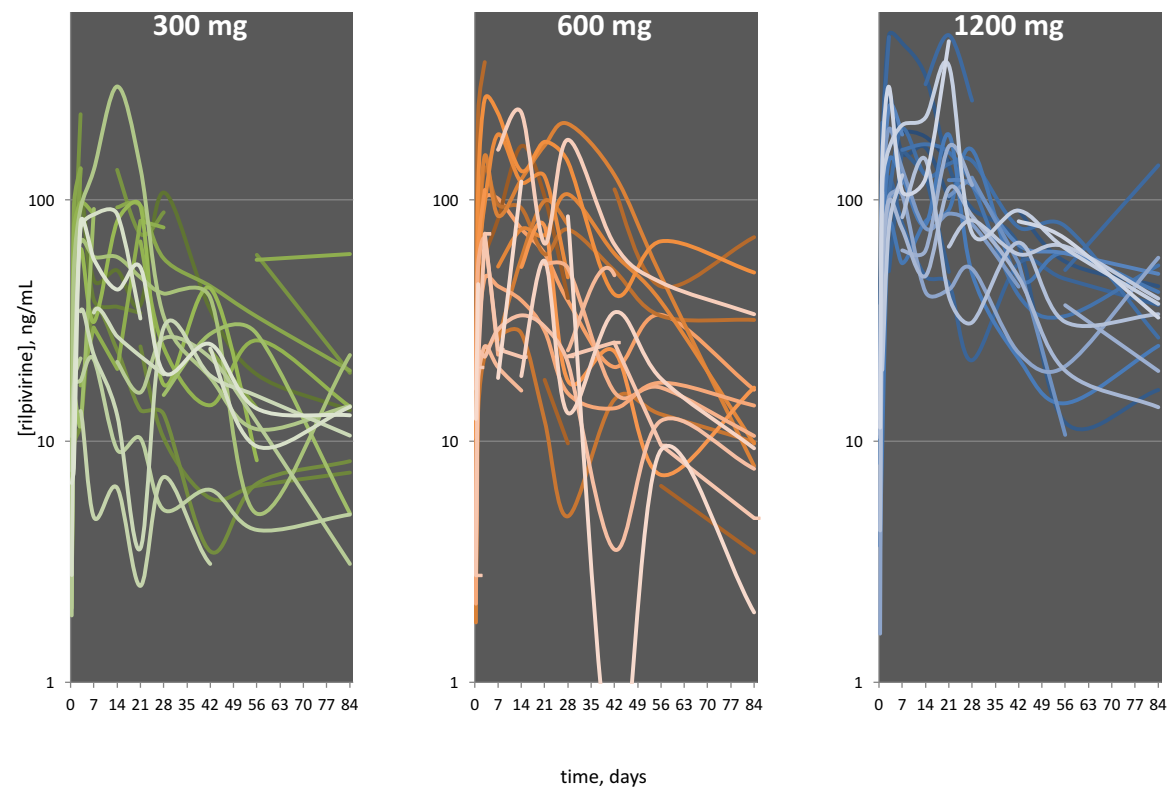
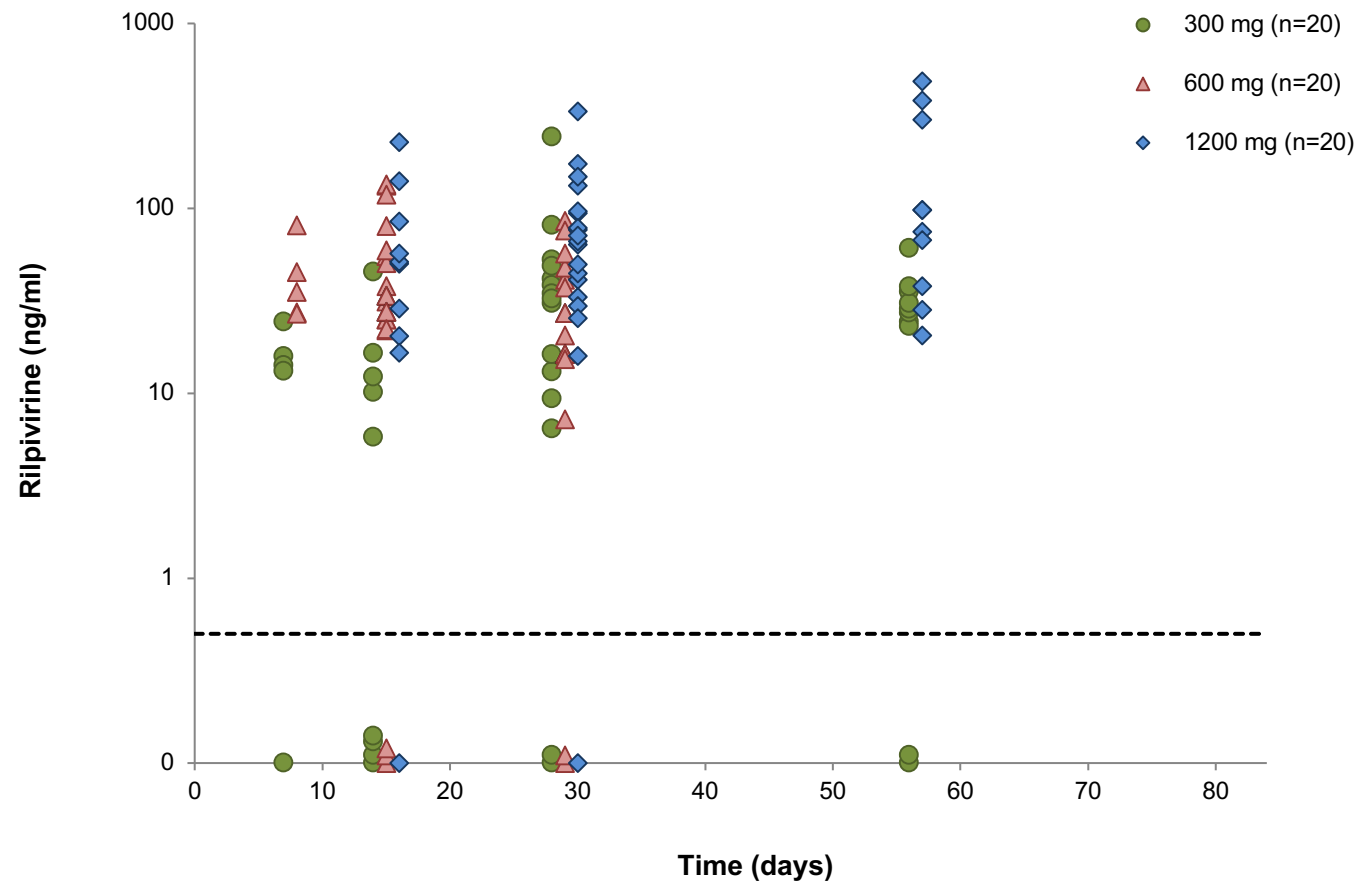


Figure 3:E Individual CERVICOVAGINAL rilpivirine concentration/time curves in participants after receiving a) 300mg, b) 600mg or c) 1200mg dose.



1

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Figure 3:F Rilpivirine concentrations in women in vaginal tissue over 84 days post dose, when receiving LA-RPV i/m at 300, 600 or 1,200mg.

1 Mean peak concentration (C_{\max}) in plasma of 33.7, 81.9, and 160.2 ng/ml was
2 attained at 7.9, 6.0, and 6.2 days (mean time to C_{\max} (T_{\max})) after a single dose of
3 300, 600, and 1,200 mg, respectively. After this peak, the concentration–time curves
4 describe a prolonged persistence of drug in plasma, with concentrations at 28 days
5 (C_{28}) of 19.3, 44.2, and 82.9 ng/ml; at 56 days (C_{56}) of 9.1, 22.6, and 45.4 ng/ml; and
6 at 84 days (C_{84}) of 6.4, 16.2, and 30.2 ng/ml after the 300, 600, and 1,200 mg dose,
7 respectively. Pairwise comparisons of plasma log area under the concentration–time
8 curve (AUC) and C_{\max} (dose normalized) did not result in any significant differences
9 in plasma exposures among the three dosing groups ($P \geq 0.1$). In addition, the 90%
10 CI of the regression slopes included unity. Dose proportionality was also
11 demonstrated for rilpivirine CVF AUC and C_{\max} (with the exception of C_{\max}
12 comparisons between the 1,200 and 300 mg doses; $P < 0.1$) using the same
13 approach. These data suggest that both systemic and compartmental RPV
14 concentrations are proportional to the administered i.m. dose.

15 In comparison, the six men receiving the 600 mg dose exhibited approximately
16 39% higher mean C_{\max} as compared with women at the same dose (114 vs. 81.9
17 ng/ml), with subsequent higher concentrations at the sampling points C_{28} (33%
18 higher) and C_{56} (21% higher) but with equivalent concentrations by day 84. This
19 resulted in an increased overall exposure (AUC_{84d}) that was 32% above that
20 measured in women at the same dose (Table 3:G RPV PK in male plasma and rectal
21 compartment (RF); (geometric mean, 90% confidence interval).

1

2

3

*Table 3:G RPV PK in male plasma and rectal compartment (RF);
(geometric mean, 90% confidence interval)*

	Plasma	RF	$[\text{RPV}]_{\text{RF}}/[\text{RPV}]_{\text{PLASMA}}$
Dose	600 mg	600 mg	600 mg
N	6	6	6
PK parameter	GM 90%CI		
C_{max} (ng/mL)	114.1 88.8, 139.4	35.7 17.7, 53.6	0.31 0.20, 0.43
T_{max} (days)	5.0 0, 11.7	6.2 0, 13.6	1.24 0, 3.94
$t_{1/2}$ (days)	30.5 26.1, 35.0	17.8 9.3, 26.3	0.61 0.24, 0.97
C_{28} (ng/mL)	58.9 42.3, 75.5	11.9 0, 35.0	0.20 0, 0.49
C_{56} (ng/mL)	27.4 20.9, 33.8	5.9 3.4, 8.5	0.22 0.03, 0.41
C_{84} (ng/mL)	15.9 13.9, 17.9	1.6 0, 3.4	0.10 0, 0.22
$\text{AUC}_{84\text{d}}$ (ng/day/mL)	3873.4 3283.7, 4463.2	935.0 350.2, 1519.8	0.24 0.10, 0.38

4

C_{max} = maximum concentration, T_{max} = time to reach C_{max} , $t_{1/2}$ = half-life, C_{28} = drug

5

concentration 28 days post dose, C_{56} = drug concentration 56 days post dose, C_{84} =

6

drug concentration 84 days post dose, $\text{AUC}_{84\text{d}}$ = area under the curve from 0 to 84

7

days.

8

1

2 **3.3.3 CERVICO-VAGINAL FLUID PHARMACOKINETICS**

3 RPV was detectable above the assay LLQ in the first CVF sample, 8-h postdose, in
4 all female participants in whom sample collection was successful (57/60; 95%), the
5 exceptions being three volunteers receiving the 300 mg dose.

6 RPV CVF mean T_{max} mirrored that in plasma of between 5 and 8 days, attaining
7 higher mean peak concentrations of 67.4, 99.3, and 199.9 ng/ml at 300, 600, and
8 1,200 mg, respectively (Table 3:F, Figure 3:D). Thereafter, RPV concentrations in CVF
9 approximated those seen in plasma, with the GM $[RPV]_{CVF}/[RPV]_{PLASMA}$ ratio in paired
10 samples maintained persistently at or above 0.8 at each dose level (Table 3:F). On
11 day 84, RPV concentrations in CVF were still measurable, with a GM of 11.7, 14.9,
12 and 36.0 ng/ml, respectively. Dose proportionality was also apparent in CVF,
13 although inter-subject variation was higher than in the plasma compartment.

14 **3.3.4 RECTAL FLUID AND TISSUE CONCENTRATIONS**

15 Rilpivirine was detectable in the first RF sample taken at 8 h postdose, in five of
16 the six male participants, and in all six on day 1 (24 h postdose). The highest
17 concentration in RF was observed at 6.2 days postdose and ranged from 15 to 92
18 ng/ml (Table 3:G, Figure 3:H), with the remaining measurable up to day 84 (from 0.4
19 to 7.4 ng/ml). RPV concentrations in RF were substantially lower than those in
20 plasma or CVF. GM ratios of concentrations between RF and plasma ranged
21 between 0.09 and 0.33 at different time points over the course of 84 days, with a
22 ratio for the overall exposure, AUC_{0-84d} RF/plasma of 0.24.

- 1 Rectal tissue RPV concentrations in men ranged from 67 to 128 ng/ml of tissue
- 2 on day 7 and from 33 to 156 ng/ml of tissue on day 14 (Table 3:H, Figure 3:I), with
- 3 tissue/plasma ratios from 0.7 to 1.2 on day 7, and from 0.5 to 1.3 on day 14.

1

Table 3:H RPV concentrations in VT and RT and their relation to plasma concentrations; (geometric mean, 90% confidence intervals)

Females		Plasma (n=20)			VT (n)			[RPV] _{VT} /[RPV] _{PLASMA}		
Dose	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg	300 mg	600 mg	1200 mg	
PK parameter	GM 90%CI									
C ₇ ng/mL	27.1	60.6	127.2	16.4	39.4	—	0.65	0.72	—	
	21.4, 32.7	45.5, 75.8	104.9, 149.4	14.5, 18.2 1<LLQ/(5)	31.2, 47.6 (5)		0.51, 0.80	0.40, 1.05		
C ₁₄ ng/mL	23.4	52.3	117.7	13.9	41.4	53.9	0.59	0.78	0.45	
	17.0, 29.9	42.6, 62.0	100.8, 134.4	8.1, 19.7 4 <LLQ/(9)	29.1, 53.8 3<LLQ/(20)	28.6, 79.4 1<LLQ/(10)	0.30, 0.87	0.55, 1.01	0.22, 0.67	
C ₂₈ ng/mL	19.3	44.2	82.9	31.8	33.8	66.6	1.70	0.80	0.83	
	16.0, 22.6	33.6, 54.7	66.6, 99.2	9.1, 54.5 2<LLQ/(15)	20.3, 47.3 3<LLQ/(15)	38.8, 94.4 1<LLQ (19)	0.58, 2.82	0.41, 1.19	0.48, 1.18	
C ₅₆ ng/mL	9.1	22.6	45.4	31.9	—	94.9	3.79	—	2.31	
	7.7, 10.6	19.1, 26.1	35.8, 54.9	27.4, 36.4 2<LLQ/(10)		33.3, 156.6 (10)	2.35, 5.23		0.29, 4.90	
Males	Plasma			RT			[RPV] _{RT} /[RPV] _{PLASMA}			
Dose	600 mg			600 mg			600 mg			
PK parameter	GM 90%CI (n)									
C ₇ ng/mL	95.5			93.7			0.98			
	79.8, 111.1 (6)			75.6, 111.8 (6)			0.85, 1.11 (6)			
C ₁₄ ng/mL	78.2			70.3			0.90			
	63.7, 92.6 (6)			34.2, 106.3 (6)			0.59, 1.20 (6)			

2

3

C₇, drug concentration 7 days postdose; C₁₄, drug concentration 14 days postdose; C₂₈, drug concentration 28 days postdose; C₅₆, drug concentration 56 days postdose; LLQ, lower limit of quantification; RT, rectal tissue, VT, vaginal tissue.

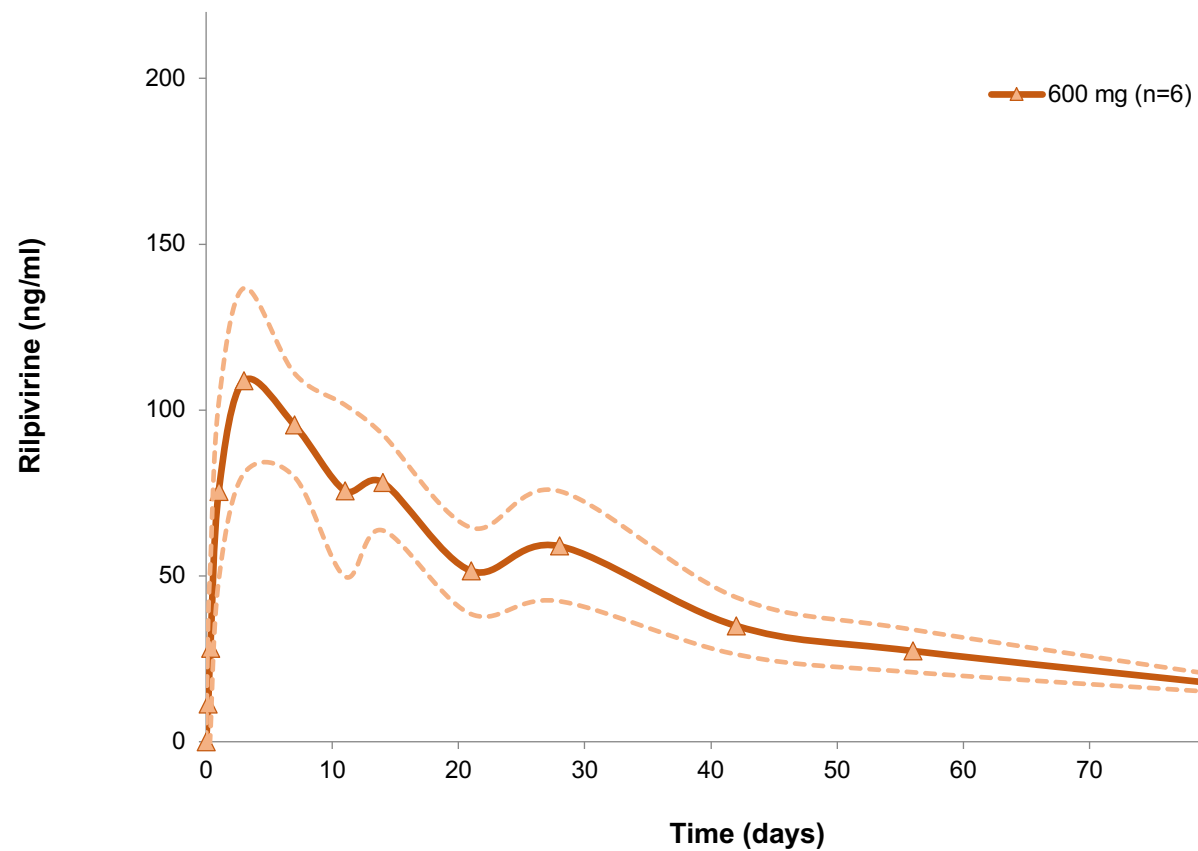


Figure 3:G Rilpivirine concentrations in men in plasma over 84 days postdose, when receiving long-acting RPV intramuscularly at 600mg

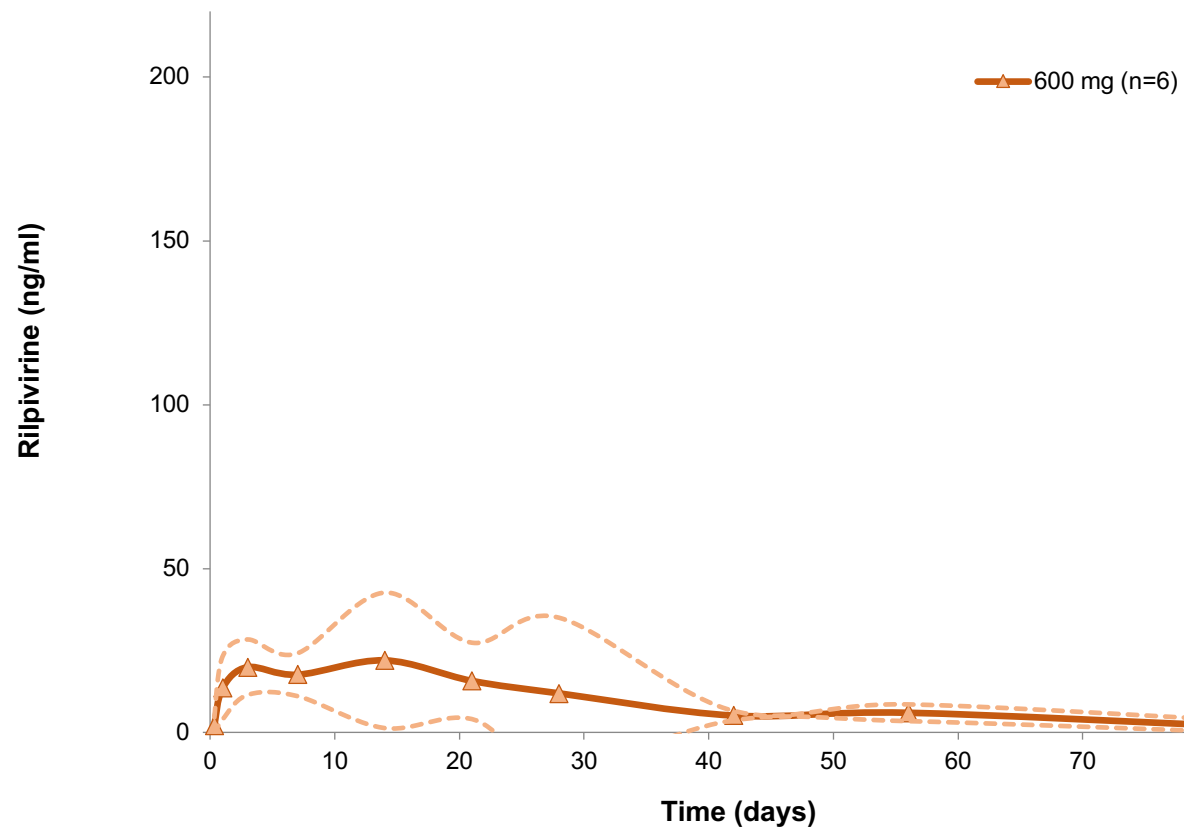


Figure 3:H Rilpivirine concentrations in men in rectal fluid over 84 days postdose, when receiving long-acting RPV intramuscularly at 600mg

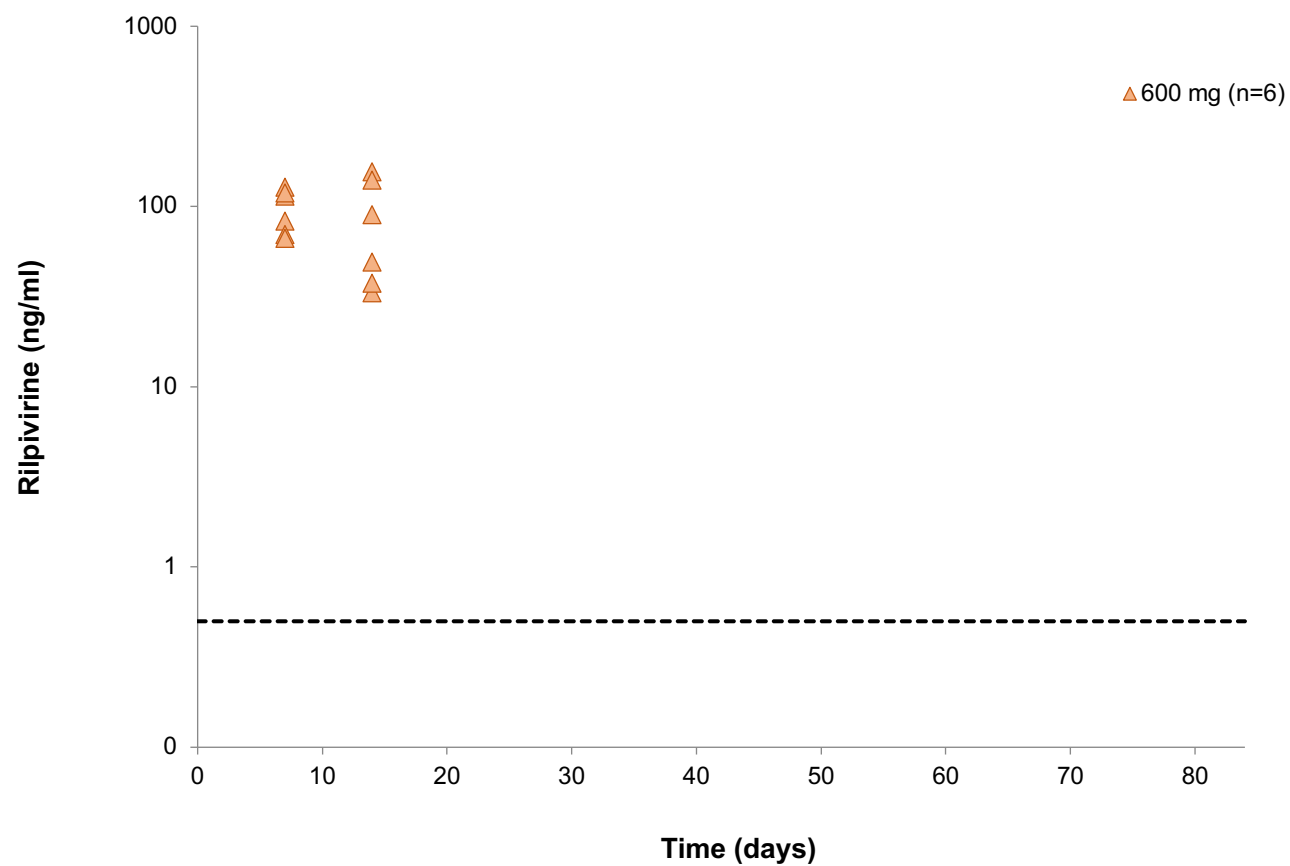


Figure 3:I Rilpivirine concentrations in men in rectal tissue over 84 days postdose, when receiving long-acting RPV intramuscularly at 600mg.

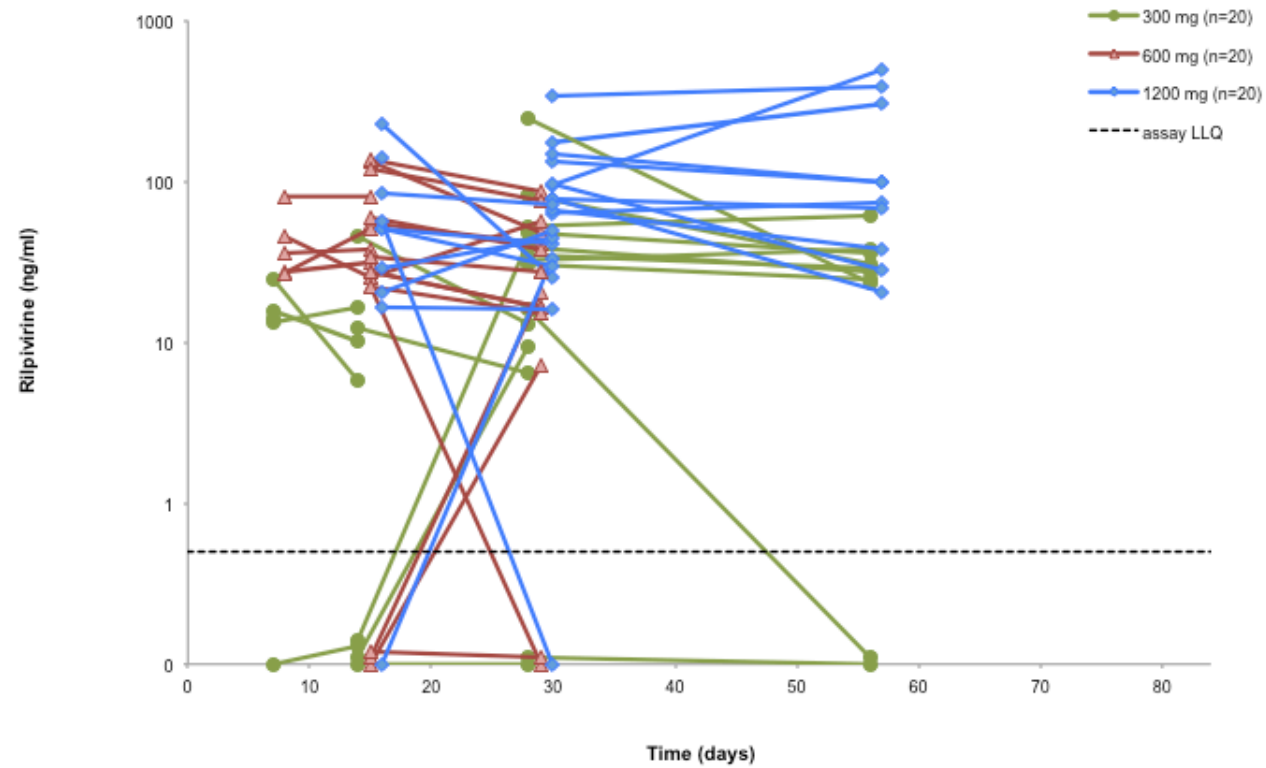


Figure 3:J Paired rilpivirine tissue concentrations from individual participants over 84 days postdose after receiving RPV intramuscularly at 300mg, 600mg and 1,200mg.

1 **3.3.5 VAGINAL TISSUE**

2 Individual vaginal tissue (VT) RPV concentrations (GM, 90% CI) on days 7, 14, 28,
3 and 56 are presented in

4 Figure 3:F and Figure 3:J, and concentrations with associated
5 [RPV]_{VT}/[RPV]_{PLASMA} ratios are shown in Table 3:H.

6 In women in whom sampling was successful, RPV VT (ng/ml) concentrations (GM
7 90% CI presented in Table 3:H, in women receiving 300 mg ranged from 13 to 24 on
8 day 7 (1 < LLQ), from 6 to 45 on day 14 (4 < LLQ), from 6 to 243 on day 28, (2 < LLQ),
9 and from 23 to 61 ng/ml on day 56; with the GM of ratios in concentration—
10 [RPV]_{VT}/[RPV]_{PLASMA}—on these days being 0.7, 0.6, 1.7, and 3.8, respectively.

11 In those receiving the 600 mg dose, concentrations ranged from 27 to 81, from
12 22 to 135 (3 < LLQ), and from 7 to 132 (3 < LLQ) in women who were on 600 mg on
13 days 7 (n = 5), 14 (n = 20), and 28 (n = 15). [RPV]_{VT}/[RPV]_{PLASMA} GM ratios were 0.7,
14 0.8, and 0.8 on days 7, 14, and 28, respectively.

15 At the highest dose (1,200 mg), concentrations ranged from 17 to 228 (1 < LLQ),
16 from 16 to 335 (1 < LLQ), and from 21 to 487 in women on days 14 (n = 10), 28 (n =
17 19), and 56 (n = 10). [RPV]_{VT}/[RPV]_{PLASMA} GM ratios were 0.5, 0.8, and 2.3 on days 14,
18 28, and 56, respectively.

19 **3.3.6 COVARIATE ANALYSIS**

20 A total of 520 paired plasma and CVF samples were available from the 60 female
21 participants in the study. Plasma and CVF concentrations were significantly

1 correlated ($r^2 = 0.518$; $P < 0.01$). Furthermore, the association remained significant
2 when stratifying by the dose administered (300 mg: $r^2 = 0.559$; $P < 0.01$; $n = 173$
3 pairs; 600 mg: $r^2 = 0.363$; $P < 0.01$; $n = 173$ pairs; 1,200 mg: $r^2 = 0.449$; $P < 0.01$; $n =$
4 174 pairs). Plasma and VT RPV concentrations from 100 paired samples were also
5 significantly correlated ($r^2 = 0.139$; $P < 0.01$; $n = 100$), but the association was
6 somewhat weaker, and the significance was lost when stratifying by dose.

7 When investigating the effects of predictors on RPV plasma concentrations, in a
8 multivariate model, female gender and body mass index (BMI) were found to be
9 independently associated with RPV C_{max} . Female gender was associated with an
10 approximately 30% decrease in RPV C_{max} ($P = 0.013$), and a one-unit (kg/m^2)
11 increase in BMI was associated with a 2.3% decrease in C_{max} ($P = 0.028$). In terms of
12 the overall RPV exposure (AUC_{84d}) in plasma, only gender was a significant univariate
13 predictor, in which women were associated with a ~28% decrease in RPV AUC_{84d} .
14 Interestingly, there was no effect of either gender or BMI on RPV plasma
15 concentrations beyond 28 days postdose. No effect of age, bodyweight, or ethnicity
16 was observed for any of the RPV PK parameters in the plasma compartment.

17 In terms of predictors of RPV concentrations in the female genital tract (CVF and
18 VT), in a multivariate analysis, age < 40 years ($P < 0.031$), BMI $> 25 \text{ kg}/\text{m}^2$ ($P = 0.005$),
19 and RPV plasma concentrations ($P = 0.005$) were significant predictors of RPV
20 concentrations in the female genital tract (CVF and VT), whereby age < 40 years and
21 BMI $> 25 \text{ kg}/\text{m}^2$ were associated with a 35–40% reduction in RPV AUC and C_{max} in
22 CVF, after adjusting for RPV plasma concentrations. There was no evidence of any
23 co-linearity between age and BMI, and the effects of age and BMI were lost beyond

1 28 days postdose. In a univariate analysis, RPV CVF concentrations and BMI (>25
2 kg/m²) were predictors of RPV concentrations in VT at day 14. However, in a
3 multivariate model, only BMI was a significant predictor, with an ~57% reduction in
4 RPV tissue concentration ($P < 0.001$). No associations between BMI (or indeed other
5 covariates) and RPV VT concentrations at days 28 or 56 were observed.

6

1 3.3.7 VIRAL INHIBITION IN CERVICOVAGINAL LAVAGE

2 RPV concentrations measured in cervicovaginal lavage (CVL) (GM; 90% CI) at days
3 28 and 56, respectively, were 0.45 ng/ml (0.08-0.83; 2 < LLQ) and 0.28 ng/ml (0.11-
4 0.45; 4 < LLQ) for women who received the 300 mg dose, and 1.90 ng/ml (0.66-
5 3.13) and 0.63 ng/ml (0-1.79) for those who were given the 1,200 mg dose.

6 However, it should be recognised that the procedure for CVL collection resulted in a
7 considerable dilution of the naturally occurring fluid. Despite this, RPV

8 concentrations in CVL and paired “undiluted” tear-test strips, both of which were
9 taken on days 28 and 56 in phase III participants, were significantly correlated ($P <$

10 0.00001; Spearman’s $r = 0.7$). CVL collected from women receiving a single dose of

11 RPV-LA 1,200 mg i.m. during the third study phase ($n = 9$) had significantly greater

12 antiviral activity on both days 28 (93 % 12%; mean % SD) and 56 (78 % 23%), as

13 compared with the baseline activity (28 % 64%) (Figure 3:K **plot A**). By contrast, CVL

14 obtained from women who were on the lower i.m. dose of 300 mg did not result in

15 a significant increase in antiviral activity at either time point, as compared with

16 baseline (Figure 3:K **plot B**) and Figure 3:L). The activity correlated significantly ($P <$

17 0.0001; Spearman’s $r = 0.8$) with RPV concentrations (Figure 3:K, **plot C**). Because

18 the endogenous antiviral activity of CVL is highly variable,¹⁰⁻¹² the data were also

19 analysed by subtracting the baseline inhibition from that observed at 28 and 56 days

20 postdose, for each subject. The RPV-LA-driven inhibition after the 1,200 mg i.m.

21 dose was higher at day 28 (71 % 67%; mean % SD) and persisted until day 56 (58 %

22 74%; mean % SD). By contrast, there was little to no increment after the 300 mg

23 dose (Figure 3:M).

1 The volunteer who tested positive for HIV antibodies on study day 84 had
2 detectable plasma HIV RNA of 370 and 175,060 copies/ml on study days 56 and 84,
3 respectively. This volunteer received the lowest studied dose (300 mg i.m.) and the
4 plasma and CVF RPV concentrations were 24.3 and 32.9 ng/ml, respectively, on day
5 28, 10.5 and 18.3 ng/ml, respectively, on day 42 (when presumed exposure to HIV
6 occurred), 6.8 and 11.2 ng/ ml, respectively, on day 56, and 7.5 and 14.0 ng/ml,
7 respectively, on day 84. There was minimal change in viral inhibition in CVL obtained
8 from this subject on day 28 (66%) or on day 56 (55%) from that at baseline (49%).
9 These findings indicate that the 300 mg dose is not sufficient to protect against HIV
10 infection.

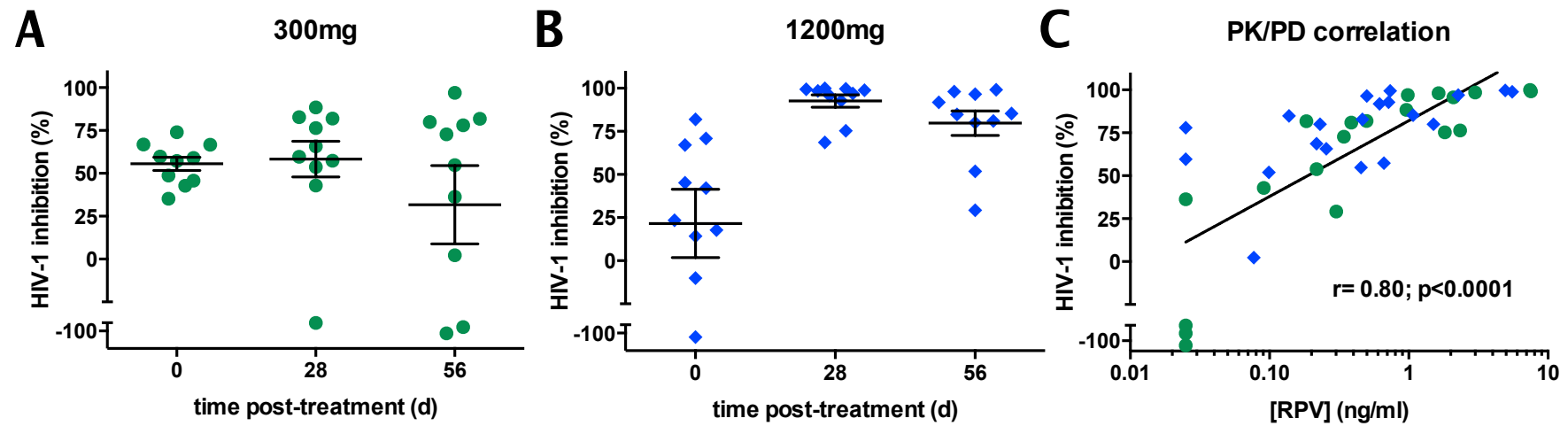
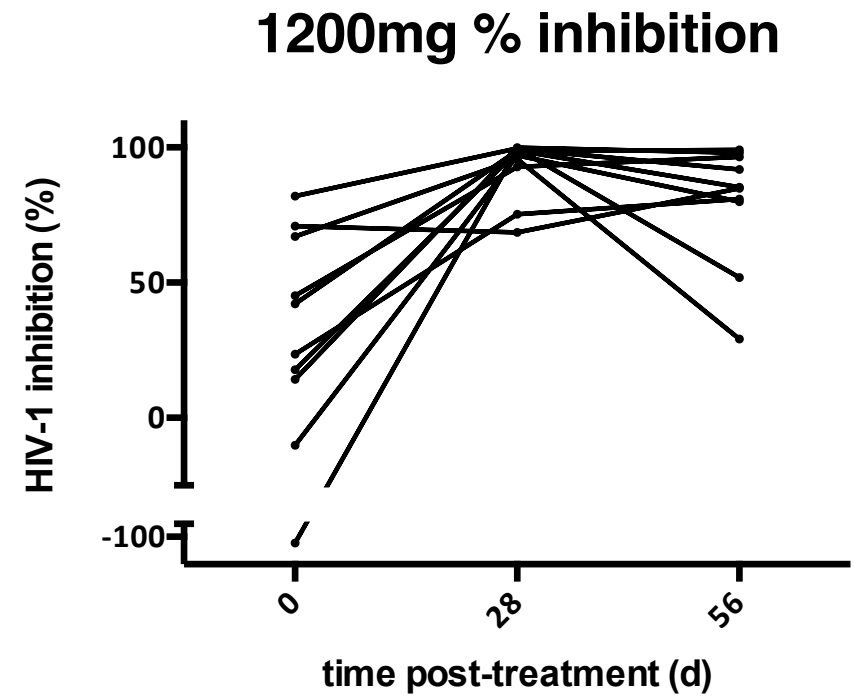
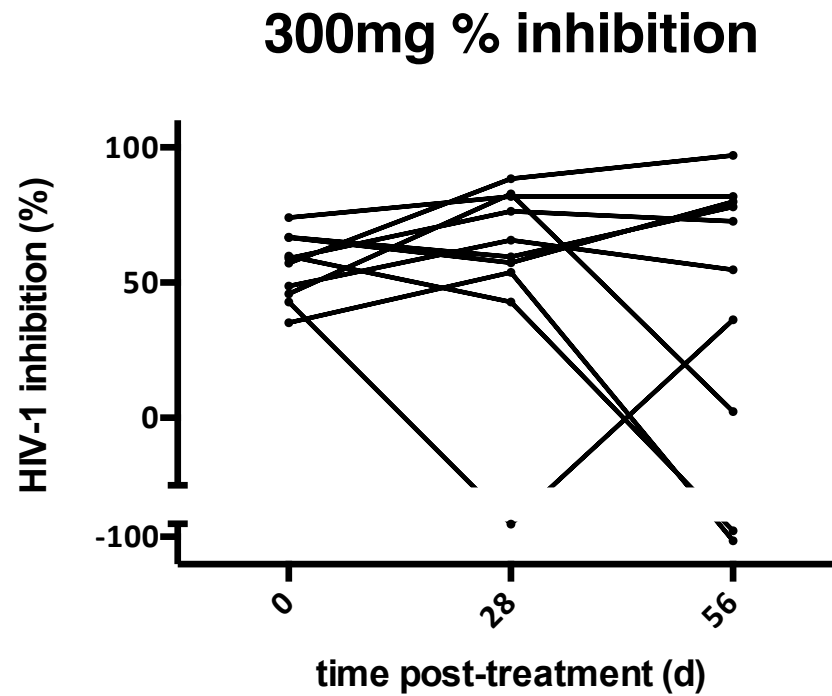


Figure 3:K HIV viral inhibition percentage measured ex vivo in cervicovaginal lavage in female participants at baseline and at 28 and 56 days postdose.

A 300mg, n = 10; **B** 1,200mg, n = 10; **C** Nonparametric Spearman's correlation between rilpivirine concentrations and viral inhibition at days 28 and 56.

1



2

3 *Figure 3:L HIV viral inhibition percentage measured ex vivo in cervicovaginal lavage in female participants at baseline and at 28 and 56*
4 *days postdose. (plotted lines from individual participants)*

Increment in HIV inhibition relative to baseline

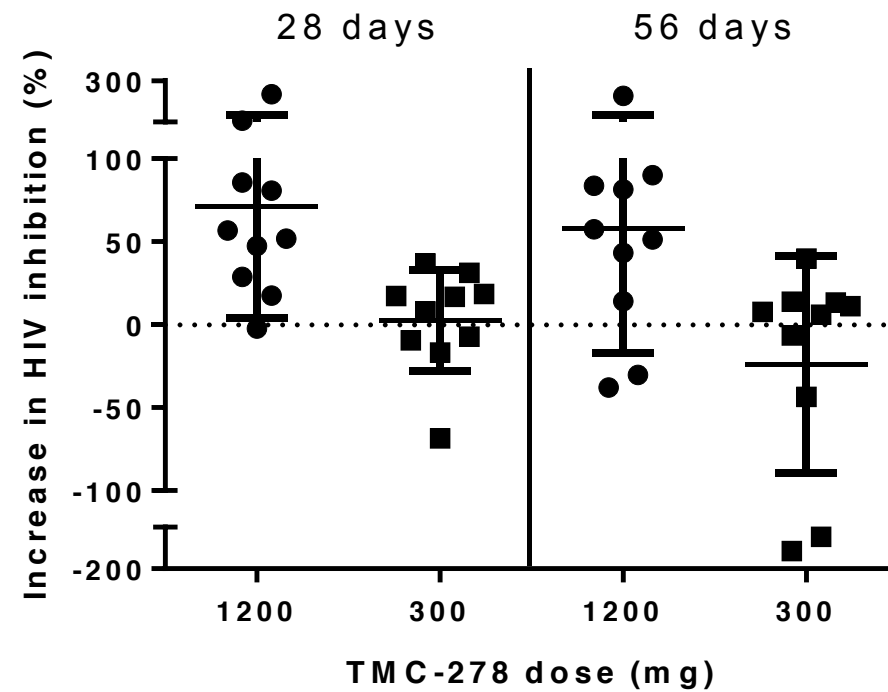


Figure 3:M Incremental change from baseline in HIV viral inhibition measured ex vivo in cervicovaginal lavage in female participants at baseline and at 28 and 56 days postdose.

3.4 DISCUSSION

This study was conducted to determine the concentrations of RPV in plasma, CVF, and VT in women after a single i.m. RPV-LA dose (300, 600, or 1,200 mg) and in the plasma, rectal compartment, and RT in men after 600 mg of RPV-LA i.m., over 84 days postdose.

Furthermore, the exploratory objective of this study was to investigate the effect of female genital-tract fluid (CVL) drug concentrations on HIV replication ex vivo, providing information on differential inhibitory effects of achieved RPV concentrations in biological fluids. The rationale for the study, study design, and dose selection and sampling intervals was based on the need to investigate the potential role of RPV-LA as an HIV PrEP agent and on the results from a previously conducted healthy volunteer study by the drug manufacturer (unpublished data, Janssen).

The study demonstrated that, after the administration of RPV-LA i.m. to both female and male healthy volunteers, measurable RPV plasma concentrations are achieved promptly (within 4 h post-administration) and persist for more than 84 days, in particular at higher doses. A secondary peak in the GM plasma concentration/time curves of the groups, observed at day 11, may reflect changes in the release of the drug from the injection site as well as complex RPV distribution kinetics (including immune cell trafficking of endocytosed nanoparticles from injection site to regional lymph nodes). It has been shown for long-acting preparations that the release profile can be influenced by changes in particle size

1 distribution as more dissolution occurs over time. However, multiple peaks were
2 observed in the individual curves (as shown in

3 Figure 3:C); thus, the secondary peak appearing in the GM profiles may simply be
4 due to increased relative sampling frequency during the first 14 days, as compared
5 with thereafter.

6 RPV concentrations in the female genital tract (CVF and VT) are also achieved
7 quickly and approximate those measured in plasma. Vaginal tissue concentrations
8 were slightly lower than genital-tract fluid, possibly reflecting cell-to-fluid flux in
9 response to concentration gradients.(Sugano et al. 2010; Van Damme et al. 2012;
10 Mesquita et al. 2013) RPV is highly (~99%) bound to plasma proteins, primarily to
11 albumin. Therefore, high accumulation of RPV in CV fluid cannot be explained solely
12 by protein binding. Instead, physicochemical properties, including the molecule's
13 dissociation constant, partition constant, and low molecular weight, may favour
14 active transport and low clearance from the CV compartment. One explanation is
15 that the more acidic environment of CVF (pH 4–5) as compared with either plasma
16 (pH 7.4) or RF (neutral to slightly basic) means that a greater percentage of RPV
17 exists in a protonated form (pK_a (logarithmic acid dissociation constant)= 5.16),
18 which may, in turn, result in mucosal “ion trapping” of the drug.

19 The RPV concentrations measured in the RT were equal to or higher than those in
20 plasma, although RPV exposure in RF was approximately 75% lower than that in the
21 plasma compartment. RF transudate may be diluted by luminal fluid derived from
22 the proximal gut, potentially indicating that this sample may not be a representative

1 surrogate for RT concentrations with systemically delivered drug. Altogether, these
2 data suggest a potential role of RPV-LA as a PrEP agent because of the exposures
3 measured at the sites of HIV transmission.

4 Notably, target protective RPV concentrations in plasma or tissues are unknown
5 and difficult to determine because of the lack of validated surrogate “efficacy”
6 markers.

7 Therefore, potential efficacy markers are inferred from (i) animal models, (ii)
8 human in vitro PD experiments, or (iii) phase III clinical trials of combination ARV
9 treatment, including oral RPV. In the treatment trials, the mean RPV C_{trough} was 80
10 ng/ml, and the upper limit of the lowest quartile of exposures, in which group the
11 virologic response was the lowest, was 50 ng/ml (unpublished data). However, a
12 minimum effective RPV concentration in vivo for prevention has not been defined to
13 date. It should be noted that the protein-corrected IC₉₀ value (90% inhibitory
14 concentration) for treatment is 12.1 ng/ml.(Azijn et al. 2010; van 't Klooster et al.
15 2010))

16 In the third stage of the study, CVL samples were collected to determine whether
17 RPV present in the vaginal lumen would inhibit HIV infection of susceptible cells *ex*
18 *vivo*. This well-established assay was previously used to assess the antiviral activity
19 of CVL samples collected from women exposed to a tenofovir gel
20 formulation.^{2,4,6,10} A limitation in quantifying drug in CVL is the unknown volume
21 of vaginal fluid suspended in the saline wash; thus, it is not possible to derive a
22 suitable correction factor for either drug or protein concentration. Nevertheless,

1 experimental analysis correlating concentrations achieved with such inhibitory
2 effects suggests that a single dose of 1,200 mg should deliver sufficient drug to the
3 genital tract to provide protection against sexual exposure to HIV; however, more
4 data, including direct challenge of cervical biopsies as another marker of
5 pharmacodynamic effect, are needed to confirm this.

6 We have also shown that BMI and gender may influence the absorption (C_{max})
7 of parenteral RPV-LA. In particular, women with a BMI of >25 kg/m² may be
8 susceptible to having lower RPV concentrations around the peak in the first 28 days
9 after dosing, both in the systemic circulation and at the site of HIV transmission. For
10 lipophilic agents such as RPV, the volume of distribution may be increased in
11 subjects with high BMI, resulting in lower drug concentrations in both the central
12 and peripheral compartments. However, these early findings are limited by an
13 under-representation of men in the study cohort, and because BMI does not
14 discriminate adipose tissue mass from muscular tissue, and men with high muscle
15 mass are not well described.

16 Indeed, differences between men and women may be related to a combination
17 of factors, including different rates of release of drug from the i.m. depot and the
18 higher body fat percentage and smaller water content in women, apart from
19 regional differences in adipose tissue distribution.

20 A further consideration is that in the female genital tract, changes in pH, stage of
21 the menstrual cycle and level of mucus production, and permeability of the vaginal
22 epithelium may also impact RPV-LA distribution in CVF and vaginal tissue, explaining

1 the high intra-individual variability observed in RPV concentrations in this
2 compartment.

3 On the basis of these data, a phase I multiple-dosing study of RPV-LA for HIV
4 prevention is currently under way investigating steady-state PK with repeated
5 administration of 600 and 1,200 mg doses using viral inhibition in tissue explants
6 (both genital and rectal) as a surrogate marker for efficacy. The consideration of
7 these aggregated data, analysed by a population PK/PD model approach, will
8 determine choice of dose magnitude and frequency in the design of a planned
9 phase II study (assessing prophylactic efficacy in higher-risk populations) and,
10 pending a favourable outcome, any subsequent global phase III studies. It is only
11 during the latter two phases that a true determination of the efficacy and thus
12 feasibility of parenteral HIV PrEP can be made.

13 ARV treatment depends on combination therapy, whereas current PrEP
14 injectable agents are investigated as single drug prevention strategies because it is
15 hypothesized that these would be sufficient to prevent HIV infection. However,
16 delayed or missed injections could lead to prolonged periods of suboptimal drug
17 exposure and increased risk of HIV acquisition; selective drug pressure in the
18 presence of a replicating virus could enable expansion of drug resistance and
19 onward transmission of resistant HIV strains. This may be especially true for
20 nonnucleoside reverse-transcriptase inhibitors, in which single viral genotype
21 mutations remarkably compromise the efficacy of this drug class,(Bannister et al.
22 2008) whose constituents have for several years been the most commonly used

1 third agents in first-line combination antiretroviral therapies.(World Health
2 Organisation 2013) 16

3 In conclusion, this study is the first to investigate RPV concentrations in the
4 female genital tract and male rectal compartment after the i.m. administration of
5 different RPV-LA doses to humans, and the first to assess the correlation between
6 compartmental drug exposure and HIV growth ex vivo. This study encourages
7 further development of RPV-LA as a potential PrEP intervention.

1 4 A CASE OF HIV SEROCONVERSION IN SSAT040

2 *AGAJ, as the attending research physician, was responsible for the diagnosis,*
3 *emergency management and ongoing follow-up of the patient described in this*
4 *seroconversion case report. As Lead clinician on this parent study, AGAJ instituted*
5 *the protocol-specified antiretroviral initiation plan for this patient and devised a*
6 *bespoke schedule of comprehensive prospective sample collection, whilst putting in*
7 *place an investigation to uncover the events leading to the proposal. AGAJ presented*
8 *a report based on the investigation of these events to the protocol steering*
9 *committee. On the basis of the ensuing discussion, a plan was enacted to engage the*
10 *clinical academic capabilities of the Mellors' Group at the University of Pittsburgh,*
11 *School of Medicine. In doing so, this new collaborative group was able to conduct a*
12 *genotypic forensic examination of archived and prospective samples, reproducing in*
13 *the lab the time course of the seroconversion. AGAJ coordinated this investigation*
14 *and drafted the brief report, as second senior author, which was published in the*
15 *Journal of Infectious Diseases, November 2015.*

16 The aim of the study SSAT040 was to determine the tolerability and
17 compartmental pharmacokinetics of a solid drug nanoparticle suspension of
18 rilpivirine in plasma, whilst simultaneously determining its distribution to tissue sites
19 of relevance to risk during sexual exposure to HIV infection; vagina and rectum.

20 As outlined in section 3.2.2, participants were systematically screened with a risk-
21 based medical and sexual history in an attempt to enrol an healthy volunteer cohort,
22 whose historical or current and likely future risk of HIV acquisition was minimised.

1 Despite this, unfortunately there was an occurrence of breakthrough HIV-1
2 infection in 1 female participant, enrolled in the 300-mg dose arm.

3 This was discovered only at the time of positive serology being confirmed from
4 the final study sampling visit when she attended for follow-up. The participant was
5 immediately recalled to clinic and received relevant clinical supportive care for a
6 newly diagnosed patient. In addition and in line with the pre-specified protocol
7 management pathway, empirical antiretroviral therapy with high genetic barrier
8 combination of unexposed drug classes was initiated.

9 In agreement with the participant, now patient, a reconstruction of events from
10 the her first study visit allowed her recall to isolate with a high degree of certainty
11 the specific sexual encounter at which transmission is likely to have occurred. This
12 was a single episode of vaginal intercourse with a new male partner, without the use
13 of barrier contraception. The partner's HIV status, was unknown at the time of the
14 encounter and he reported having a negative HIV antibody test within the three
15 months prior. Though the partner was visiting the UK at the time of the event, he
16 returned to his home country in southern Africa and was not available during the
17 course of the follow-up of the index case.

18 This breakthrough infection was evaluated with the consent of the patient,
19 allowing the collection and ongoing archive of follow up samples for at least six
20 months after seroconversion, for plasma, serum and isolated peripheral blood
21 mono-nuclear cells. We also obtained consent from the patient to engage the
22 technical capability of a collaborating group at the Division of Infectious Diseases,

1 Department of Medicine, University of Pittsburgh School of Medicine, Pennsylvania
2 allowing the anonymous transfer of data and samples for further investigation.

3 Herein, are reported the levels of RPV in plasma and cervicovaginal fluid (CVF)
4 and the emergence of NNRTI-resistant HIV-1, with discussion of the potential impact
5 of this resistance on subsequent treatment with first-line NNRTIs

6 The duration of TMC278LA detectability after injection has subsequently been
7 formally investigated and demonstrated to exceed 150 days(McGowan et al. 2016),
8 but the levels of active drug required to prevent HIV-1 infection have not yet been
9 determined.

10 The prevalence of RPV-associated mutations is only 5% in treatment-naive
11 individuals but is as high as 59% in patients with no response to NNRTI-containing
12 first-line antiretroviral therapy (ART). (Anta et al. 2013; Lambert-Niclot et al. 2014;
13 Parczewski et al. 2014; Theys et al. 2015)

14 The July 2014 International AIDS Society USA HIV-1 drug resistance mutation
15 update recognised 17 mutations associated with RPV drug resistance, including
16 L100I, K101E/P, E138A/G/K/Q/R, V179L, Y181C/I/V, Y188L, H221Y, F227C, and
17 M230I/L,(Wensing et al. 2014) which confer up to 50-fold resistance to RPV.(Azijn et
18 al. 2010; Wensing et al. 2014; Basson et al. 2015)

19 During the ECHO and THRIVE trials, which evaluated RPV in a background of 2
20 NRTIs, E138K with M184I most commonly emerged in individuals with virologic
21 failure. Of the samples from patients with virologic failure in these trials who

1 exhibited phenotypic resistance to RPV, 46% had virus with cross-resistance to
2 nevirapine (NVP), 86% had virus with cross-resistance to efavirenz (EFV), and 91%
3 had virus with cross-resistance to etravirine (ETR).(Cohen et al. 2013)

4 .

5 **4.1 MATERIALS AND METHODS**

6 SSAT040 study outline and conduct is reported above. Participants for the
7 SSAT040 study were verified to be HIV seronegative by the 4th Generation GS HIV
8 Combo Ag/Ab EIA HIV-1/2+O (Bio- Rad) prior to enrolment and were deemed to be
9 at low risk for HIV infection as indicated by self-report. Plasma samples obtained at
10 multiple time points from the participant with breakthrough infection were
11 retrospectively tested for HIV-1 RNA (COBAS AmpliPrep/COBAS TaqMan HIV-1 Test,
12 v2.0, Roche Molecular Diagnostics) and for HIV-1 antibody at multiple time points,
13 using the Bio-Rad 4th Generation GS HIV Combo Ag/Ab HIV-1/2+O EIA. Plasma and
14 CVF were collected at regular intervals over 84 days for pharmacokinetic analysis.
15 Resistance testing was performed on a residual sample collected following
16 pharmacokinetic analysis. Sufficient plasma specimens for resistance testing were
17 not available at all time points.

18 **4.1.1 RESISTANCE ANALYSIS**

19 Resistance analysis was conducted in the laboratories at the University of
20 Pittsburgh School of Medicine, by the Division of Infectious Diseases group led by
21 John Mellors.

1 HIV-1 RNA from plasma samples was isolated using the ViroSeq 2.0 extraction
2 module (Celera). In addition, RNA from plasma samples collected in heparinised
3 tubes was precipitated in 3 M LiCl prior to amplification, to alleviate polymerase
4 chain reaction (PCR) inhibition. Protease and full-length reverse transcriptase (amino
5 acids 1 through 560) were amplified using in-house primers (OF1-BCD-5'-
6 GAGGGACACCAAATGAAAG AYTG-3' and 4232 5'-CCTGACTTTGGGGATTGTAGGGA
7 AT-3') in a 1-step reverse-transcription PCR, using SuperScript III One-Step
8 (Invitrogen), followed by a second nested PCR, using in-house primers (Bcl1-5'-
9 TAAGACAGTATGATCAAA TACTTATAGAAATTTGTGG-3' and Xho1-5'-TAACTTTT
10 CCCTCGAGATGTGTACAATCTAATTGCC-3'), to amplify full-length reverse
11 transcriptase.

12 Sequence was generated by means of 6 bidirectional primers, using Sanger
13 sequencing methods. HIV-1 drug resistance mutations were identified using the
14 HIVdb program v7 (Stanford University) [14]. Allele-specific PCR for K101K and
15 K101E was performed using RNA isolated for standard genotyping. Complementary
16 DNA species were amplified and quantified in a first-round PCR, using Phusion Hot
17 Start II High Fidelity Polymerase (Thermo Fisher Scientific) and HIV-1 subtype C
18 reverse transcriptase– specific primers 28F 5'-AAACAATGGCCATTGACAGAAGA-3'
19 and 80R 5'-GTTTCATACCCCATCCAAAGAAATG-3'.

20 A total of 1×10^6 copies of first-round PCR product were used in an allele-specific
21 PCR reaction, using AmpliTaq Gold (Applied Biosystems) and the following patient-
22 specific primers: forward wild-type primer 5'-GGAATACCGCACCCAGCAGGATTCA-3',
23 forward 101E primer 5'-GGAATACCGCACCCAGCAGGATT CG-3', and reverse primer

1 5'-CTCTGGAATATTGCTGGTG ATCCTT-3'. The detection limit for K101E is 0.1%, based
2 on testing a panel of K101:101E PCR amplicon mixtures included in each assay.

3

4 **4.1.2 PHENOTYPIC ANALYSIS**

5 Full-length reverse transcriptase amplicons created for genotyping from the day
6 115 sample were used to generate recombinant patient-derived virus, using a
7 modified HIV-1xxLAI vector [15]. PCR amplicons from the day 115 sample were
8 cloned into Bcl1- Xho1 linearized xxLAI plasmids, using InFusion cloning technology
9 (Clontech). DNA generated from single-colony bacterial isolates was selected based
10 on having either K101 or 101E and having identical sequence for the remainder of
11 the reverse transcriptase gene. Midiprep DNA from these isolates were used to
12 transfect 293T cells, using Lipofectamine2000 (Invitrogen), and viral supernatants
13 were collected 48 hours after transfection. Patient-derived viral stocks were used in
14 drug susceptibility assays using TZM-bl cells and normalized for output of 100
15 relative light units in virus control wells. Fifty percent inhibitory concentrations
16 (IC50) were determined for RPV, nevirapine (NVP), efavirenz (EFV), and etravirine
17 (ETR).

18

19 **4.2 RESULTS**

20 **4.2.1 HIV-1 INFECTION AND RPV LEVELS**

21 One female participant in the SSAT040 study acquired HIV-1 infection through
22 heterosexual intercourse without a condom with a new male partner approximately

1 41 days following receipt of a 300-mg single intramuscular injection of TMC278LA.
2 The male partner was subsequently found to have acute HIV-1 infection at the time
3 of transmission. Retrospective testing showed that the SSAT040 participant had
4 undetectable plasma HIV-1 RNA (level, <100 copies/mL) on the day she received the
5 TMC278LA injection (day 0). HIV-1 RNA was first detected on day 57 (level, 370
6 copies/mL), followed by seroconversion on day 84. The peak viremia level (644 925
7 copies/mL) occurred at day 115 and declined after initiation of ART with tenofovir/
8 emtricitabine and ritonavir-boosted darunavir (800/100 mg) on day 115 but
9 remained detectable through day 275 (level, 125 copies/mL) until finally achieving
10 viral suppression below the limit of quantitation (level, <40 copies/mL) on day 309
11 (Table 4:A, Figure 4:A). Both plasma and CVF drug concentrations peaked on day 15
12 (68.1 ng/mL and 294.9 ng/mL, respectively) and then dropped 6.5-fold and 16-fold
13 respectively by day 41 (the suspected day of HIV exposure), but low levels of RPV
14 were still detectable in plasma on day 226 (4.0 ng/mL). RPV drug concentrations
15 were approximately 10.5 ng/mL and 18.3 ng/mL in plasma and CVF, respectively, at
16 the time of HIV-1 exposure.

Table 4:A Rilpivirine Concentration, HIV Infection History and Drug Resistance Selection in a Seroconverter from the SSAT040 Study

Days post- IM injection ^a	[RPV] ng/mL			HIV Serostatus	Plasma HIV-1 RNA (c/mL)	CD4 (%)	Resistance Mutations ^b	% K101E ^c
	Plasma	CVF	Vaginal Tissue					
SCREENING				Neg				
0	0				<100			
0.17	4.08							
0.34	18.21	7.54						
1	37.42	51.28			<100			
4	53.62	91.83			<100			
7	58.16	133.25			<100			
11	42.6				<100			
15	68.11	294.89			<100			
22	24.34	135.31			<100			
28	24.33	32.89	243.23	Neg	<100			
44 ^d	10.49	18.3			<100			
57	6.78	11.2	23.05	Neg	370			
84	7.51	13.97		Pos	175060		none	0
115 ^e	4.05			Pos	644925	410 (22%)	K101EK ^f	19.4
122					74964	526 (34%)		
136					18006	692 (44%)		

<i>Days post- IM injection^a</i>	[RPV] ng/mL	HIV Serostatus	Plasma HIV-1 RNA (c/mL)	CD4 (%)	Resistance Mutations ^b	% K101E ^c
151	13.75		6204	617 (49%)	K101EK	
175			2998	605 (49%)		
199	6.00		3558	834 (53%)	none	0.1
211			1184			
226	4.00		769			
275			125			
309			<40			

Abbreviations: IM, intramuscular; RPV, rilpivirine; CVF, cervicovaginal fluid; c/mL, copies/mL; neg, negative; pos, positive

^a Participant was given a single dose of 300 mg TMC278LA.

^b Resistance mutations were determined by standard genotyping.

^c % K101E was determined by allele-specific PCR, with a limit of detection of 0.1%

^d HIV exposure self-reported to occur on day 41. Day 44 samples are the first available after reported exposure.

^e ART initiation with Truvada® and ritonavir-boosted darunavir (800/100mg)

^f Susceptibility of d115 patient-derived recombinant HIV-1 isolates with K101E (HIV-1_{d115/K101E}) to RPV, nevirapine (NVP), efavirenz (EFV) and etravirine (ETR) was compared with d115 wild type HIV-1 isolates (HIV-1_{d115/WT}).

HIV-1_{d115/K101E} had an RPV IC₅₀ of 1.6 nM (4.3-fold resistance) compared to HIV-1_{d115/WT} (IC₅₀ 0.4 nM).

HIV-1_{d115/K101E} had an NVP IC₅₀ of 368 nM (7.9-fold cross-resistance) compared to HIV-1_{d115/WT} (IC₅₀ 46 nM).

HIV-1_{d115/K101E} had an EFV IC₅₀ of 2.8 nM (4.0-fold cross-resistance) compared to HIV-1_{d115/WT} (IC₅₀ 0.7 nM). HIV-1_{d115/K101E} had an ETR IC₅₀ of 4.0 nM (4.0-fold cross-resistance) compared to HIV-1_{d115/WT} (IC₅₀ 1.0 nM)

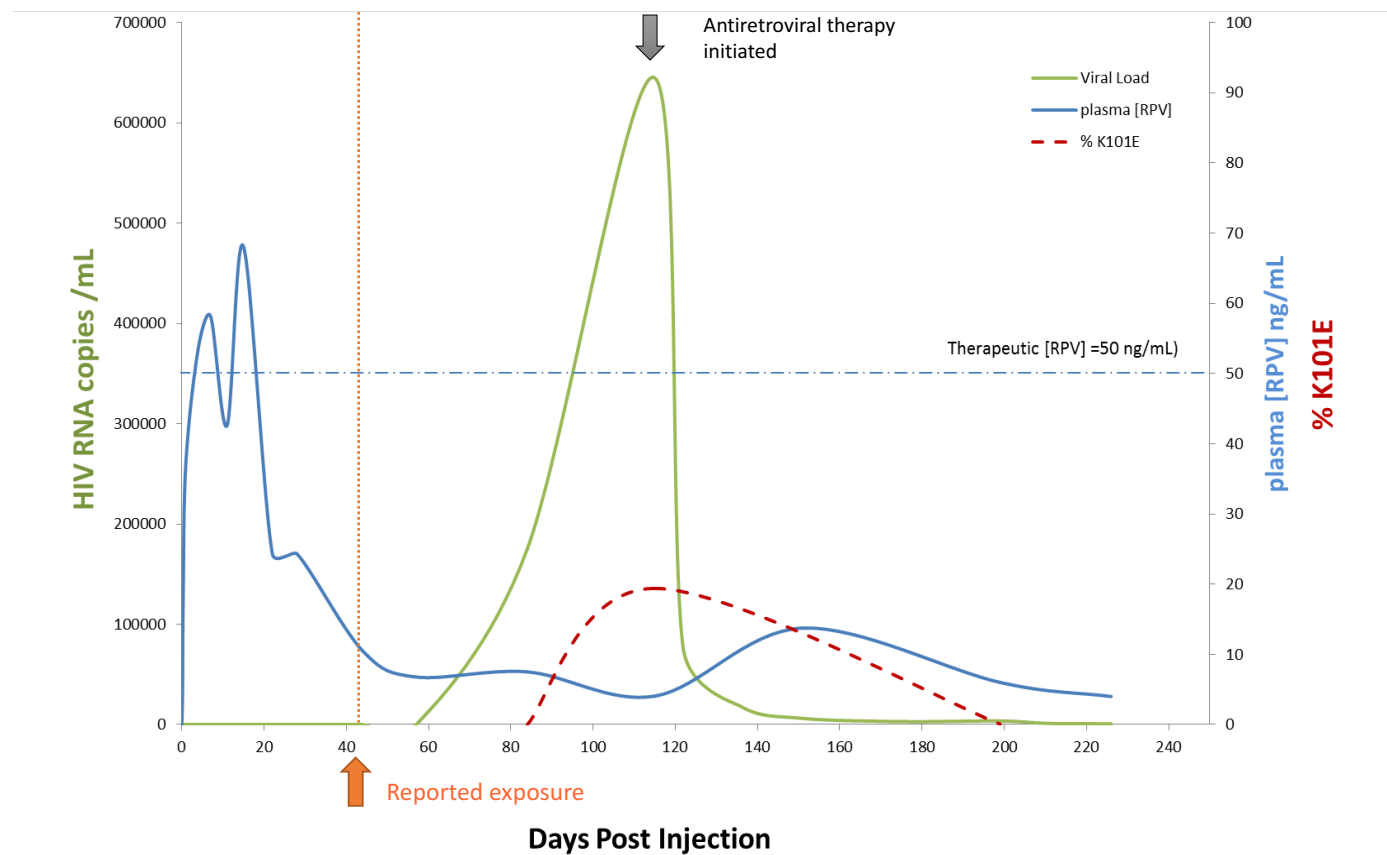


Figure 4:A Overlaid chronologic plot summarising HIV-1 RNA Copies/mL, Plasma Rilpivirine Concentration and Selection of K101E in seroconversion case.

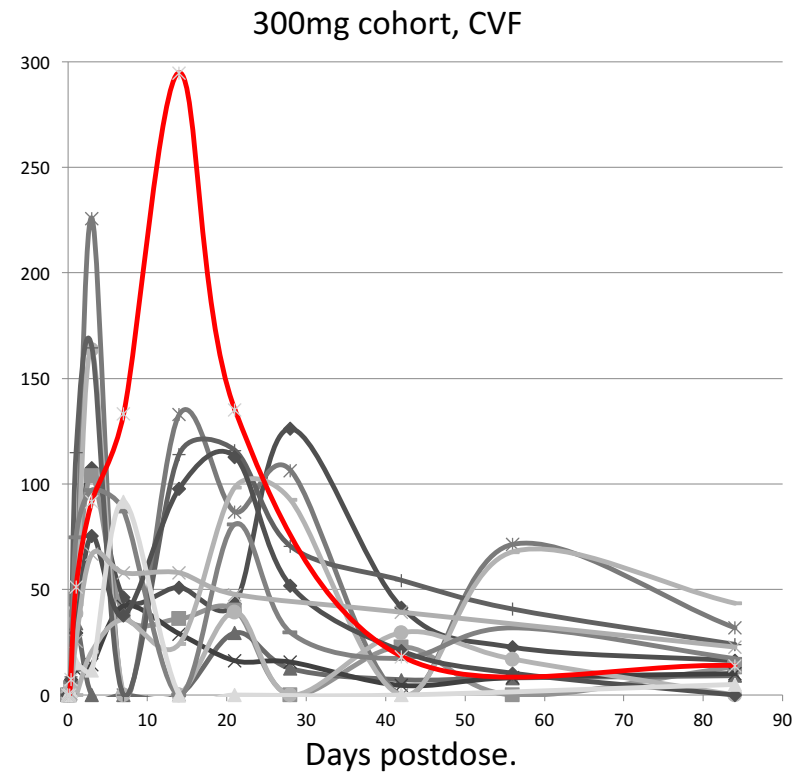
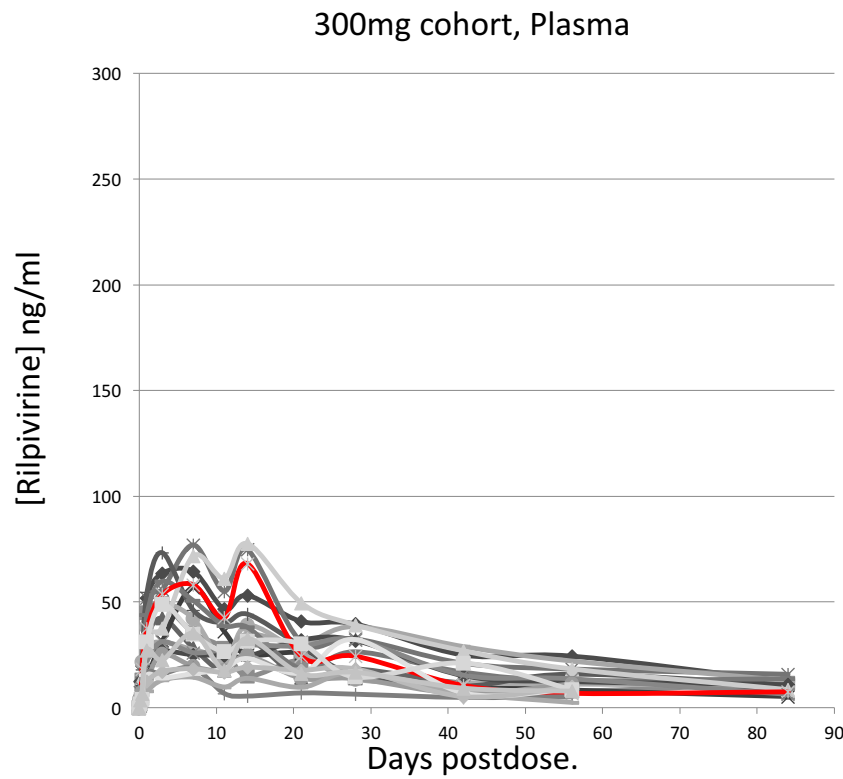


Figure 4:B Individual plasma and CVF concentrations over 84 days following a single intramuscular dose of 300mg.

Concentration plot for seroconversion case in red..

1

2 **4.2.2 SELECTION OF HIV-1 DRUG RESISTANCE**

3 After evaluation of residual plasma samples, collected for pharmacokinetic
4 analysis, for the presence of selected or transmitted drug resistance (Table 4:A),
5 infection with wild-type virus, as determined by both standard population
6 sequencing and allele- specific PCR with an assay sensitivity of 0.1% for 101E, was
7 confirmed during acute infection on day 84. This was 43 days after the estimated
8 date of HIV-1 infection. At the peak viremia level, on day 115, however, a mixed
9 population of 101K/E was detected by both population sequencing and allele-
10 specific PCR (19% 101E). By day 199, the predominant viral population was wild
11 type, with only 0.1% 101E detected by allele-specific PCR (Table 4:A). Samples from
12 the participant's sex partner were not available for testing.

13 **4.2.3 CROSS-RESISTANCE TO NNRTIS**

14 The day 115 patient-derived recombinant viral isolates containing either 101E or
15 101K was used to evaluate phenotypic resistance to RPV and cross-resistance to
16 NVP, EFV, and ETR.

17 Day 115 isolates containing 101E had 4.3-fold greater resistance (IC₅₀, 0.58
18 ng/mL) to RPV, compared with day 115 isolates containing K101 (IC₅₀, 0.13 ng/mL).

19 Isolates containing 101E had 7.9-fold greater cross-resistance to NVP and 4.0-fold
20 greater cross-resistance to both EFV and ETR (IC₅₀, 98 ng/mL vs 12 ng/ mL for NVP,
21 0.88 ng/mL vs 0.22 ng/mL for EFV, and 1.8 ng/mL vs 0.44 ng/mL for ETR (Table
22 4:B)).

1 Table 4:B Seroconversion ex vivo phenotypic analysis

	RPV		NVP		EFV		ETR	
	Mea	Fold	Mea	Fold	Mea	Fold	Mea	Fold
	n IC ₅₀	Change	n IC ₅₀	Change	n IC ₅₀	Change	n IC ₅₀	Change
	(ng/mL)		(ng/mL)		(ng/mL)		(ng/mL)	
<i>K101K</i>	0.13	-	12	-	0.22	-	0.44	-
<i>K101E</i>	0.58	4.3	98	7.9	0.88	4.0	1.8	4.0

2

3 Full-length reverse-transcriptase amplicons created for genotyping from the day
4 (d) 115 sample were used to generate recombinant patient-derived virus using a
5 modified HIV-1_{xxLAI} vector (see Materials and Methods).

6 Patient-derived viral stocks were used in drug susceptibility assays using TZM-bl
7 cells and normalized for output of 100 RLU in virus control wells. Susceptibility to
8 RPV, nevirapine (NVP), efavirenz (EFV) and etravirine (ETR) of d115 patient-derived
9 recombinant HIV-1 isolates with K101E (HIV-1_{d115/K101E}) was compared with d115
10 wild type HIV-1 isolates (HIV-1_{d115/WT}) to generate fold-change resistance values.

11

12 **4.2.4 DISCUSSION**

1 RPV concentrations in plasma and CVF after a single injection of 300 mg of
2 TMC278LA were insufficient to prevent infection in 1 female participant from the
3 SSAT040 study. The infection event occurred 41 days following TMC278LA
4 administration, when the plasma RPV concentration was 10.5 ng/mL and the CVF
5 RPV concentration was 18.3 ng/mL. Although the concentration of RPV required for
6 prevention is not known, the RPV levels at the time of HIV exposure were well below
7 the proposed target concentration of 50 ng/mL that is the minimum needed for
8 virologic response when RPV is used as ART.(Cohen et al. 2013) Of note, the infected
9 participant displayed higher peak concentrations of RPV in CVF and vaginal tissue,
10 compared with other participants in the same 300-mg arm (who may or may not
11 have had exposure to HIV), but free drug concentrations and the pharmacodynamic
12 effect of RPV in these sites remains undefined.(McGowan et al. 2016) RPV levels
13 continued to be low but detectable through the testing period of 226 days.

14 High levels of viral replication combined with low but detectable levels of RPV led
15 to the selection of resistant variants containing K101E. Initial infection occurred with
16 wild-type virus, as demonstrated by both standard and sensitive resistance testing,
17 and thus resistance was selected by TMC278 and not transmitted from the
18 participant's partner. A mixed population of viruses, consisting of approximately
19 80% wild-type and 20% mutant 101E variants, emerged by day 115, coinciding with
20 peak viremia.

21 Phenotypic testing of the day 115 mutant showed resistance to RPV (4-fold) and
22 cross-resistance to NVP (7-fold), EFV (4-fold), and ETR (4-fold), which indicates that
23 the K101E mutant could negatively influence the virologic response to first-line

1 NNRTI-containing ART. The 101E variants declined to undetectable levels in parallel
2 with the rapid decline in viremia level after ART initiation (Figure 4:A).

3 This case report is a unique instance of well-documented infection with wild-type
4 HIV-1 and subsequent selection of resistant virus by continued drug exposure from a
5 long-acting drug formulation. The 300mg dose of TMC278LA was insufficient to
6 prevent infection and limit viral replication but was high enough to select RPV-
7 resistant virus, illustrating a negative consequence of a long-acting antiretroviral
8 formulation. RPV concentrations that prevent HIV infection prevention are
9 undefined, but the 300mg dose of TMC278 did not achieve RPV levels above the
10 proposed therapeutic concentration for virologic suppression (50 ng/mL). Further
11 studies are needed to determine a target RPV concentration for HIV-1 prevention
12 and to evaluate the risk of resistance during breakthrough infection with higher
13 doses of TMC278. Both plasma and vaginal concentrations of RPV must be
14 considered when selecting the appropriate dose for HIV prevention.

15 It is important to note the limitations inherent in the investigation of this
16 seroconversion whilst receiving chemoprophylaxis which was insufficient to
17 abrogate the establishment of infection. These limitations are inherent to the design
18 of the primary study protocol, wherein the intention was to avoid HIV infection by
19 selecting volunteer participants at low behavioural risk of infection for
20 pharmacokinetic exploration, without testing pharmacodynamic efficacy of
21 prevention of the investigational therapy.

1 Firstly, resistance testing was done using residual plasma specimens obtained
2 after pharmacokinetic testing, so all intermediate time points were not available for
3 standard or sensitive resistance testing.

4 Additionally, despite prolonged exploration of a variety of logistic means to allow
5 the identified sex partner of the index case to consent to collection, processing and
6 shipping of samples from his country of origin in Southern Africa, these repeated
7 efforts were ultimately unsuccessful over a period of greater than 12 months. The
8 objective of obtaining samples would have served two purposes. The primary
9 importance, which became evident on discussion of the partner's access to clinical
10 care within his home country, was that though therapy was accessible to him, this
11 would likely be based on empirical initiation of a NNRTI-based antiretroviral therapy,
12 as primary resistance testing would only be available at through commercial
13 laboratories. Therefore, it would be important to confirm that he was indeed
14 infected with wild-type virus based on the evidence of the investigation of the
15 transmitted virus, rather than the harbouring the mutation seen as a minority
16 species. Provision of this result to the patient would help to guide his choice, with a
17 local healthcare provider, of initial therapy ensuring predicted full activity of all
18 components. Secondly, confirming his infection with wild-type virus, would
19 corroborate the finding that the founder virus in the recipient was wild type at
20 codon 101, based on sensitive allele-specific PCR.

21 Nonadherence to daily or coitally dependent PrEP has emerged as the major
22 barrier to PrEP effectiveness. Long-acting formulations have the advantage of
23 infrequent dosing, but sustained, low drug concentrations for months after an

1 injection is problematic for the selection of drug resistance when breakthrough
2 infection occurs. The post-hoc interrogation of the events occurring in this
3 seroconversion highlights the importance of frequent HIV-1 testing with PrEP use
4 and the need to develop a safe method for discontinuing long-acting products to
5 avoid infection and drug resistance.

6 Elimination of rilpivirine is primarily through undergoing oxidative metabolism
7 mediated by the cytochrome P450 (CYP) 3A system, with 85% recovery of
8 radiolabelled dose in faeces and 6% in urine, with only trace amounts as the
9 unchanged moiety (<1%). Given its high binding affinity to plasma proteins,
10 predominantly albumin (99.7%), haemodialysis is unlikely to remove a significant
11 proportion of the administered dose. In Table 4:A and Figure 4:A between the time
12 of initiation of a ritonavir-boosted protease inhibitor combination on day 115 and
13 the final PK follow-up sample on day 226, the plasma concentration of rilpivirine
14 remains above 4 ng/mL across this period. Plasma concentrations at the two
15 intervening (days 151 and 199) were 13.75 and 6.00 ng/mL respectively. The
16 individual PK plots over time (Figure 3:C, Figure 3:E) illustrate a high degree of
17 variability in sequential concentrations at interval, such that this apparent secondary
18 peak in the presence of ritonavir may simply be sampling of the peaks and trough in
19 that “tail” , rather than causally related to reduce elimination through cytochrome
20 inhibition in the presence of ongoing release from the depot. It would not be
21 unreasonable to infer that during the period of almost 4 months from day 115
22 period, the whole body clearance of rilpivirine was closely matched to the rate of
23 release from the depleting depot. This raises a number of unanswered questions

1 with implications for registration phase study design and subsequent real-world use.
2 Taking a base assumption that the 300mg dose has been demonstrated herein not
3 to afford pharmacokinetic exposure able to sustain exposure above the putative
4 target minimum threshold (50 ng/mL), planned future development phases have
5 been able to eliminate, on the basis of futility, both the 300mg dose as well as
6 higher doses which at predicted steady state, had concentrations at the end of the
7 desired dosing interval lower than the above target threshold.

8 Thus most importantly, the dose of injectable long-acting formulations such as
9 TMC278LA must be high enough and delivered at a frequency which enough to
10 achieve drug levels that prevent HIV-1 infection.

11

5 SSAT049: RILPIVIRINE AND BOOSTED-DARUNAVIR DUAL THERAPY

AGAJ acted as Chief Investigator for this study and was responsible for designing the concept and securing industry funding in the form of an unrestricted grant, writing the protocol and leading the submissions for both ethical and regulatory approvals. AGAJ wrote standardisation manual and study report forms, as well as convening the Protocol Steering Committee to oversee the safe conduct of the study. AGAJ was responsible for the clinical care of all participants on study, including their recruitment from ambulatory clinics, obtaining informed consent and collecting clinical history and assessing physical examination and determining eligibility. Whilst on study, AGAJ was fully responsible for clinical management of these patients. AGAJ participated in the analysis of the bioanalytic output and presented the data as a poster at the 2014 Conference on Retroviruses and Opportunistic Infections and was lead author on the manuscript which has now been published in HIV Clinical Trials, 2018, (Jackson et al. 2017).

5.1 BACKGROUND

Although combination antiretroviral therapy (cART) using three drugs, is recommended by international guidelines for the treatment of HIV infection,(Churchill et al. 2016) (Panel on Antiretroviral Guidelines for Adults and

1 Adolescents 2016) strategies which spare the use of drug classes, such as “dual
2 therapies” in which combinations of 2 effective agents from different classes based
3 on their mechanism of action, have been more recently been explored. There are a
4 number of studies in which the putative aim of the avoidance of a drug class was to
5 explore the potential to improve aspects of tolerability, adherence, and reduce
6 observed toxicity and cost.

7 The NEAT001/ANRS143 study compared a combination of ritonavir-boosted
8 darunavir 800mg/100mg once daily combined with either; raltegravir 400mg twice
9 daily as the NtRTI-sparing dual-class regimen, or with tenofovir disoproxil fumarate/
10 emtricitabine 245mg/200mg once daily fixed-dose formulation as the standard
11 triple class regimen in a randomised, non-inferiority comparison in previously
12 untreated HIV-positive patients starting therapy for the first time in 15 European
13 countries.(Raffi et al. 2014) The NtRTI sparing dual therapy was found to be non-
14 inferior to standard therapy with treatment failure rates after 96 weeks of 17.8%
15 and 13.8% respectively with a difference of 4.0%, with 95% confidence interval
16 overlapping zero. The rate of occurrence of adverse events, classified as *serious* or
17 which required modification of therapy, were similar between both groups. The
18 expert study group came to the conclusion that this boosted-PI combination with
19 integrase strand-transfer inhibitor (INSTI) could be a reasonable option for patients
20 starting therapy with a CD4 count greater than 200 cells per μL .

21 Another head to head, open-label non-inferiority study conducted by the
22 OLE/RIS-EST13 study group in 32 hospitals in Spain and France, compared the
23 outcomes of patients already on a combination of ritonavir-boosted lopinavir,

1 combined with a two drug NRTI backbone containing either lamivudine or
2 emtricitabine with another nucleotide/nucleoside.(Arribas et al. 2015) Within the
3 study, 250 patients were randomised to either continue three drug therapy or to
4 switch (reduce) to a combination of lopinavir/ritonavir 400mg/100mg twice daily
5 with lamivudine 300mg once daily. Over 48 weeks after switching, the dual therapy
6 rate of maintenance of treatment response was 87.8% (108/123), compared to
7 86.6% (110/127) in those remaining on three drugs. With non-inferior efficacy by
8 statistical tests and similar safety, in terms of adverse events and discontinuations,
9 this combination of a boosted PI and NRTI was also demonstrated to be a suitable
10 switch option.

11 More recently, a proof-of-concept pilot trial explored in a single-arm study design
12 of 20 therapy naïve patients starting once-daily dual combination of the INSTI
13 dolutegravir 50mg with lamivudine 300mg; the PADDLE study.(Cahn et al. 2017) All
14 patients were screened for baseline genotype and started therapy with a viral load
15 below 100,000 copies/mL (maximum 36,000 c/mL). Plasma viraemia was rapidly
16 suppressed in all patients within the first 8 weeks, to below assay limits of
17 quantification. Of the 20, one patient committed suicide and another experienced
18 protocol-defined low level virologic failure at week 36, with return to suppression by
19 the final visit of the study whilst remaining on therapy. This study is of particular
20 interest as the dolutegravir and lamivudine (at these doses) are two components of
21 the licensed single tablet fixed-dose once-daily combination (Triumeq®; dolutegravir
22 50mg/ abacavir 600mg/ lamivudine 300mg) (ViiV Healthcare 2014) but avoiding the
23 potential toxicities which have been associated with the use of abacavir.

1 Within the aforementioned guidelines, cART containing two nucleoside reverse
2 transcriptase inhibitors (NRTIs) plus a third agent are the most commonly
3 recommended combinations (Churchill et al. 2016; Panel on Antiretroviral
4 Guidelines for Adults and Adolescents 2016). However, long-term exposure to NRTI
5 may lead to the development of adverse events with two of the most commonly
6 used; tenofovir disoproxil fumarate(Gilead Sciences Inc. 2002) [potential long-term
7 renal or bone toxicity] or abacavir sulfate(ViiV Healthcare 2000) [HLA-B*5701
8 related potentially severe hypersensitivity reactions and retrospectively observed
9 increased risk of myocardial infarction].

10 Additionally, there are important considerations outside the immediate patient
11 which are related to successes in treatment coverage which has co-evolved with
12 secular trends in the public health epidemic.

- 13 I. the possible transmission of virus already containing resistance
14 mutations, such that patients may have a viral genotype with less than
15 optimal susceptibility to the commonly used NRTIs,(Baxter et al. 2015)
- 16 II. the low genetic barrier of certain NRTIs like lamivudine (3TC) and the
17 likelihood of some patients to harbour resistance to this or other NRTIs if
18 they have a history of poor adherence(Churchill et al. 2016) and
- 19 III. the rapidly increasing use of tenofovir disoproxil fumarate
20 (TDF)/emtricitabine (FTC)-base pre-exposure prophylaxis (PrEP) for HIV
21 that may increase the development of resistance in individuals who
22 seroconvert on PrEP,(Dimitrov et al. 2016), are other important factor to
23 consider when investigating optimal alternative cART.

1 Consequently, there is importance in the evaluation of the of dual-therapy
2 options, particularly where the patterns of use of NRTI-sparing may show an
3 increased frequency of occurrence.

4 Rilpivirine, is an antiretroviral agent approved in Europe and the USA for
5 treatment of therapy-naïve HIV-1 infected adult patients (with a viral load \leq
6 100,000 copies/mL) in combination with other agents. (Molina et al. 2012; Sanford
7 2012) A member of the non-nucleoside reverse transcriptase inhibitor (NNRTI) class,
8 with a diarylpyrimidine structure, its activity mediated by non-competitive inhibition
9 of HIV-1 reverse transcriptase. Rilpivirine is primarily undergoes oxidative
10 metabolism mediated by the cytochrome P450 (CYP) 3A system, therefore drugs
11 that induce or inhibit CYP3A may affect its clearance. The recommended dose is 25
12 mg given once daily, taken with a meal of at least 533 kcal.(Sanford 2012) (Janssen-
13 Cilag Ltd 2011)

14 Darunavir, is a member of the protease inhibitor (PI) class. When co-administered
15 with low dose ritonavir it is approved, in combination with other antiretroviral
16 agents, for the treatment of patients with HIV-1. In treatment-naïve patients the
17 recommended dose is 800 mg once daily with ritonavir 100 mg once daily taken
18 with food.(Sherer 2007) (Janssen-Cilag Ltd 2015)

19 Ritonavir is a potent inhibitor of cytochrome CYP3A4 and when administered at a
20 low dose of 100 mg once daily, it causes clinically significant increases in the plasma
21 exposure of protease inhibitors and numerous other drugs processed by this
22 common metabolic pathway.(Moyle and Back 2001)

1 A phase I study investigating the drug interaction between rilpivirine dosed at
2 150 mg once daily and darunavir/ritonavir in HIV-negative healthy volunteers
3 showed an increase in rilpivirine pharmacokinetic parameters ranging between 80
4 to 180% with no changes in darunavir or ritonavir concentrations. (Santoscoy et al.
5 2008)

6 However, data in adult antiretroviral naïve patients living with HIV (PLWH) on the
7 currently approved rilpivirine dose of 25 mg once daily are lacking. Yet, a
8 combination of rilpivirine and ritonavir-boosted darunavir could potentially form a
9 once daily NRTI-sparing treatment, as shown in a switch study (Maggiolo et al. 2016)
10 and in a study in adolescents by. (Foca et al. 2016) Hence, this study investigated the
11 antiviral activity, steady state pharmacokinetics and safety of rilpivirine plus
12 darunavir/ritonavir in therapy-naïve HIV-infected adults.

13

14 5.2 METHODS

15

16 5.2.1 PARTICIPANTS

17

18 Written informed consent was obtained from PLWH, aged 18 to 65 years old,
19 naïve to cART with a VL > 1000 copies/mL and a CD4 count > 50 cells/mm³.
20 Participants were excluded if they had significant acute or chronic
21 psychiatric/medical illnesses that could have interfered with the ability of taking part

1 in the study, anomalies and risk factors for QTc prolongation or clinical laboratory
2 determinations; positive screens for hepatitis B/C; baseline transmitted resistance
3 compromising rilpivirine and darunavir efficacy; use of other drugs known to
4 interact with the study drugs.

5

6 **5.2.2 STUDY DESIGN**

7

8 This was a single-arm, open-label study approved by the City and East Research
9 Ethics Committee and the Medicines and Healthcare products Regulatory Agency
10 (MHRA), UK (EUdraCT - 2012-002663-10; Clinicaltrials.gov NCT01736761).

11 Following recruitment of the first 10 participants with baseline viral loads below
12 100,000 copies/mL (group A), a protocol steering committee convened to review
13 viral load responses over the first four weeks of therapy and to advise on whether to
14 proceed with recruitment of participants with baseline viral loads above 100,000
15 copies/mL (group B).

16 After successful screening, participants were administered rilpivirine 25 mg plus
17 darunavir/ritonavir 800/100 mg once-daily, and given weekly appointments until
18 week 4 for drug concentration measurement (trough concentration, C_{trough}), HIV-
19 RNA testing and resting ECG monitoring. At week 4, they were admitted to the
20 research unit for 24 hour pharmacokinetic sampling, blood was taken pre-dose, 1, 2,
21 4, 6, 8, 12 and 24 hours post-dose. Follow up appointments for safety laboratory

1 test, viral load measurement and ECG monitoring were at weeks 6, 8, 10, 12, 24, 36
2 and 48 post cART initiation.

3 On the pharmacokinetic day, the study medication was taken with a standardised
4 breakfast (534 kcal) and 240 mL of water. Compliance with study drug
5 administration was assessed by pill counting by the study staff throughout the study
6 period.

7

8 **5.2.3 BIOANALYSIS (DRUG PLASMA CONCENTRATION MEASUREMENT)**

9

10 Blood samples were collected for the measurement of rilpivirine, darunavir,
11 ritonavir concentrations into lithium heparin containing-blood tubes (six mLs) at
12 each time-point, protected from light and immediately inverted several times and
13 then kept on ice or refrigerated until centrifugation. Within 30 minutes of blood
14 collection, each blood sample was centrifuged for 10 minutes at 1200 g at 4°C.

15 Plasma was then aliquoted equally into three 2.0 mL tubes (light protected) and
16 stored at -20°C. Samples were shipped on dry ice to the Liverpool Bioanalytical
17 Facility for analysis.

18 Plasma drug concentrations were determined using protein precipitation of
19 analyte and stable isotope labelled internal standard using validated high-pressure
20 liquid chromatography tandem mass spectrometry methods, as previously
21 described [13,14]. The assay was validated over a calibration range of 15-15000

1 ng/mL (darunavir), 5-5000 ng/mL (ritonavir) and 0.5-400 ng/mL (rilpivirine). The
2 accuracy (percentage bias) and precision (% coefficient of variation) were less than
3 15%.

4

5 **5.2.4 DATA ANALYSIS**

6

7 All HIV-RNA for the studied subjects were transformed to the Log10 scale and a
8 linear mixed model fitted, with the trend estimated using a cubic spline.
9 Furthermore, the mean of the decreases in Log10 viral load between successive
10 visits have been compared for group A and group B at each visit time-point using a
11 two-sample randomization t-test (R Foundation for Statistical Computing, Vienna,
12 Austria).

13 The calculated pharmacokinetic parameters for rilpivirine, darunavir and ritonavir
14 were the plasma concentration measured 24 hours after the observed dose
15 (C_{trough}), the maximum observed plasma concentration (C_{max}) and the area under
16 the plasma concentration curve from 0 to 24 hours (AUC₀₋₂₄). All pharmacokinetic
17 parameters were calculated using actual blood sampling time and non-
18 compartmental modelling techniques (WinNonlin Phoenix, version 6.1; Pharsight
19 Corp., Mountain View, CA).

20 Descriptive statistics, including geometric mean (GM) and 95% confidence
21 intervals (95% CI) were calculated for all pharmacokinetic parameters. Inter

1 individual variability in drug pharmacokinetic parameters was expressed as a
2 percentage coefficient of variation [CV, (standard deviation/mean) x 100].

3

4 5.3 RESULTS

5

6 Fifty-two PLWH and naïve to cART were screened for the study and 37 were
7 enrolled. One subject withdrew for personal reasons, therefore 36 completed the
8 study; baseline characteristics are summarised in Table 3. Thirty-five were males and
9 33 were Caucasian (two Indians and one Pakistani); median (range) age was 35 (21-
10 58) years. Baseline median (range) CD4 count was 388 (170-1375) cells/mm³. Pre-
11 cART resistance testing showed that no patient had any resistance to NNRTIs or
12 protease inhibitors, while four had baseline resistance to NRTIs expressed as a single
13 thymidine analogue mutation (TAM). Subjects with viral genotypes which included
14 TAMs were re-questioned to confirm that their medical history did not include the
15 use of antiretroviral medication as pre-exposure or post-exposure prophylaxis. All
16 confirmed the absence of exposure to therapy whilst infected, and the TAM were
17 assumed to have been transmitted at the time of infection.

Table 3 Baseline demographic and clinical characteristics of the study participants

	Group A (n=18)	Group B (n = 18)	All subjects (n = 36)
Gender (male:female)	17:1	18:0	35:1
Age in years (median [range])	36 [21 – 58]	35 [21 – 54]	34 [21 – 58]
Mode of transmission			
Men who have sex with men	16	17	33
Heterosexual	1	-	1
Unknown	1	1	2
HIV RNA (copies/mL), median [range]	91,999 [807 – 5,595,624]	24,403 [807 – 78,113]	215,120 [105,885 – 5,595,624]
HIV subtype			
B ₁ (n)	15	13	28
Non-B ₁ (n)	3	5	8
Non-B subtypes were AG, D, C and AE			
CD4 count (cells/mm ³); median [range]	388 [308 – 404]	450 [204 – 1375]	370 [170 – 787]
N with CD4 <200 cells/mm ³	1	None	1
QT _{cf} (msec); median [range]	360 [308 – 404]	358 [324 – 404]	360 [308 – 404]
Body Mass Index (kg/m ²); median [range]	23 [16 – 35]	22 [20 – 28]	23 [16 – 35]

1

2 **5.3.1 VIRAL LOAD DYNAMICS**

3

4 Eighteen patients had a baseline viral load below 100,000 copies/mL (group A)
5 and 18 with a baseline viral load above 100,000 copies/mL (group B) at screening
6 (Table 1). Viral load decay curves are illustrated in (Figure 5:A, Figure 5:B, Figure
7 5:C).

8 All but one (viral load = 63 copies/mL) study patients achieved viral load < 50
9 copies/mL by week 36 and all by week 48 (Table 1). Overall median (range) viral load
10 reduction (Log10 copies/mL) was 1.3 (0.6-1.9) over the first week of treatment, with
11 no major differences between group A and B (Table 5:D).

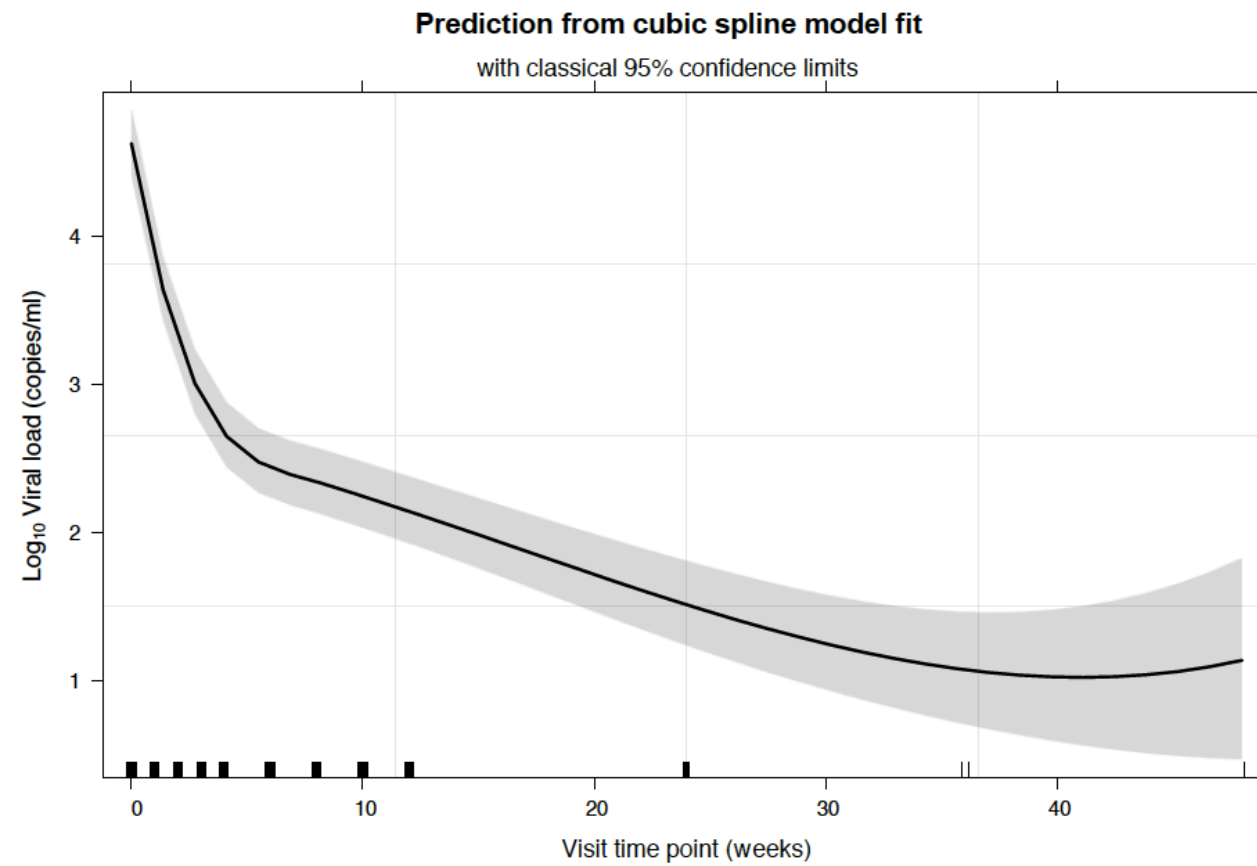


Figure 5: A Geometric mean viral load decay over 48 weeks treatment with rilpivirine/ darunavir/ ritonavir in 36 study patients.

1

2

3

Table 5:D Viral load (VL) decay in the 36 patients who completed the study, and in group A (VL < 100,000 copies/mL) and group B (VL >= 100,000 copies/mL) over the study period.

Between visit Log ₁₀ viral load decrease (copies/ml)	Subgroup of HIV RNA at baseline								
	Overall			U = under 100,000 copies/mL			A= At least 100,000 copies/mL		
	N = 36			N = 18			N = 18		
	n	Median	IQR	n	Median	IQR	n	Median	IQR
Screening to baseline	36	-0.05	-0.2 to 0.2	18	0.12	-0.2 to 0.2	18	-0.14	-0.2 to 0.0
Baseline to wk 1	36	1.30	1.2 to 1.6	18	1.30	1.1 to 1.5	18	1.35	1.2 to 1.7
Wk 1 to wk 2	34	0.36	0.2 to 0.6	17	0.35	0.2 to 0.4	17	0.39	0.2 to 0.7
Wk 2 to wk 3	34	0.17	0.1 to 0.3	17	0.17	0.1 to 0.4	17	0.17	0.1 to 0.3
Wk 3 to wk 4	32	0.14	0.0 to 0.2	15	0.03	-0.1 to 0.2	17	0.19	0.0 to 0.2
Wk 4 to wk 6	31	0.24	0.1 to 0.4	14	0.27	0.0 to 0.5	17	0.24	0.1 to 0.4
Wk 6 to wk 8	30	0.19	0.0 to 0.4	12	0.01	-0.1 to 0.1	18	0.28	0.2 to 0.4
Wk 8 to wk 10	27	0.25	0.0 to 0.4	9	0.25	0.1 to 0.4	18	0.18	0.0 to 0.4
Wk 10 to wk 12	26	0.17	0.0 to 0.3	9	0.16	-0.0 to 0.3	17	0.18	0.0 to 0.5
Wk 12 to wk 24	6	0.73	0.6 to 1.1	0	-	-	6	0.73	0.6 to 1.1
Wk 24 to wk 36	1	0.03	0.0 to 0.0	0	-	-	1	0.03	0.0 to 0.0
Wk 36 to wk 48	0	-	-	0	-	-	0	-	-

4

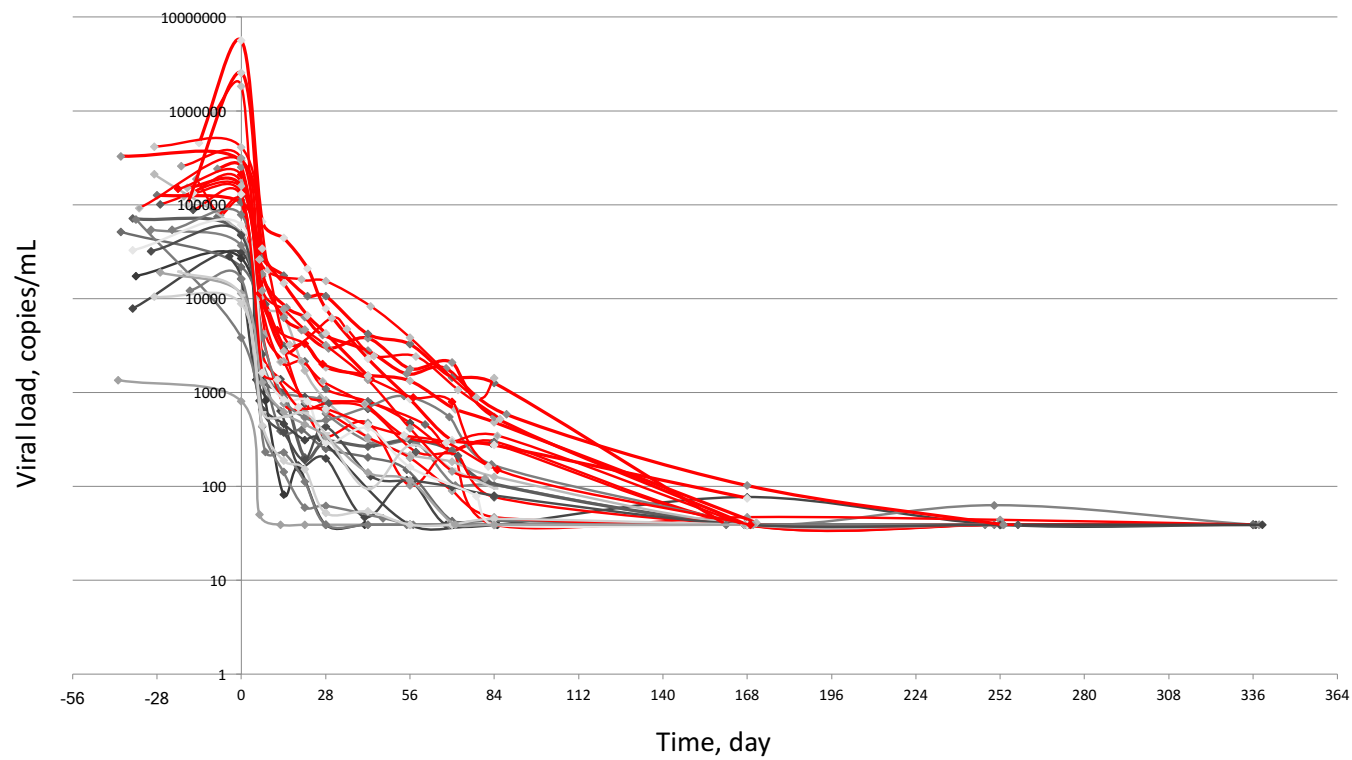


Figure 5:B Individual patient viral load kinetics over 48 weeks treatment with darunavir/ritonavir/rilpivirine (baseline VL over 100,000 copies/mL in red)

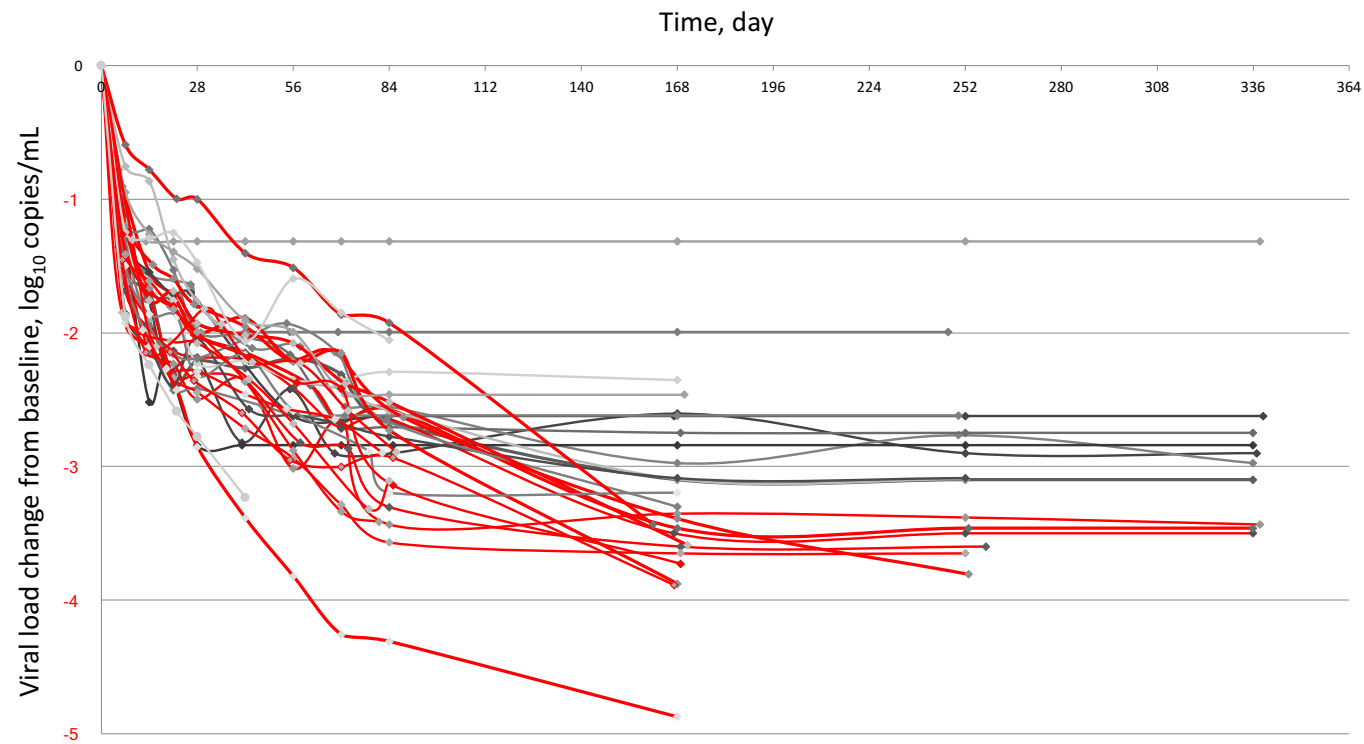


Figure 5:C Individual patient viral load kinetics (delta from baseline0 over 48 weeks treatment with darunavir/ritonavir/rilpivirine (baseline VL over 100,000 copies/mL in red)

1

2 **5.3.2 PHARMACOKINETICS**

3

4 GM plasma concentration versus time curves for rilpivirine, darunavir and
5 ritonavir are shown in (Figure 5:D,Figure 5:E) and their pharmacokinetic parameters
6 summarised in Table 5:E.

7 Although approximately 45% higher, rilpivirine AUC measured in this study was
8 within the range of those reported from Phase III pharmacokinetic sub-
9 studies.(Molina et al. 2012) All subjects had darunavir Ctrough values above the
10 protein binding adjusted IC50 of 550 ng/mL, and the darunavir pharmacokinetic
11 parameters measured were similar to those reported from Phase III studies (Table
12 5:E).(Sherer 2007)

Table 5:E Rilpivirine, darunavir and ritonavir pharmacokinetic parameters over a 24 hour dose interval at steady state in 36 patients living with HIV, expressed as geometric mean and 95% confidence intervals.

PK parameter	Rilpivirine	Darunavir	Ritonavir
N = 36	Geometric mean (95% C.I.)		
AUC_{0-24h}	3036	82598	4455
ng.h/mL	(2876 – 3969)	(76508 – 113143)	(4098 – 64752)
CV%	49	59	68
C_{max}	188	8381	503
ng/mL	(175 – 248)	(7802 – 11279)	(463 – 845)
CV%	53	56	90
C_{trough}	116	1728	42
ng/mL	(106 – 171)	(1661 – 2669)	(38 – 60)
CV%	72	71	68

AUC_{0-24h} ; area under the curve between 0 to 24 hours. C_{max}; maximum concentration. C_{24h}; concentration at 24 hours. CV ; coefficient of variation

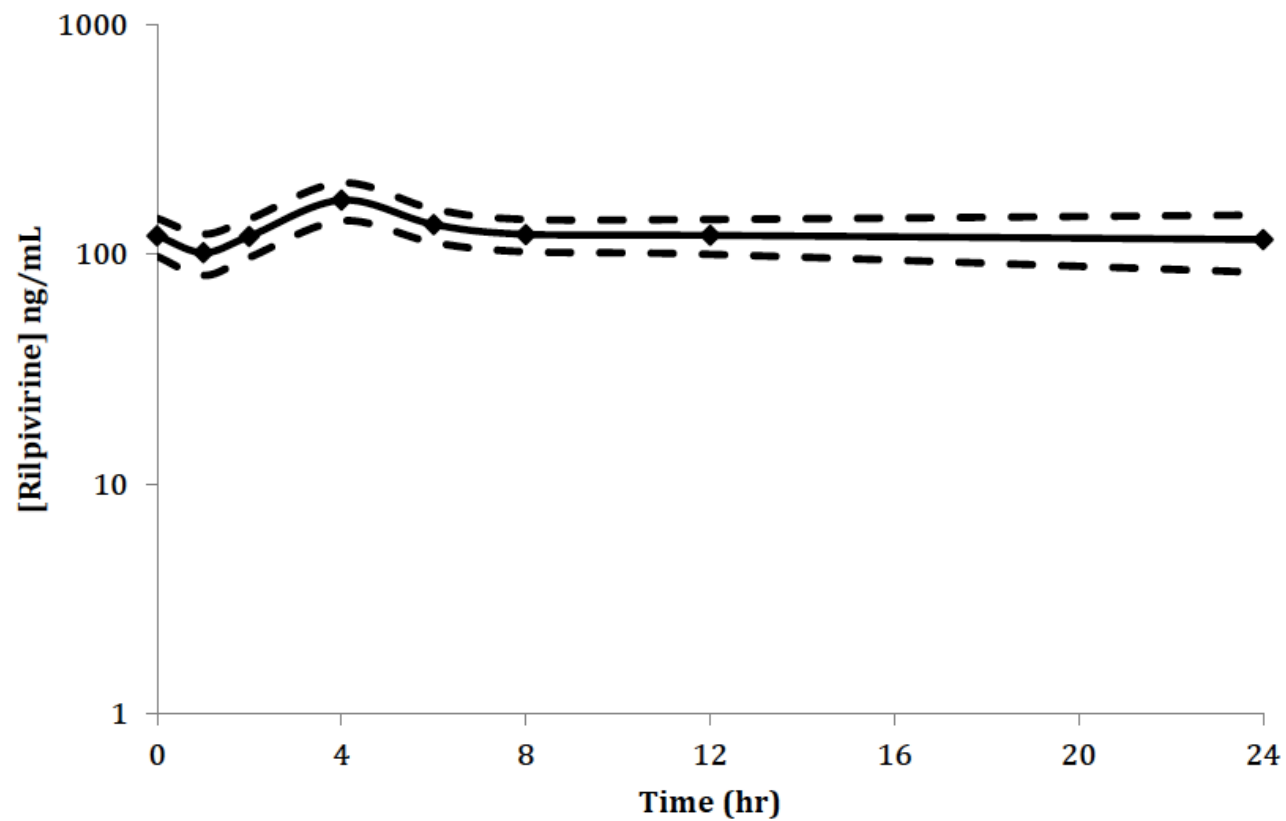


Figure 5:D Geometric mean (GM, solid line) and 95% confidence interval (95%C.I., dotted lines) plasma concentrations of rilpivirine.

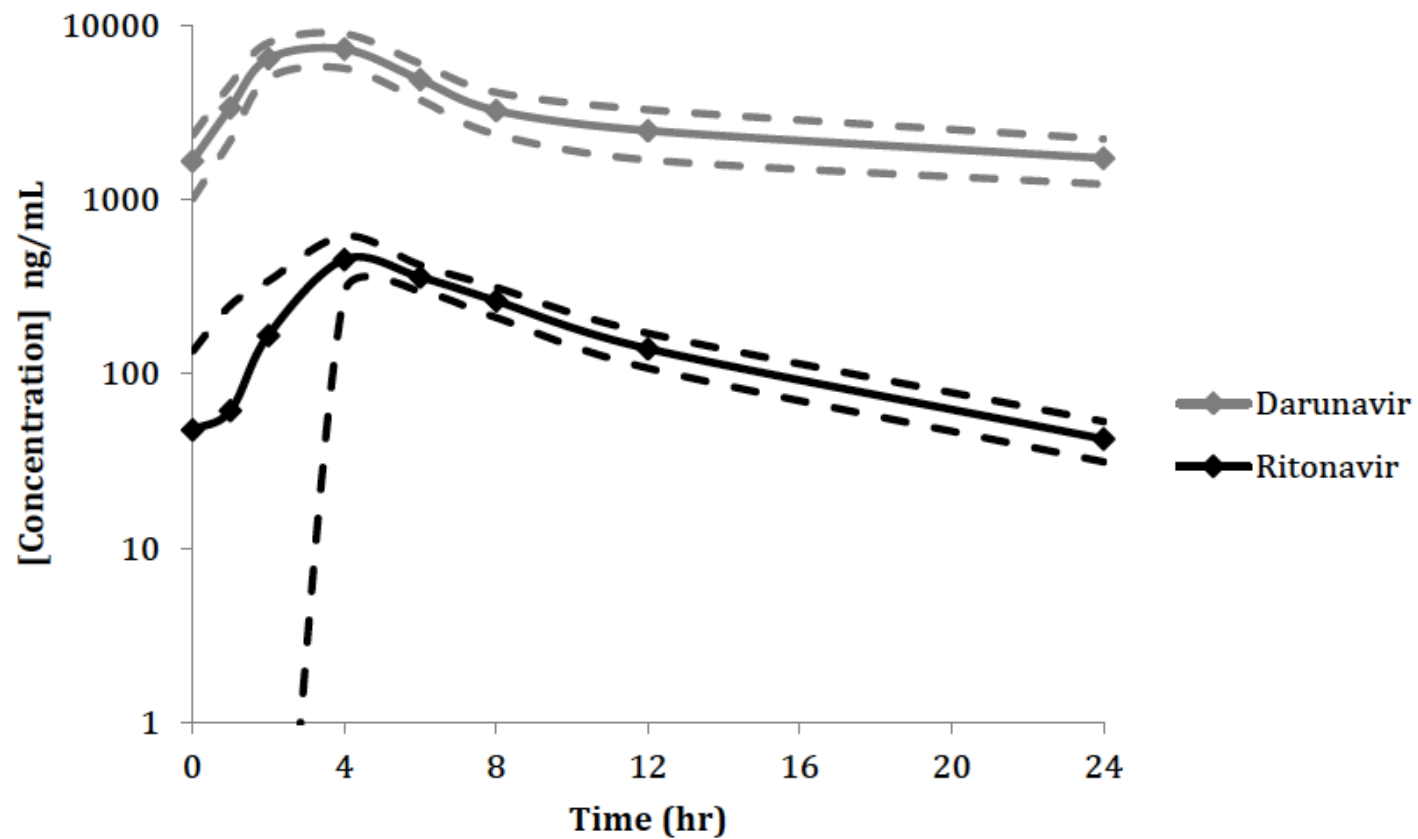


Figure 5:E Geometric mean (GM, solid line) and 95% confidence interval (95% C.I., dotted lines) plasma concentrations of darunavir/ritonavir.

1

2 **5.3.3 SAFETY AND TOLERABILITY**

3

4 Study drugs were well tolerated with no subjects discontinuing rilpivirine and
5 darunavir/ritonavir because of drug related toxicity. Three serious adverse events
6 were recorded during the study, none of which were deemed by the investigator to
7 be related to study medication: i) admission to hospital for treatment of prostatitis
8 complicated by peri-urethral abscess; ii) overnight admission to hospital to treat
9 dehydration and confusion caused by recreational drug intoxication; iii) laparotomy
10 and bowel resection with temporary stoma to repair rectal trauma.

11 There were no other adverse events higher than Grade 2 of severity. Two
12 patients experienced a transient self-limiting generalised rash (without mucosal
13 involvement) following cART initiation, which resolved with continued dosing.

14 No significant change from baseline and no QTcF interval greater than 450 msec
15 were recorded during the study and no laboratory parameter abnormalities of
16 greater severity than Grade 2 were measured.

17

18 **5.4 DISCUSSION**

19

20 In this pharmacokinetic/pharmacodynamic study in HIV-1-infected treatment-
21 naive participants, the NRTI-free combination containing rilpivirine and

1 darunavir/ritonavir led to achievement of an undetectable viral load in all subjects at
2 week 48 (and all but one who had a viral load of 36 copies/mL, by week 36). No
3 differences in viral load decrease or achievement of an undetectable viral load were
4 observed between patients with a baseline viral load above and below 100,000
5 copies/mL.

6 Rilpivirine exposure was slightly increased by ritonavir co-administration via
7 CYP3A4 inhibition; however, its AUC was still within the range observed in historical
8 controls.(Molina et al. 2012) Importantly, no changes in ECG QTcF were seen during
9 the study. The study drugs were well tolerated with no drug related adverse events
10 or laboratory parameters higher than Grade 2.

11 While an association between efficacy and baseline viral load has been
12 demonstrated with other cART combinations containing two or three active
13 drugs,(Sax et al. 2009; Taiwo et al. 2011; Raffi et al. 2014) this small but intensive
14 study showed that rilpivirine plus darunavir/ritonavir is efficacious independently of
15 baseline viral load. A potential explanation of this finding could be the slightly higher
16 concentrations that were measured for rilpivirine in the presence of
17 darunavir/ritonavir or the fact that being a pharmacokinetic/pharmacodynamic
18 study, participants were followed closely and adherence monitored frequently, in
19 particularly during the first four weeks of treatment in which this was conducted
20 weekly.

21 As rilpivirine doses three times higher than the recommended dose of 25 mg
22 once-daily can prolong the QTc interval,(Sanford 2012) and an increase in rilpivirine

1 concentrations was expected because of ritonavir co-administration and therefore
2 inhibition of rilpivirine metabolism. ECG recordings throughout the study were
3 available and showed no change in QTcF from baseline. Therefore, no correlation
4 between QTcF change and rilpivirine C_{max} could be calculated.

5 Dual antiretroviral treatments containing a boosted protease inhibitor-based
6 therapy, (e.g. a boosted protease inhibitor plus one agent from another class) is not
7 the preferred strategy recommended by most recent guidelines. However, over the
8 past 15 years numerous studies have been published showing how dual
9 antiretroviral therapies may be beneficial in clinical practice. Importantly, a recent
10 meta-analysis showed minor differences in terms of virological efficacy when
11 comparing dual to triple cART. (Achhra et al. 2016)

12 Furthermore, it was observed that although a similar risk of serious adverse
13 events in patients on dual and triple therapies, the former had a lower risk of
14 adverse events leading to discontinuation, suggesting that dual therapies may be
15 better tolerated. (Achhra et al. 2016)

16 Interpretation of this study should take into account its single-arm design and the
17 small number of patients studied, and a larger randomized trial is warranted to draw
18 definite conclusions. However, interestingly in the past, single arm Phase IIb trials
19 have predicted more definitive findings than those in Phase III clinical trials: the
20 ACTG A5262 study was followed by NEAT001 that showed that the dual regimen
21 containing raltegravir plus darunavir/ritonavir did not achieve non-inferiority in
22 individuals with high baseline viral loads and low CD4 counts.(Taiwo et al. 2011; Raffi

1 et al. 2014) Beyond this study, the efficacy of RPV/DRV/r in maintaining an
2 undetectable viral load in suppressed patients as a switch strategy and in
3 adolescents has also been confirmed.(Foca et al. 2016; Maggiolo et al. 2016)

4 There is increasing interest from healthcare practitioners involved in
5 management of HIV patients in routine practice, outside academic clinical studies, in
6 the use of reduced number of drugs used in standard antiretroviral regimens. A
7 variety of different innovative combinations, which avoid the use boosted protease
8 inhibitors, are being studied utilising drugs with a high genetic barrier such as
9 dolutegravir, in combination with an NRTI (e.g. lamivudine, 3TC)(Cahn et al. 2017) or
10 an NNRTI (e.g. rilpivirine). {Llibre:2017uh} The outcome of these studies is important
11 because new dual combinations may limit drug-related toxicities in both early and
12 long-term antiretroviral use. Importantly, however, dual regimens need to be
13 carefully selected and to date large randomised clinical trial data on dual therapy
14 are available only for boosted protease inhibitor-containing combinations.(Cahn et
15 al. 2014; Arribas et al. 2015; Cahn et al. 2017)

16 To summarise, this study has demonstrated that the combination of rilpivirine
17 and darunavir/ritonavir shows efficacy in therapy-naïve HIV infected patients and is
18 generally well-tolerated without exhibiting any safety signals of concern. Given this,
19 it is potentially useful as an alternative strategy in patients for whom standard cART
20 is not an option due to resistance or toxicity.

21

1

2 6 SSAT055: NANO-EFAVIRENZ AND NANO-LOPINAVIR

3

4 *AGAJ helped to design this study and wrote the first drafts of the protocol, leading*
5 *on the steps to obtain regulatory and ethical approval. In particular, the candidate*
6 *was responsible for the design of efavirenz-containing arms of this study, by ensuring*
7 *that in accordance with the European Medicines Agency guidance on bioequivalence,*
8 *they would potentially fulfil the minimum requirements necessary for registration*
9 *purposes. The study was delayed for logistic reasons, and resumed only after AGAJ*
10 *had changed employment and was no longer clinically responsible for study conduct.*
11 *However, on resumption of study activities, AGAJ served as a member of the protocol*
12 *steering committee, and conducted the pharmacogenomics analyses on samples*
13 *obtained from volunteers in the study.*

14

15 6.1 BACKGROUND

16 Chapters 3 and 4 of this manuscript, explore the use of a solid drug nanoparticle
17 (SDN) suspension of a non-nucleoside reverse transcriptase inhibitor. This study
18 served as a work of conceptual proof, in determining the potential utility or (and)
19 futility of administering the active pharmaceutical ingredient, rilpivirine, parenterally
20 to an intramuscular site to act as a pre-exposure agent for the prevention of HIV
21 infection.(Jackson et al. 2014; Penrose et al. 2015)

1 The strategic approach of manipulating an existing active pharmaceutical
2 ingredient (API) into a new formulation presents an appealing opportunity. The
3 ability to modify a poorly aqueously soluble drug as nanometer scaled particles
4 presents a potentially advantageous means of modifying the physiological and
5 chemical properties of these drugs and their interaction with either enteral or
6 parenteral absorptive tissues and organs, by enhanced absorption and distribution
7 kinetics. This, whilst at the same time using a lower total amount of API with lower
8 per administration doses, reduced frequency of dosing and potentially lower peak
9 post-absorptive concentrations with prolonged therapeutic coverage.

10 Several nanomedicine strategies can be hypothesised to improve drug delivery
11 and, in general, polymer materials are used in main approaches underpinning the
12 science of nanomedicines. SDN are structured as stabilised particles, with each
13 particle composed entirely of the active drug, unlike the variety of nano-scale
14 formulation approaches which utilise a nano-carrier as a vehicle for the active
15 moiety (polymer micelles, liposomes or vesicles, polymer particles, or drug-polymer
16 conjugates).

17 The advantage of creating SDN using products which have previously been
18 developed through registrational clinical trial programmes, thus achieving full
19 marketing authorisation with well-defined efficacy, tolerability and safety in
20 standard formulation, allows some simplification of the route to a viable licensed
21 product.

1 In the case of rilpivirine (discussed in chapters 3 and 4) this transition and
2 repurposing, by adopting a new mode of delivery from an orally ingested tablet for
3 enteral absorption to a parenteral long-acting intramuscular depot, is not simply a
4 matter of demonstrating bioequivalence between routes of administration. A range
5 of new questions arose during the course of its development. There were physio-
6 chemical factors relating to the stability of the new formulation, its impact on
7 quality of the manufactured product and the resilience of the product real-world
8 use. Additionally, there were patient and healthcare provider considerations of
9 acceptability and reliability of such a product as well as how to manage adverse
10 reactions with a non-reversible product with no current means of removal once
11 administered.

12 Even beyond this, these sets of challenges questions do not cover the range of
13 host-related factors which may differ between the use of an API in established
14 infection, in comparison to how the API must interact with the range of host
15 susceptibility factors from the whole-body systematic level through to local organ
16 and tissue specific innate protective mechanisms at rectal and genital sites and their
17 interaction with local vaginal and gut flora.

18 The suspension-based antiretroviral products which have been studied to date
19 for intramuscular injections (rilpivirine and cabotegravir – an integrase inhibitor
20 being developed by ViiV Healthcare) have typically utilised particles whose average
21 diameter is $<10\mu\text{m}$ which are created through mechanical attrition processes such
22 as nano-milling and high-pressure homogenisation, often termed a “top-down”
23 approach. This is a batched process, which uses macroscopic solid form API as a

1 starting material and within a sterile contained system, gradually reduces the size of
2 the solid by continuously grinding the API against spherical small-diameter ceramic
3 beads over a prolonged period of time. The size and uniformity of the of the desired
4 SDN are time dependent, with greater durations required for smaller particles
5 within a more narrow range of particle size.

6 For example, the G001 formulation of rilpivirine under current development by
7 Janssen is a sterile suspension is based on NanoCrystal® technology, involving a wet
8 bead milling process (NETZSCH Lohnmahltechnik GmbH, Bobingen, Germany) to
9 produce particles of pure rilpivirine of average size 200nm over a milling duration of
10 50 days.

11 A novel alternative to nano-milling for the production of SDN is the “bottom-up”
12 approach using an innovative emulsion-templated spray-dry manufacturing process.
13 (Zhang et al. 2008; Grant and Zhang 2011) In this bottom-up approach, the drug
14 nanoparticles are formed from molecules in an emulsion. The emulsion is freeze-
15 dried by spray freezing into cold liquid to prepare a dry SDN material which can
16 either be readily re-suspended in aqueous medium, forming a nano-particulate
17 suspension, or packaged into capsules for oral administration. To date, a series of
18 candidate SDN antiretrovirals have been generated at the University of Liverpool,
19 formerly in collaboration with IOTA NanoSolutions Ltd (Liverpool, UK) and supported
20 by the Research Councils UK and Engineering and Physical Sciences Research Council
21 funding.

1 This study will explore the pharmacokinetic exposures obtained in HIV-negative
2 healthy volunteers, with oral nano-formulated versions of two existing approved
3 antiretroviral agents (efavirenz and lopinavir) which have been synthesised to GMP
4 standards using the Liverpool technology. These will be compared to the exposures
5 obtained with the existing marketed formulations (Sustiva® - efavirenz and Kaletra®
6 - lopinavir/ritonavir) with the ultimate aim of determining a dose of each
7 nanoformulation which is bioequivalent to the licensed product.

8 For Kaletra® the comparative dose will be the 400/100mg (2x 200mg/50mg film
9 coated tablets) licensed twice-daily dose for treatment of HIV-infected adult
10 patients.

11 For Sustiva®, {BristolMyersSquibbPharmaceuticalsLimited:2002ww} the aim is to
12 study both the licensed dose of 600mg once a day {sustiva smpc ref} and a lower
13 dose of 400mg (2 x 200mg capsules). This lower dose is based on the published data
14 within the ENCORE-1 study (clinicaltrials.gov identifier NCT01011413), , which
15 showed matched effectiveness of the daily 400mg dose to 600mg of efavirenz,
16 when dosed daily with tenofovir disoproxil fumarate/ emtricitabine 245mg/200mg
17 once-daily fixed dose combination.

18

1

2

6.2 METHODS

3

4 This study is a phase I, open-label prospective pharmacokinetic study
5 investigating two new antiretroviral formulations - NANO-efavirenz (NANO-EFV) and
6 NANO-lopinavir (NANO-LPV) in parallel and employing an adaptive design with two
7 stages. The results obtained in the primary stage are used to inform the NANO-
8 antiretroviral doses selected for investigation in the secondary stage.

9 The primary objective of the first phase was to investigate the pharmacokinetics
10 of NANO-EFV and NANO-LPV in HIV-negative healthy volunteers after receiving a
11 single dose and after multiple dosing at steady-state.

12 6.2.1 PARTICIPANTS

13 The study was conducted within clinical research ward facility of the St Stephen's
14 Centre, Chelsea and Westminster Hospital in London, UK. Written informed consent
15 was obtained from male and non-pregnant, non-lactating female healthy volunteers
16 recruited through a database of volunteers and NRES-approved advertisement
17 within the local Trust. Volunteers were aged between 18 to 65 years, inclusive with
18 a recorded body mass index (BMI) from 18.0 to 30.0 kg/m². Participants were
19 screened by medical history (confirmed with their GP), physical examination and
20 screening laboratory assessments and were excluded if they had any significant
21 acute or chronic psychiatric or medical illness, any prescribed or over-the-counter
22 use of medication, electrocardiographic anomalies or QTcF \geq 450 msec, elevated

1 transaminase (ALT and AST) > 1.25 times the upper limit of normal, positive serology
2 for a bloodborne virus (HIV, Hepatitis B or C), recreational drug use (by history and
3 urine drug screen), any known personal or family history of cardiac disease or
4 sudden cardiac death, participation in a clinical trial of an investigational medicinal
5 product within 3 months prior to screen date, and for females of childbearing
6 potential a positive pregnancy test at any point at or after screening.

7

8 **6.2.2 PHARMACOGENOMICS**

9

10 For both stages, in all groups and all arms, volunteers were invited to consent to
11 a separate sub-study in which a single plasma sample was collected at any point on
12 or after day 1, de-identified through a linked anonymised protocol and stored and
13 shipped to the University of Liverpool for later analysis for gene polymorphisms
14 relevant to the metabolic disposal of the studied drug.

15

16 **6.2.3 BIOANALYSIS (DRUG PLASMA CONCENTRATION MEASUREMENT)**

17

18 Blood samples were collected for the measurement of efavirenz, lopinavir and
19 ritonavir concentrations from an indwelling catheter, or direct venepuncture if not
20 available, at each sampling time. Six mLs of whole blood was collected into an
21 evacuated lithium heparin blood collection tube at each timepoint, immediately

1 inverted several times to mix and placed on wet ice or refrigerated until
2 centrifugation, within 90 minutes of collection. Each sample was spun for 10
3 minutes at 2000 g at 4 °C and then equal aliquots transferred to three opaque
4 polypropylene capped storage tubes for storage at -20°C or lower until shipping to
5 the pharmacology research laboratories at Liverpool. Concentrations of efavirenz,
6 lopinavir and ritonavir in plasma were measured using a validated HPLC – tandem
7 mass spectrometry method. (Else et al. 2010; Amara et al. 2011) with a lower limit
8 of quantification of 25 ng/mL for efavirenz and 75 ng/mL for lopinavir.

9

10 **6.2.4 BIOANALYSIS (PHARMACOGENETIC ANALYSIS)**

11

12 Whole blood was collected in a 4mL evacuated EDTA collection tube from those
13 participants who gave written informed consent to the substudy. Samples were
14 deidentified with a secondary unlinked code and the sample was stored at 20°C
15 until shipping to the pharmacology research laboratories at Liverpool. Genomic DNA
16 was extracted from a 250µL aliquot of thawed whole blood whole blood through
17 use of the manufacturers protocol (E.Z.N.A Blood DNA Mini Kit; Omega biotek; Nor-
18 cross, GA). Extracted DNA was quantified using NanoDrop (Thermo Fisher Scientific,
19 Wilmington, DE). Genotyping was completed using real- time allelic discrimination
20 polymerase chain reaction (PCR) assay on a DNA Engine Chromo4 system (Bio-Rad
21 Laboratories, Hercules, CA). The PCR protocol followed denaturation at 95°C for 10
22 min, followed by 50 cycles of amplification at 92°C for 15 sec and annealing at 60°C

1 for 1 min 30 sec. Taqman Genotyping Master mix and assays were purchased from
 2 Life Technologies (Paisley, Renfrewshire, UK). Opticon Monitor v. 3.1 software (Bio-
 3 Rad Laboratories) was used to obtain allelic discrimination plots and identify
 4 genotypes.

5

6 *Table 6:A Pharmacogenetic single nucleotide polymorphisms assayed for*
 7 *participants in each treatment group.*

Efavirenz	Lopinavir
CYP2B6 516G>T (rs3745274)	CYP3A4*22 99366316G>A (rs35599367)
CYP2B6 983T>C (rs28399499)	CYP3A5*3 6986A>G (rs776746)
SLCO1B1 521T>C (rs4149056)	SLCO1B1 521T>C (rs4149056)

8

9 Possession of three homozygous wild-type CYP2B6 15582C>T/ 516G>T/ 983T>C
 10 (CC/GG/TT) is predictive of EFV C₂₄ in the lowest concentration stratum in a
 11 genome-wide association PK correlation study of ACTG trials. (Holzinger et al. 2012)

12

13 **6.2.5 STUDY ENROLMENT**

14

1 Screening procedures were common to both arms and both stages of the study,
2 which were conducted in parallel; firstly, to group B investigating NANO-LPV, then to
3 group A, the NANO-EFV cohort in the primary stages.

4 After analysis of the primary results by a protocol steering committee.

5

6.2.6 (NANO-EFV) GROUP A: PRIMARY STAGE PHARMACOKINETIC PROTOCOL

Three to five volunteers were to receive 50mg of NANO-EFV orally once daily over a 21-day period, with a witnessed dose on day 1 followed by a 72-hour PK profile (pre-dose within ten minutes of dosing, 2, 3, 4, 6, 8, 12, 18, 24, 36, 60 and 72 hours post-dose). Daily dosing from day 4-21 followed, with a second PK profile at steady state following the decay in plasma concentrations over 10 days (pre-dose within ten minutes of dosing, 2, 3, 4, 6, 8, 12, 18, 24, 36, 60, 72, 84, 132, 180 and 228 hours post last dose). In addition, during daily dosing a blood sample for PK trough analysis was taken on days 7, 14 and 17.

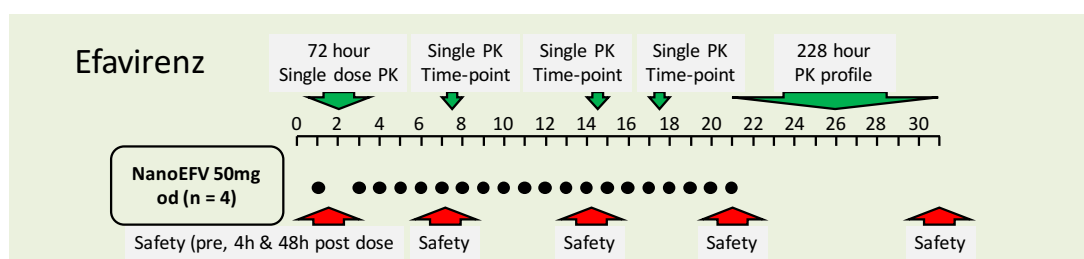


Figure 6:A Efavirenz protocol schedule of events

Efavirenz side effects related to the central nervous system were assessed by the volunteers completing a short 10-item questionnaire, grading symptoms on a four-point ordinal scale from none to severe.

1 **6.2.7 (NANO-LPV) GROUP B PRIMARY STAGE: PHARMACOKINETIC PROTOCOL**

2

3 Three to five volunteers were to receive 400mg of NANO-LPV orally twice daily
4 over a 7-day period, with a witnessed dose on day 1 followed by a 12-hour PK profile
5 (pre-dose within ten minutes of dosing, 1, 2, 4, 8, 12 hours post-dose). Twice daily
6 dosing from day 1-7 followed, with a second PK profile at steady state following the
7 decay in plasma concentrations over 3 days (pre-dose within ten minutes of dosing,
8 1, 2, 4, 8, 12, 24, 32, 48 and 56 hours post last dose). In addition, during twice-daily
9 dosing a blood sample for PK trough analysis was taken on day 3.

10 From days 8 to 21 participants took no medication in a washout period.

11 On day 22, volunteers received 200mg of NANO-LPV twice daily with 100mg
12 ritonavir (Norvir®) twice daily over a 7-day period, with a witnessed dose on day 22
13 followed by a 12-hour PK profile (pre-dose within ten minutes of dosing, 1, 2, 3, 4, 8,
14 12 hours post-dose). Twice daily dosing from day 22-28 followed, with a second PK
15 profile at steady state following the decay in plasma concentrations over 3 days
16 (pre-dose within ten minutes of dosing, 1, 2, 3, 4, 8, 10, 12, 24, 32, 48 and 56 hours
17 post last dose). In addition, during twice-daily dosing a blood sample for PK trough
18 analysis was taken on day 24.

19

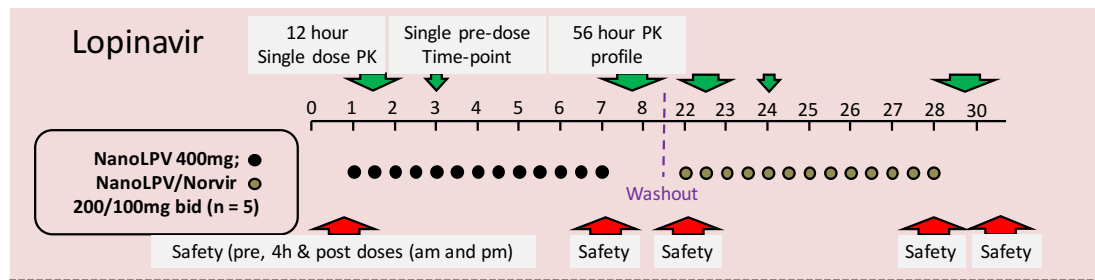


Figure 6:B Lopinavir protocol schedule of events

6.3 PRIMARY STAGE RESULTS

At the time of this manuscript the SSAT055 remains an active ongoing clinical trial of investigational medicinal products. Both primary phase arms and the secondary phase NANO-LPV exploration have been completed, however the secondary phase efavirenz study has not yet been initiated.

Pharmacokinetic analyses and pharmacodynamic analyses have been conducted on samples from completed stages, however the formal GCLP compliant pharmacokinetic report is not yet available at the time of submission of this thesis. Similarly, data on individual participant's clinical history and safety had not undergone final reconciliation, quality checks and database lock, prior to clinical study report being compiled.

The results presented henceforth represent a compilation of the aggregated raw data and simple pharmacokinetic profiles.

6.3.1 GROUP A PRIMARY STAGE (NANO-EFV)

Four participants were enrolled to the primary stage of Group A and completed both single dose and steady state pharmacokinetic profiles having taken NANO-EFV 50mg once daily for 21 days.

1 After the first dose: maximum plasma efavirenz concentrations C_{\max} geometric
2 mean (GM) with 90% confidence intervals was 155 ng/mL (98 – 211 ng/mL)
3 measured at the 3 hour timepoint. At 24 hours, C_{24h} GM(90% CI) was 41ng/mL (29 –
4 52 ng/mL) and after 72 hours C_{72h} had declined to 25ng/mL, the limit of
5 quantification.

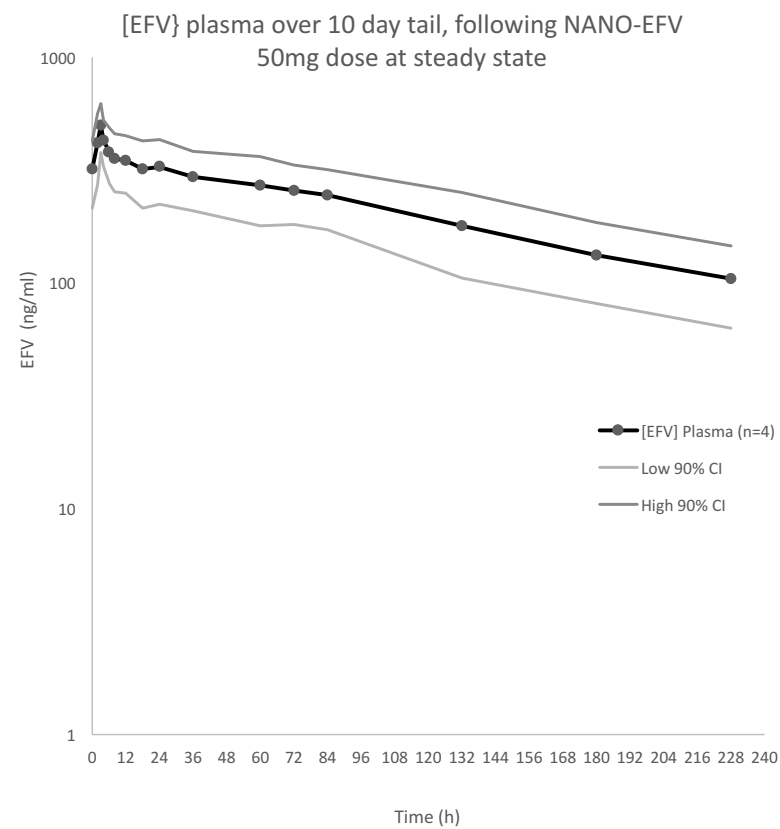
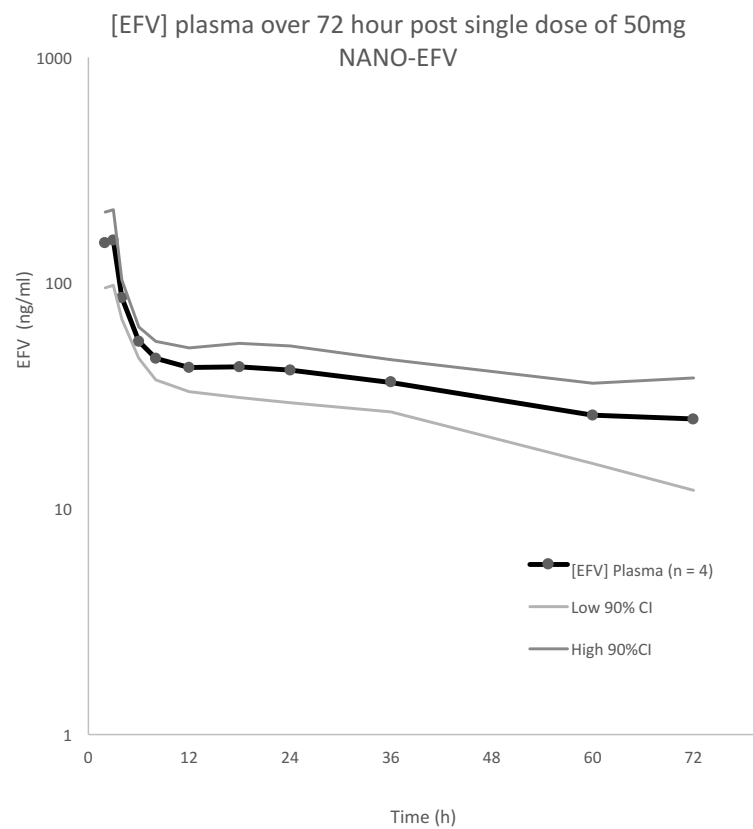
6 At steady state, GM (90%CI) C_{\max} efavirenz was again measured at 3 hours after
7 dose at 501ng/mL (379 – 624 ng/mL) declining to 328 ng/mL (223 – 432 ng/mL) and
8 105 ng/mL (63 – 146 ng/mL) at C_{24h} and C_{228h} , respectively. Geometric mean PK
9 profiles and 90% confidence intervals are illustrated in Figure 6:C

10 *Table 6:B NANO-Efavirenz; single nucleotide polymorphisms in substudy*
11 *participants*

EFV patients (n=4)	<u>Genotype frequencies</u>		
CYP2B6 516G>T (rs3745274)	GG	GT	TT
	2	0	2
CYP2B6 983T>C (rs28399499)	TT	CT	CC
	4	0	0
SLCO1B1 521T>C (rs4149056)	TT	CT	CC
	4	0	0

12

- 1 Single nucleotide genetic polymorphisms in CYP2B6 516G>T, CYP2B6 983T>C and
- 2 SLC1B1 521T>C results are presented in Table 6:A but have not been analysed as
- 3 covariates due to small numbers in the primary stage.



1

2

Figure 6:C Primary stage NANO-EFV, single dose and steady state PK profiles, Geometric mean (90% confidence intervals)

1 6.3.2 GROUP B PRIMARY STAGE (NANO-LPV)

2

3 Five participants were enrolled to the primary stage of Group B and completed
4 both 7 day dosing periods of 400mg NANO-LPV twice daily and after washout
5 200mg NANO-LPV + 100mg ritonavir twice daily, with 12 hour PK profile after single
6 dose and 56-hour PK profile following the final dose on day 7 at steady state.

7

8 Geometric mean PK profiles and 90% confidence intervals for plasma lopinavir
9 are displayed in Figure 6:D.

10 *Single dose:*

11 GM (90% CI) lopinavir concentrations peaked at 2 hours following a 400mg oral
12 dose of NANO-LPV; C_{max} 111ng/mL (61-162) and were below LLQ in all participants
13 at 8 hours post dose. Conversely after 200mg NANO-LPV with 100mg ritonavir, C_{max}
14 occurred at 4 hours post dose; 976 ng/mL with high variability 90% CI (-1105 to
15 3057 ng/mL), declining to 645 ng/mL (-444 to 1734 ng/mL) at 12 hours post-dose.

16 *Steady State*

17 After 7 days dosing with 400mg NANO-LPV twice daily, lopinavir C_{max} {GM, (90%
18 CI)} was 93 ng/mL (23 – 164 ng/mL) at 2-hour post dose, with all below LLQ at 12
19 hours post dose.

1 Conversely, when NANO-LPV 200mg was taken twice daily with a dose of 100mg
2 ritonavir, C_{max} was recorded 4 hours post dose at 7002 ng/mL (5263 – 8741 ng/mL),
3 with C_{12h} , and C_{24h} were 3068 ng/mL (1307 – 4828 ng/mL) and 580 ng/mL (-84 to
4 1243 ng/mL), respectively and all concentrations were below the limits of
5 quantification by 48 hour post-dose.

6 ***Ritonavir***

7 Plasma ritonavir PK profiles after single dose and steady state in the ritonavir
8 periods are shown in Figure 6:E.

9

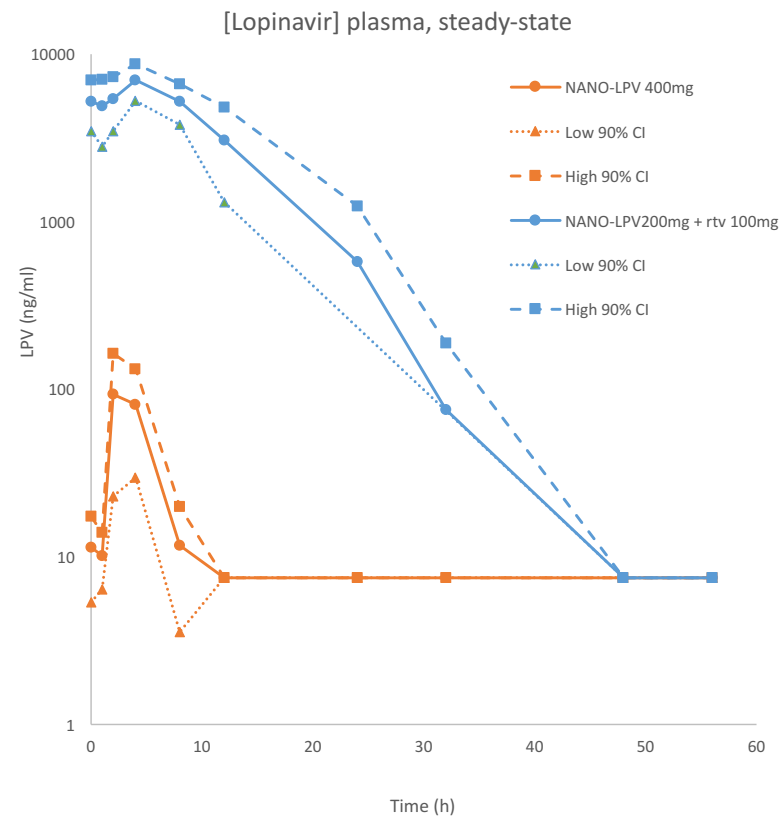
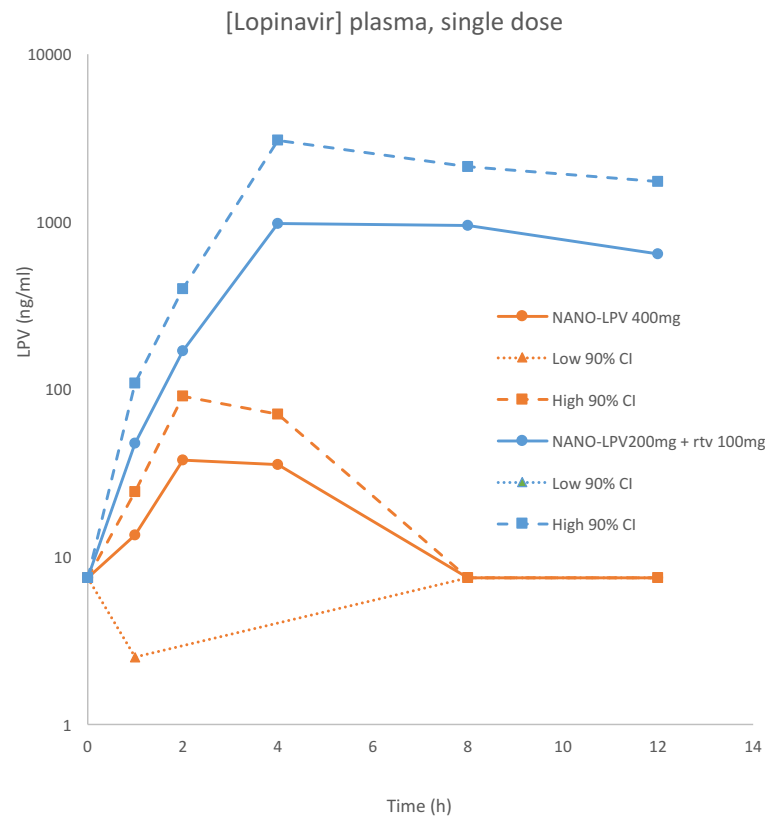


Figure 6:D Primary Stage Group B; Lopinavir plasma PK profiles with NANO-LPV +/- rtv at single dose and steady state, geometric mean (90% confidence intervals)

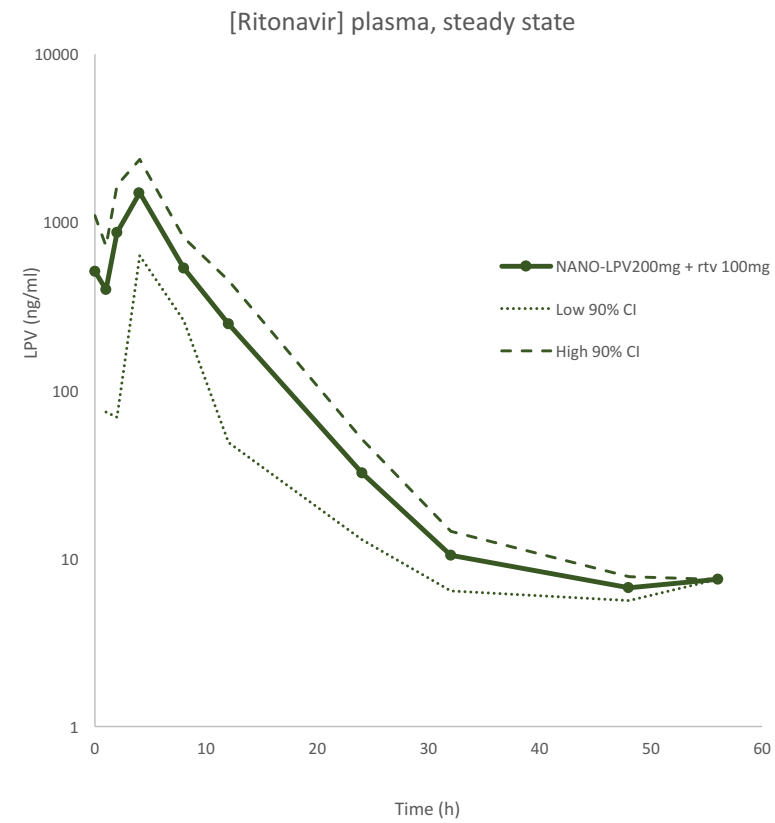
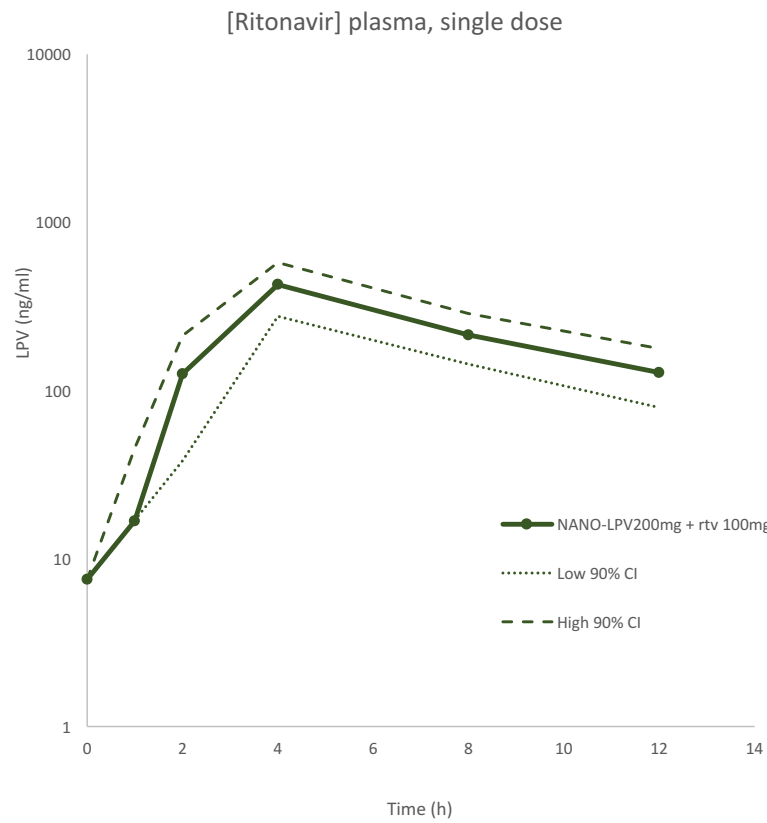


Figure 6:E Primary Stage Group B; Ritonavir plasma PK profile when dosed with NANO-LPV at single dose and steady state, geometric mean (90% confidence intervals)

6.4 MODELLING THE PRIMARY STAGE RESULTS TO INFORM PROTOCOL STEERING COMMITTEE

Nonlinear mixed-effects modeling was applied as described previously by Dickinson et al, (Dickinson et al. 2011) was developed to explore the relationship between lopinavir and ritonavir concentrations over 72 h after attaining steady state with NANO-LPV without or with ritonavir, in order to assess other lopinavir and ritonavir dosing strategies compared to the standard 400-mg–100-mg twice-daily dose.

The modelling methodology conducted by Dr. Dickinson is briefly described below.

Nonlinear mixed-effects modeling was applied by using NONMEM (version VI 2.0, level 1.1, double precision; ICON Development Solutions, Ellicott City, MD) (3) with first-order conditional estimation with interaction (FOCE-I). The model fit was assessed by statistical and graphical methods. The minimal objective function value (OFV) (equal to a $-2 \log$ likelihood) was used as a goodness-of-fit diagnostic, with a decrease of 3.84 points corresponding to a statistically significant difference between nested models ($P < 0.05$, χ^2 distribution, and 1 degree of freedom).

Graphical diagnostics were performed with Microsoft Office Excel 2007 for Windows (Microsoft Corporation, Redmond, WA). Standard errors of the parameter estimates were determined with the COVARIANCE option of NONMEM, and individual Bayesian parameter and concentration estimates were determined with

1 the POSTHOC option. The model-building process was in 3 stages: (i) a separate
2 model was developed for lopinavir, (ii) a separate model was developed for
3 ritonavir, and, finally, (iii) a combined model was developed, incorporating the
4 influence of ritonavir concentrations on lopinavir clearance

5 The Population PK method was used to describe the clinical data and to simulate
6 alternative dosing strategies in order to inform PSC selection of

7

8 1. Kaletra® (LPV/RTV, 400/100 mg bid; n=16) LPV and RTV modelled using a
9 sequential approach with a direct response E_{\max} model (Dickinson et al. 2011)

10

11

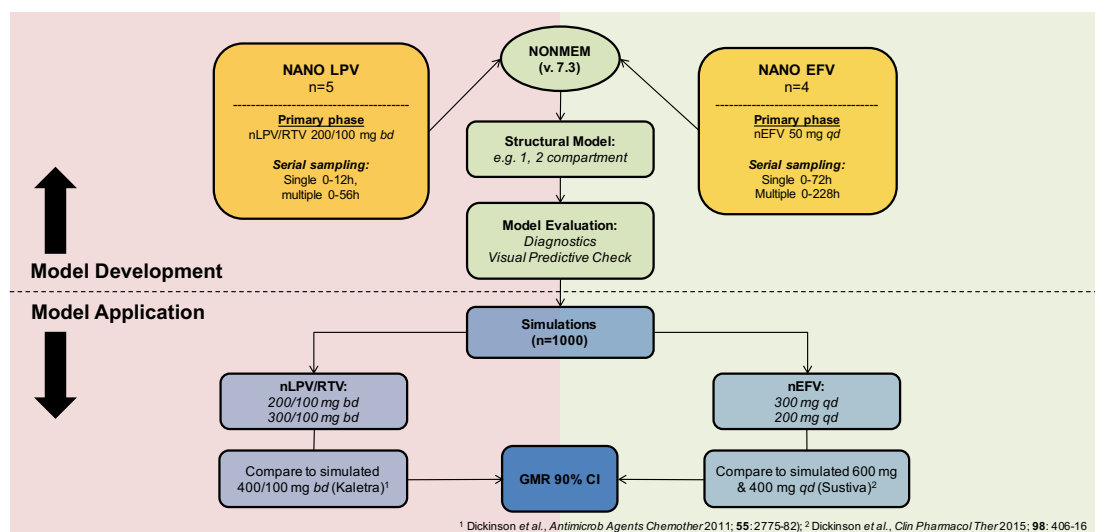
12 2. NanoLPV/RTV (nLPV/RTV, 200/100 mg bid; n=5) nLPV and RTV modelled
13 simultaneously with an indirect response E_{\max} model (enzyme turnover; using
14 only the multiple dose nLPV/RTV data)

15

1 Simulations:

- 2 1. LPV/RTV 400/100 mg bid, n=1000
- 3 2. nLPV/rtv 200/100 mg bid, n=1000
- 4 3. nLPV/rtv 300/100 mg bid, n=1000

5 Steady state was not assumed, but the model dosed with the same number and
6 frequency of doses as the clinical study



8 A summary of the comparative simulation dosing with ritonavir boosted NANO-
9 LPV 200/100mg compared to full-dose Kaletra and NANO-LPV/rtv 300/100mg is
10 shown in Table 6:C. Comparing the 12-hour areas under the curve, 200/100mg
11 NANO-LPV/rtv was predicted from the simulation to give comparable PK exposure to
12 that of standard dose Kaletra, whilst the higher 300/100mg NANO-LPV/rtv

Table 6:C Population PK simulation output comparing dosing strategies, based on clinical data output from primary stage

Comparison of Kaletra vs. nLPV/RTV

Comparison of simulated C_{12} , C_{24} , AUC_{0-12} , AUC_{0-24} , and C_{max} made by means of GMR, 90% CI (difference considered significant if the CI does not cross 1 – shown in bold)

	Geometric mean			GMR (90% CI)*	GMR (90% CI)*
LOPINAVIR	<i>nLPV 200 mg</i>	<i>nLPV 300 mg</i>	<i>LPV 400 mg</i>	<i>nLPV 200 vs. LPV</i>	<i>nLPV 300 vs. LPV</i>
C_{12} (mg/L)	4.16	6.24	4.02	1.04 (0.99-1.08)	1.55 (1.49-1.62)
C_{24} (mg/L)	0.66	0.99	0.24	2.77 (2.53-3.04)	4.16 (3.80-4.56)
AUC_{0-12} (mg.h/L)	72.35	108.53	79.07	0.92 (0.89-0.94)	1.37 (1.34 -1.41)
AUC_{0-24} (mg.h/L)	101.64	152.46	100.76	1.01 (0.98-1.04)	1.51 (1.47-1.56)
C_{max} (mg/L)	10.69	16.04	9.97	1.07 (1.05-1.10)	1.61 (1.57-1.65)

	Geometric mean		GMR (90% CI)*
RITONAVIR	<i>RTV 100 mg (nano)</i>	<i>RTV 100 mg</i>	
C_{12} (mg/L)	0.214	0.123	1.74 (1.57-1.92)
C_{24} (mg/L)	0.017	0.013	1.31 (1.06-1.61)
AUC_{0-12} (mg.h/L)	9.46	3.59	2.64 (2.54-2.74)
AUC_{0-24} (mg.h/L)	11.85	4.33	2.74 (2.62-2.88)
C_{max} (mg/L)	2.08	0.59	3.55 (3.43-3.67)

* Nanoformulation as reference

NANO-EFAVIRENZ

Simulated doses of NANO-EFV 300 mg or 200 mg once daily were predicted

approximate simulated doses of EFV (Sustiva) 600 or 400 mg once daily.

{Humanconfirmationo:2017wf}

6.4.1 PROTOCOL STEERING COMMITTEE

Both primary stages completed with no adverse events above grade 2, no

discontinuation of therapy and no serious adverse events.

1 The Protocol Steering Committee consisted of the clinical investigators, members
2 of the antiretroviral pharmacokinetic group and department of chemistry at the
3 University of Liverpool, and an academic clinician and a biostatistician who were
4 both extensively experienced in the clinical development studies of antiretroviral
5 therapies. Guided by the results of the population PK simulation, the protocol
6 steering committee agreed to proceed to the secondary stage investigation of
7 NANO-LPV/rtv with the 200/100mg BD dose, comparing in two-phase randomised-
8 order sequential dosing period with washout, against Kaletra® 400/100mg BD.

9

6.5 SECONDARY STAGE: NANO-LOPINAVIR

Ten volunteers were randomly allocated to Arm 1 or Arm 2 in a 1:1 allocation to determine the order in which they received the investigational product and nano-formulation.

The protocol for study conduct of the secondary stage was as described in section 6.2.7 above, with the minor alteration to add PK timepoints at 3 hour and 10 hour post-dose to better characterise the timing and magnitude of peak concentration.

Arm 1: Five volunteers received lopinavir/ritonavir as Kaletra® tablets at a dose of 400/100mg twice daily for 7 days, then following a 2-week washout period, received 200mg NANO-lopinavir and 100mg ritonavir (Norvir®) twice daily for 7 days.

Arm 2: Five volunteers received 200mg NANO-LPV and 100mg ritonavir (Norvir®) twice daily for 7 days, then following a 2-week washout period, received lopinavir/ritonavir as Kaletra® tablets at a dose of 400/100mg twice daily for 7 days

1 6.5.1 RESULTS

2

3 *Steady State*

4 After 7 days dosing with Kaletra® 400/100mg BD and NANO-LPV/rtv 200/100mg
5 BD, steady state pharmacokinetic profiles for 56 hours post dose were obtained as
6 plotted in Figure 6:F.

7 With Kaletra®, C_{\max} GM (90% CI) of 9928 ng/mL (7418 – 12438 ng/mL) was
8 attained at 4 hour post dose, with C_{12h} of 4880ng/mL (3392 – 6369ng/mL).

9 In comparison with NANO-LPV/rtv, C_{\max} was 4954 ng/mL (3749 -6160 ng/mL)
10 with C_{12h} at 2458 ng/mL (1807 – 3110 ng/mL).

11 Results of genomic DNA analysis for SNP of relevance to lopinavir metabolism for
12 both stages are presented in Table 6:D. The predominant concordance between
13 participants in genotype and small number of participants precludes further
14 covariate analysis at this point.

15

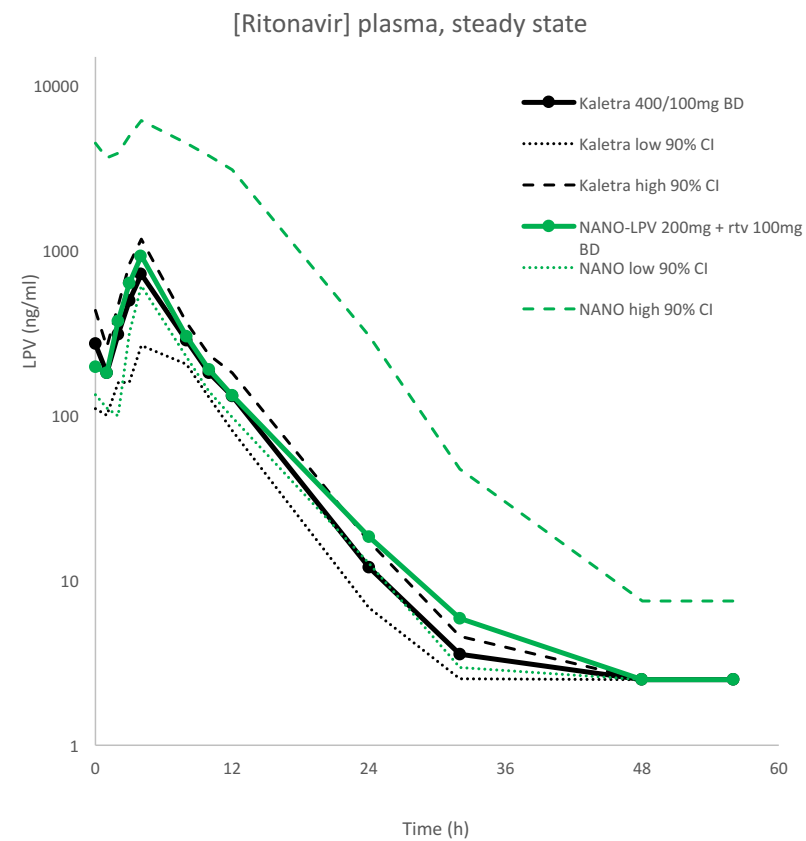
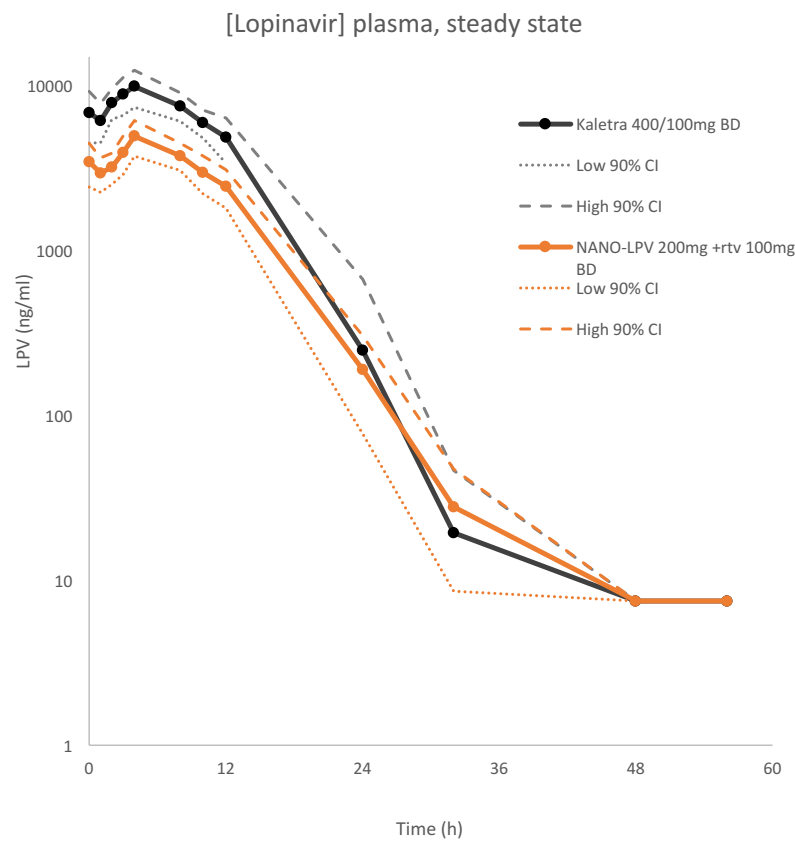


Figure 6:F Secondary Stage: PK profiles (GM, 90% CI) for Kaletra 400/100 BD and NANO-LPV 200mg plus ritonavir 100mg BD, after dosing to steady state for 7 days.

1

Table 6:D NANO-Lopinavir pharmacogenomic results

LPV patients (n=13)	<u>Genotype frequencies</u>		
CYP3A4*22 99366316G>A (rs35599367)	GG	GA	AA
	13	0	0
CYP3A5*3 6986A>G (rs776746)	AA	AG	GG
	7	3	3
SLCO1B1 521T>C (rs4149056)	TT	CT	CC
	12	1	0

2

6.6 DISCUSSION

This preliminary data from an ongoing study pilot study, showed that both orally administered SDN formulations of efavirenz and lopinavir were generally well tolerated at the studied doses in small numbers of HIV-1 negative healthy volunteers. The data so far confirm the potential of a 50% dose reduction compared to the licensed formulations using a novel approach to the manufacture of solid dose nano-formulations.

These results are currently preliminary, and even on completion of this adaptive-design pilot study will have been limited to human studies in a less than two-dozen volunteers requiring further confirmation in larger studies. Regardless, the current European Medicines Agency guidance on investigation of bioequivalence (European Medicines Agency 2010) provides a good indication for a route to regulatory approval and how it could potentially be achieved at the next stage of human study.

The projected statement on the dose-reducing potential of the SDN for efavirenz compared to that of the licensed standard formulation is currently based on population PK modelling, using a common model developed within the same collaborative group to compare with a statistically-powered clinical study in HIV-1 infected patients receiving treatment; the ENCORE1 study. (Dickinson et al. 2016) This study established the non-inferior virologic efficacy of a 400mg dose of efavirenz compared to 600mg, both given with 2 NRTI. Though lower plasma PK exposures were observed with the 400mg dose, wherein the modelled predictions

1 for some participants indicated concentrations at 12 hours after dosing, C_{12h} below
2 the putative minimum effective concentration of 1000 ng/L, no significant
3 difference in the virologic failure rate was observed compared to those above this
4 threshold.

5 On the basis of the ENCORE1 results, the 400mg dose of efavirenz is being
6 considered by global non-governmental bodies for guidance approval with the
7 intended benefit anticipated to be in low and middle-income countries with large
8 populations infected by HIV. Ongoing studies are being conducted to provide
9 evidence that this dose reduction does not have detrimental effects due to lowered
10 therapeutic efficacy in two clinical situations of concern to healthcare in these
11 societies; i) in women of child-bearing age during pregnancy and ii) when co-
12 administered with rifampicin containing regimens for the treatment of tuberculosis.
13 (Boffito et al. 2017)

14 With future confirmation of the bioequivalence of 200mg SDN formulation of
15 efavirenz to standard formulation at 400mg, there are potentially significant savings
16 to be gained both in cost of drug and in the amount of active pharmaceutical
17 ingredient used within the context of a global manufacturing capacity which is not
18 infinite (Table 6:E). If confirmed in larger future studies, the approach has the
19 potential for savings up to 243 million USD per year while also freeing up
20 manufacturing capacity up to 930 tons per year.

1

2 *Table 6:E Predicted cost and product efficiency gains with the use of solid*
3 *dose nanoformulations replacing standard formulations of lopinavir and*
4 *efavirenz.*

SDN (dose)	Comparator drug ¹	Est. cost of SDN pppy (USD) ²	Est. savings in L&MICs during first 5 yrs of launch ³	
			Cost saved (USD)	API saved
Lopinavir / ritonavir (200/100 QD)	400/100 BD.	~\$ 183	~\$ 216 Mn	331 tons
Efavirenz (300mg BD)	600 mg QD	~\$ 24	~\$ 471 Mn	~2,759 tons
	400 mg QD	~\$ 24	~\$ 113 Mn	~920 tons
Efavirenz (200mg BD)	400 mg QD (if approved)	~\$ 16	~\$ 314 Mn	~1,839 tons

5

- 6 1. Dose reduction predicted from the presented pharmacokinetic data
7 2. University of Liverpool estimated SDN process may add \$4-16/kg to API cost, lower-end
8 estimates shown here, updated with improved understanding of yield loss
9 3. High-level estimates based on ARV demand in low and middle income countries (L&MIC)
10 2015-2025 jointly forecast by Medicines Patent Pool & World Health Organisation (2016)
11 pppy = per person, per year

12

13 In a similar dose minimisation proposal to that described for standard
14 formulation efavirenz (Dickinson et al. 2016), this group has previously investigated
15 lopinavir (Jackson et al. 2011) and lamivudine (Else et al. 2012). All three could
16 potentially be candidates for SDN formulation, as the approach has wide
17 applicability to drugs from several classes for numerous indications. In addition to
18 other development programmes for other oral applications, parenteral long-acting
19 SDN applications are currently ongoing.

1 7 SUMMARY

2 In summarising the independent projects from which this thesis is constituted,
3 each was designed with the aim of addressing a clinical question of relevance to
4 current and future practice in the management of treatment and prevention of HIV.
5 Each was successful in achieving its stated objectives, providing positive
6 confirmations of the overriding hypothesis being tested.

7 A common feature of all three pieces of work was the incorporation as an
8 integral feature of the design of the protocols of a feedback decision loop utilising
9 pharmacokinetic methods to adjudicate safety, efficacy and viability of the ongoing
10 study. This is a common feature within pharmaceutical manufacturer's early stage
11 human clinical studies with multi-phase single-ascending dose and multiple-
12 ascending dose structures. In research from academic and healthcare research
13 groups in the UK, this has previously been an uncommon strategy but is being used
14 with increasing frequency with adaptive study designs. These often tend to be post-
15 licensing studies which aim to explore and to define clinical effectiveness of the use
16 of therapies, and may combine therapies to explore their synergistic effects. The
17 potential for improving the efficient use of research funds, physical and human
18 resources as well as the ability to build into safeguards based on target exposures
19 within a therapeutic window, which has the ethical implications of avoiding
20 exposure to futile doses, particularly in healthy volunteer studies. Within this
21 context, the UK Health Research Authority has recognised the current and future
22 trend towards the use of such study designs and this year has introduced specific

1 training on the research ethics pertaining to them, into the curriculum of the
2 National Research Ethics Service training for committee members and chairs.

3

4 **SSAT040** demonstrated that an nano-formulation of the non-nucleoside reverse
5 transcriptase inhibitor, rilpivirine, when dosed intramuscularly was generally well
6 tolerated as a single dose with no safety signals of concern apart from transient
7 local injection site discomfort. Each studied dose provided prolonged plasma
8 exposure for at least four weeks, at levels consistent with therapeutic efficacy. With
9 regards its potential as a pre-exposure prophylaxis agent, the tissue drug
10 penetration within the female genital tract and male rectum was comparable to
11 plasma exposures. Testing these achieved female genital secretory fluid
12 concentrations in an *ex vivo* model showed that viral inhibition could be achieved at
13 these drug levels. On the basis of these study results the McGowan group at the
14 Magee-Womens Research Institute in Pittsburgh, again supported by the same
15 funder and manufacturer, have completed the next phase of study with 600mg and
16 1200mg doses in a protocol designed to specifically test viral inhibition in freshly
17 biopsied rectal and vaginal tissue samples. (McGowan et al. 2016) Interestingly,
18 though significant viral suppression was observed in rectal tissue, persisting for
19 months after a single dose, no viral suppression was detected in cervical or vaginal
20 tissue. This latter finding has had negative implications on the future development
21 decision on long-acting rilpivirine for HIV prevention.

22

1 **SSAT049** as a pilot study, demonstrated the efficacy of this oral once daily
2 combination of darunavir, boosted by ritonavir with rilpivirine for effective viral
3 suppression in small numbers of therapy naïve HIV-infected adults. At the time of
4 creating the study protocol as a concept, the idea of co-formulating these agents to
5 a single-tablet fixed dose combination was discussed in non-specific term with the
6 drug manufacturer. Since that time, evolution of the therapeutic landscape has
7 rendered these ideas moot, and in fact the manufacturer continued with the
8 commercial decision to develop a co-formulation of darunavir boosted by cobicistat
9 as a single-tablet, (Janssen-Cilag Ltd 2014) and very recently has received approval
10 for both these agents combined with a dual-NRTI backbone of tenofovir
11 alafenamide and emtricitabine in Symtuza®.(Janssen-Cilag Ltd 2017)

12

13 **SSAT055**, though not yet complete, has demonstrated the potential of a novel
14 manufacturing process for solid-drug nanoparticle formulations and the potential
15 for oral SDN formulations to provide advantageous pharmacokinetic exposure, with
16 a reduction in the absolute amount of active pharmaceutical ingredient used. Whilst
17 these advantages of dose reduction/ minimisation would be applicable to all global
18 regions, the anticipated greatest benefit - should further development of the
19 process chemistry and manufacturing be proven viable for large scale – is expected
20 to be in low and middle-income countries where the majority of 19 million
21 untreated people living with HIV reside. There has been a significant recent
22 announcement to this regard from the World Health Organisation and the
23 governments of South Africa and Kenya (with multiple global health stakeholders:

1 the Joint United Nations Programme on HIV/AIDS (UNAIDS), the Clinton Health
2 Access Initiative (CHAI), the Bill & Melinda Gates Foundation (BMGF), Unitaid, the
3 United Kingdom's Department for International Development (DFID), the United
4 States President's Emergency Plan for AIDS Relief (PEPFAR), the U.S. Agency for
5 International Development (USAID), and the Global Fund to Fight AIDS, Tuberculosis
6 and Malaria, with Mylan Laboratories Limited and Aurobindo Pharma). A pricing and
7 licensing agreement has been concluded to provide a generic, single tablet regimen
8 containing the integrase inhibitor dolutegravir (with lamivudine and tenofovir
9 disoproxil fumarate) to healthcare purchasers in low and middle-income countries
10 for the price of US\$75 per year {WorldHealthOrganisation:uy}. This is an important
11 advance in the global access to antiretroviral therapies which addresses a major
12 discrepancy between resource-poor and resource-rich countries; that of pricing and
13 availability of guideline recommended first-line options (dolutegravir, efavirenz and
14 nevirapine are the three WHO recommended third agent). The availability of a
15 dolutegravir based single-tablet regimen at an accessible purchase price would
16 appear to make efforts to optimise doses using SDN of the older third-agents,
17 efavirenz and lopinavir/ritonavir, obsolete.

18 There are three counterpoints to this assumption which point to a continued
19 demand for efavirenz based therapy. Firstly, the global prevalence of persons living
20 with HIV are likely to continue to increase, due to a reduction in mortality with
21 successful viral suppression whilst rate of decline in incidence lags behind. To
22 achieve a ninety percent target of patients on therapy, will require significant
23 additional capacity for manufacturing API for both existing products under license

1 agreements, as well as newly introduced products. Secondly, patients currently
2 stable and established on efavirenz based regimens are likely to remain on their
3 current therapy, thus maintaining this need for product and opportunity for its
4 improvement. Finally, the proof-of-concept for oral SDN reformulations will likely
5 apply in just the same way to other antiretroviral regimens.

1 8 ACKNOWLEDGEMENTS

2 The work contained within this thesis manuscript is the product of the
3 collaborative efforts of multiple agencies, whose contributions I wish to
4 acknowledge.

5 Firstly, I wish to thank my clinical supervisor at SSAT, Professor Marta Boffito,
6 whose leadership, tutelage, guidance and encouragement were instrumental in
7 stimulating an interest in clinical pharmacology and both initiating and completing
8 this work.

9 I also thank Professor Saye Khoo and Professor Andrew Owen for their guidance,
10 good humour, patience and ongoing supervision.

11 St. Stephen's AIDS Trust

12 Professors Brian Gazzard and Mark Nelson.

13 All patients and healthy volunteers; both those who participated in this study as
14 well as the many with whom I have had the privilege to play some part in their
15 clinical care, whilst they frequently displayed remarkable acts of altruism bravery in
16 taking the informed risk of participating in research.

17 Chris Higgs, Zeenat Karolia, Natalia Seymour, Lisa Ringner-Nackter, Elisa
18 Bisdomini, Carl Fletcher, Ruth Bateson and the regulatory team, & Gary Lo.

19

1 University of Liverpool.

2 Marco Siccardi, Laura Else, Deidre Egan and the team in the bioanalytic facility,
3 Laura Dickinson and the PK modelling team, fellow doctoral candidate Megan Neary
4 and Justin Chiong.

5 Professor Ian MacGowan at the Magee-Womens Research Institute, Pittsburgh.

6 SSAT040 was supported by a research grant from the Bill and Melinda Gates
7 Foundation. The study medication was supplied by Janssen Infectious Diseases. I
8 also thank Stephen Becker, Kim Shaffer, and Joe Romano from the Bill and Melinda
9 Gates Foundation and Herta Crauwels, René Verloes, and Peter Williams from
10 Janssen Infectious Diseases for their advice during this study. Betsy Herold and
11 Pedro Mesquita at the Albert Einstein College of Medicine who agreed at an early
12 stage to be collaborating co-authors and were critical to the design and analysis of
13 the ex vivo pharmacodynamic aspect of this study.

14 I wish to thank John Mellors, Kerri Jo Penrose, Urvi Parikh and Kristen Hamanishi
15 who willingly made available their technical expertise, unique facility for virologic
16 investigation and analysis which was critical to successful laboratory reconstruction
17 of the events surrounding the seroconversion case which occurred.

18 Some of the results of this study were presented at the 19th Conference on
19 Retroviruses and Opportunistic Infections, 5–8 March 2012, Seattle, WA. The study
20 manuscript is published in Clinical Pharmacology and Therapeutics, June 2014. The

1 seroconversion case is published in the Journal of Infectious Diseases November
2 2015.

3 SSAT049 was supported by a grant from Janssen Pharmaceuticals. Preliminary
4 results from this study were presented at the 21st Conference on Retroviruses and
5 Opportunistic Infections, Boston MA, 3-6 March, 2014.

6 SSAT055 as a clinical study was supported by a funding grant held by Professors
7 Owen and Rannard at the University of Liverpool. Portions of the results from this
8 study were presented at the Conference on Retroviruses and Opportunistic
9 Infections, Seattle WA February 13-16, 2017.

10

11

1 9 REFERENCES

2

3 Achhra AC, Mwasakifwa G, Amin J, Boyd MA. Efficacy and safety of contemporary
4 dual-drug antiretroviral regimens as first-line treatment or as a simplification
5 strategy: a systematic review and meta-analysis. *The Lancet HIV*. 2016
6 Aug;3(8):e351–60.

7 Amara AB, Tjia J, Dutton J, Else LJ, Back DJ, Khoo S. Development and validation of a
8 HPLC-MS/MS assay to quantify the antiretroviral (ARV) drug, efavirenz and its
9 major metabolites in plasma. British Mass Spectrometry Society Meeting.
10 Cardiff; 2011.

11 Anta L, Llibre JM, Poveda E, Blanco JL, Alvarez M, Pérez-Elías MJ, et al. Rilpivirine
12 resistance mutations in HIV patients failing non-nucleoside reverse transcriptase
13 inhibitor-based therapies. *AIDS*. 2013 Jan 2;27(1):81–5.

14 Arribas JR, Girard P-M, Landman R, Pich J, Mallolas J, Martínez-Rebollar M, et al.
15 Dual treatment with lopinavir-ritonavir plus lamivudine versus triple treatment
16 with lopinavir-ritonavir plus lamivudine or emtricitabine and a second
17 nucleos(t)ide reverse transcriptase inhibitor for maintenance of HIV-1 viral
18 suppression (OLE): a randomised, open-label, non-inferiority trial. *The Lancet*
19 *infectious diseases*. 2015 Jul;15(7):785–92.

20 Azijn H, Tirry I, Vingerhoets J, de Béthune M-P, Kraus G, Boven K, et al. TMC278, a
21 next-generation nonnucleoside reverse transcriptase inhibitor (NNRTI), active
22 against wild-type and NNRTI-resistant HIV-1. *Antimicrobial Agents and*
23 *Chemotherapy*. 2010 Feb 1;54(2):718–27.

24 Baert L, van 't Klooster G, Dries W, ois MF, Wouters A, Basstanie E, et al.
25 Development of a long-acting injectable formulation with nanoparticles of
26 rilpivirine (TMC278) for HIV treatment. *Eur J Pharm Biopharm*. 2009 Jul
27 31;72(3):502–8.

28 Baeten JM, Donnell D, Ndase P, Mugo NR, Campbell JD, Wangisi J, et al.
29 Antiretroviral Prophylaxis for HIV Prevention in Heterosexual Men and Women.
30 *N Engl J Med*. 2012 Jul 11;367(5):120711140017009–410.

31 Bannister WP, Ruiz L, Cozzi-Lepri A, Mocroft A, Kirk O, Staszewski S, et al.
32 Comparison of genotypic resistance profiles and virological response between
33 patients starting nevirapine and efavirenz in EuroSIDA. *AIDS*. 2008 Jan
34 30;22(3):367–76.

- 1 Basson AE, Rhee S-Y, Parry CM, El-Khatib Z, Charalambous S, De Oliveira T, et al.
2 Impact of drug resistance-associated amino acid changes in HIV-1 subtype C on
3 susceptibility to newer nonnucleoside reverse transcriptase inhibitors.
4 Antimicrob Agents Chemother. American Society for Microbiology; 2015
5 Feb;59(2):960–71.
- 6 Baxter JD, Dunn D, White E, Sharma S, Geretti AM, Kozal MJ, et al. Global HIV-1
7 transmitted drug resistance in the INSIGHT Strategic Timing of AntiRetroviral
8 Treatment (START) trial. HIV Medicine. 2015 Apr;16 Suppl 1(Suppl 1):77–87.
- 9 Bedimo RJ, Drechsler H, Jain M, Cutrell J, Zhang S, Li X, et al. The RADAR study: week
10 48 safety and efficacy of RAltegravir combined with boosted DARunavir
11 compared to tenofovir/emtricitabine combined with boosted darunavir in
12 antiretroviral-naïve patients. Impact on bone health. Law M, editor. PLoS ONE.
13 2014;9(8):e106221.
- 14 Boffito M, Lamorde M, Watkins M, Pozniak A. Antiretroviral dose optimization: the
15 future of efavirenz 400 mg dosing. Curr Opin HIV AIDS. Current Opinion in HIV
16 and AIDS; 2017 Jul;12(4):339–42.
- 17 Cahn P, Andrade-Villanueva J, Arribas JR, Gatell JM, Lama JR, Norton M, et al. Dual
18 therapy with lopinavir and ritonavir plus lamivudine versus triple therapy with
19 lopinavir and ritonavir plus two nucleoside reverse transcriptase inhibitors in
20 antiretroviral-therapy-naïve adults with HIV-1 infection: 48 week results of the
21 randomised, open label, non-inferiority GARDEL trial. The Lancet infectious
22 diseases. 2014 Jul;14(7):572–80.
- 23 Cahn P, Rolón MJ, Figueroa MI, Gun A, Patterson P, Sued O. Dolutegravir-lamivudine
24 as initial therapy in HIV-1 infected, ARV-naïve patients, 48-week results of the
25 PADDLE (Pilot Antiretroviral Design with Dolutegravir LamivudinE) study. J Int
26 AIDS Soc. 2017 May 9;20(1):1–7.
- 27 Churchill D, Waters L, Ahmed N, Angus B, Boffito M, Bower M, et al. British HIV
28 Association guidelines for the treatment of HIV-1-positive adults with
29 antiretroviral therapy 2015. HIV Medicine. 2016 Aug;17 Suppl 4(Suppl 1):s2–
30 s104.
- 31 Cohen CJ, Molina J-M, Cassetti I, Chetchotisakd P, Lazzarin A, Orkin C, et al. Week 96
32 efficacy and safety of rilpivirine in treatment-naïve, HIV-1 patients in two Phase
33 III randomized trials. AIDS. 2013 Mar 27;27(6):939–50.
- 34 Cohen MS, Chen YQ, McCauley M, Gamble T, Hosseinipour MC, Kumarasamy N, et
35 al. Prevention of HIV-1 infection with early antiretroviral therapy. N Engl J Med.
36 2011 Aug 11;365(6):493–505.
- 37 Dalgleish AG, Moyle GJ, Easterbrook P, Gazzard BG. AIDS: clinical and scientific
38 issues past, present and future. Q J Nucl Med. 1995 Sep;39(3):156–62.

- 1 Dickinson L, Amin J, Else L, Boffito M, Egan D, Owen A, et al. Comprehensive
2 Pharmacokinetic, Pharmacodynamic and Pharmacogenetic Evaluation of Once-
3 Daily Efavirenz 400 and 600 mg in Treatment-Naïve HIV-Infected Patients at
4 96 Weeks: Results of the ENCORE1 Study. Clin Pharmacokinet. Springer
5 International Publishing; 2016 Jul;55(7):861–73.
- 6 Dickinson L, Boffito M, Back D, Else L, Hentig von N, Davies G, et al. Sequential
7 population pharmacokinetic modeling of lopinavir and ritonavir in healthy
8 volunteers and assessment of different dosing strategies. Antimicrob Agents
9 Chemother. American Society for Microbiology; 2011 Jun;55(6):2775–82.
- 10 Dimitrov DT, Boily M-C, Hallett TB, Albert J, Boucher C, Mellors JW, et al. How Much
11 Do We Know about Drug Resistance Due to PrEP Use? Analysis of Experts'
12 Opinion and Its Influence on the Projected Public Health Impact. Apetrei C,
13 editor. PLoS ONE. 2016;11(7):e0158620.
- 14 Eisingerich AB, Wheelock A, Gomez GB, Garnett GP, Dybul MR, Piot PK. Attitudes
15 and acceptance of oral and parenteral HIV preexposure prophylaxis among
16 potential user groups: a multinational study. Tachedjian G, editor. PLoS ONE.
17 2012;7(1):e28238.
- 18 Else LJ, Tjia J, Jackson AGA, Panchala SD, Egan D, Boffito M, et al. Quantification of
19 rilpivirine in human plasma, cervicovaginal fluid, rectal fluid and genital/rectal
20 mucosal tissues using liquid chromatography–tandem mass spectrometry.
21 Bioanalysis. 2014 Jul;6(14):1907–21.
- 22 Else LJJ, Jackson AGA, Puls RR, Hill AA, Fahey PP, Lin EE, et al. Pharmacokinetics of
23 Lamivudine and Lamivudine-Triphosphate after Administration of 300
24 Milligrams and 150 Milligrams Once Daily to Healthy Volunteers: Results of the
25 ENCORE 2 Study. Antimicrob Agents Chemother. 2012 Mar 1;56(3):1427–33.
- 26 Else LL, Else L, Watson VV, Watson V, Tjia J, Tjia JJ, et al. Validation of a rapid and
27 sensitive high-performance liquid chromatography-tandem mass spectrometry
28 (HPLC-MS/MS) assay for the simultaneous determination of existing and new
29 antiretroviral compounds. J Chromatogr B Analyt Technol Biomed Life Sci. 2010
30 Jun 1;878(19):1455–65.
- 31 European Medicines Agency. Committee for Medicinal Products for Human Use
32 (CHMP) Guideline on the Investigation of Bioequivalence. ema.europa.eu Jan
33 20, 2010.
- 34 Foca M, Yogev R, Wiznia A, Hazra R, Jean-Philippe P, Graham B, et al. Rilpivirine
35 Pharmacokinetics Without and With Darunavir/Ritonavir Once Daily in
36 Adolescents and Young Adults. Pediatr Infect Dis J. 2016 Sep;35(9):e271–4.
- 37 Ford N, Lee J, Andrieux-Meyer I, Calmy A. Safety, efficacy, and pharmacokinetics of
38 rilpivirine: systematic review with an emphasis on resource-limited settings. HIV.
39 2011 Apr;3:35–44.

1 Gazzard BG. HIV disease and the gastroenterologist. *Gut*. 1988 Nov;29(11):1497–
2 505.

3 Gilead Sciences Inc. Viread 245 mg film-coated tablets - Summary of Product
4 Characteristics (SPC) - (eMC) [Internet]. medicines.org.uk. 2002 [cited 2017 Sep
5 26]. Available from: <https://www.medicines.org.uk/emc/medicine/9008>

6 Grant N, Zhang H. Poorly water-soluble drug nanoparticles via an emulsion-freeze-
7 drying approach. *Journal of Colloid and Interface Science*. 2011 Apr;356(2):573–
8 8.

9 Grant RM, Lama JR, Anderson PL, McMahan V, Liu AY, Vargas L, et al. Preexposure
10 Chemoprophylaxis for HIV Prevention in Men Who Have Sex with Men. *N Engl J*
11 *Med*. 2010;363(27):2587–99.

12 Holzinger ER, Grady B, Ritchie MD, Ribaud HJ, Acosta EP, Morse GD, et al. Genome-
13 wide association study of plasma efavirenz pharmacokinetics in AIDS Clinical
14 Trials Group protocols implicates several CYP2B6 variants. *Pharmacogenet*
15 *Genomics*. 2012 Dec;22(12):858–67.

16 Jackson AGA, Else L, Higgs C, Karolia Z, Khoo S, Back D, et al. Pharmacokinetics and
17 pharmacodynamics of the nucleoside sparing dual regimen containing rilpivirine
18 plus darunavir/ritonavir in treatment-naïve HIV-1-infected individuals. *HIV*
19 *clinical trials*. Taylor & Francis; 2017 Nov 30;:1–7.

20 Jackson AGA, Else LJ, Mesquita PMM, Egan D, Back DJ, Karolia Z, et al. A
21 compartmental pharmacokinetic evaluation of long-acting rilpivirine in HIV
22 negative volunteers for pre-exposure prophylaxis (PrEP). *Clin Pharmacol Ther*.
23 2014 May 26;96(3):314–23.

24 Jackson AGA, Hill A, Puls R, Else L, Amin J, Back D, et al. Pharmacokinetics of plasma
25 lopinavir/ritonavir following the administration of 400/100 mg, 200/150 mg and
26 200/50 mg twice daily in HIV-negative volunteers. *J Antimicrob Chemother*.
27 2011 Mar;66(3):635–40.

28 Janssen-Cilag Ltd. Edurant 25 mg tablets - Summary of Product Characteristics (SPC)
29 - (eMC) [Internet]. medicines.org.uk. 2011 [cited 2017 Sep 27]. Available from:
30 <https://www.medicines.org.uk/emc/medicine/25490>

31 Janssen-Cilag Ltd. REZOLSTA 800 mg/150 mg film coated tablets - Summary of
32 Product Characteristics (SPC) - (eMC) [Internet]. medicines.org.uk. 2014 [cited
33 2017 Sep 28]. Available from:
34 <https://www.medicines.org.uk/emc/medicine/29480>

35 Janssen-Cilag Ltd. PREZISTA 800 mg film coated tablets - Summary of Product
36 Characteristics (SPC) - (eMC) [Internet]. medicines.org.uk. 2015 [cited 2017 Sep
37 27]. Available from: <https://www.medicines.org.uk/emc/medicine/31092>

- 1 Janssen-Cilag Ltd. Symtuza - European public assessment report (EPAR) European
2 Medicines Agency. ema.europa.eu. 2017 Sep.
- 3 Kozal MJM, Lupo SS, Dejesus EE, Molina J-MJ, McDonald CC, Raffi FF, et al. A
4 Nucleoside- and Ritonavir-Sparing Regimen Containing Atazanavir Plus
5 Raltegravir in Antiretroviral Treatment-Naïve HIV-Infected Patients: SPARTAN
6 Study Results. *HIV clinical trials*. 2012 Jan 1;13(3):119–30.
- 7 Lambert-Niclot S, Charpentier C, Storto A, Fofana D, Soulié C, Fourati S, et al.
8 Rilpivirine, emtricitabine and tenofovir resistance in HIV-1-infected rilpivirine-
9 naive patients failing antiretroviral therapy. *Journal of Antimicrobial*
10 *Chemotherapy*. 2014 Apr;69(4):1086–9.
- 11 Lodi S, Sharma S, Lundgren JD, Phillips AN, Cole SR, Logan R, et al. The per-protocol
12 effect of immediate versus deferred antiretroviral therapy initiation. *AIDS*
13 [Internet]. *AIDS*; 2016 Nov 13;30(17):2659–63. Available from:
14 <http://Insights.ovid.com/crossref?an=00002030-201611130-00012>
- 15 Maggiolo F, Di Filippo E, Valenti D, Serna Ortega PA, Callegaro A. NRTI Sparing
16 Therapy in Virologically Controlled HIV-1 Infected Subjects: Results of a
17 Controlled, Randomized Trial (Probe). *J Acquir Immune Defic Syndr*. 2016 May
18 1;72(1):46–51.
- 19 Marrazzo J, Ramjee G, Nair G, Palanee T, Mkhize B, Nakabiito C, et al. Pre-exposure
20 Prophylaxis for HIV in Women: Daily Oral Tenofovir, Oral
21 Tenofovir/Emtricitabine, or Vaginal Tenofovir Gel in the VOICE Study (MTN 003).
22 20th Conference on Retroviruses and Opportunistic Infections (CROI)
23 [Internet]. Atlanta, GA; 2013. Available from:
24 <http://www.retroconference.org/2013b/Abstracts/47951.htm>
- 25 Marrazzo JM, Ramjee G, Richardson BA, Gomez K, Mgodini N, Nair G, et al. Tenofovir-
26 based preexposure prophylaxis for HIV infection among African women. *N Engl J*
27 *Med*. 2015 Feb 5;372(6):509–18.
- 28 McGowan I, Dezzutti CS, Siegel A, Engstrom J, Nikiforov A, Duffill K, et al. Long-acting
29 rilpivirine as potential pre-exposure prophylaxis for HIV-1 prevention (the
30 MWRI-01 study): an open-label, phase 1, compartmental, pharmacokinetic and
31 pharmacodynamic assessment. *The Lancet HIV*. 2016 Dec;3(12):e569–78.
- 32 Mesquita PM, Srinivasan P, Johnson TJ, Rastogi R, Evans-Strickfaden T, Kay MS, et al.
33 Novel preclinical models of topical PrEP pharmacodynamics provide rationale
34 for combination of drugs with complementary properties. *Retrovirology*.
35 2013;10(1):113.
- 36 Meyers K, Rodriguez K, Moeller RW, Gratch I, Markowitz M, Halkitis PN. High
37 interest in a long-acting injectable formulation of pre-exposure prophylaxis for
38 HIV in young men who have sex with men in NYC: a P18 cohort substudy.
39 Garcia-Lerma JG, editor. *PLoS ONE*. 2014;9(12):e114700.

- 1 Molina J-M, Clumeck N, Redant K, Rimsky L, Vanveggel S, Stevens M, et al. Rilpivirine
2 versus efavirenz in HIV-1 patients with baseline viral load $\leq 100\,000$ copies/ml:
3 week 48 Phase III analysis. *AIDS*. 2012 Dec 29;:-.
- 4 Morrison CS, Chen P-L, Kwok C, Baeten JM, Brown J, Crook AM, et al. Hormonal
5 contraception and the risk of HIV acquisition: an individual participant data
6 meta-analysis. *Beyrer C, editor. PLoS Med*. 2015 Jan;12(1):e1001778.
- 7 Moyle GJ, Back D. Principles and practice of HIV-protease inhibitor
8 pharmacoenhancement. *HIV Med*. 2001 Apr;2(2):105–13.
- 9 Panel on Antiretroviral Guidelines for Adults and Adolescents. **Guidelines for the**
10 **Use of Antiretroviral Agents in HIV-1-Infected Adults and Adolescents.**
11 *aidsinfo.nih.gov*. 2016 Jul.
- 12 Parczewski M, Urbańska A, Maciejewska K, Witak-Jędra M, Leszczyszyn-Pynka M.
13 Transmitted drug resistance to rilpivirine among antiretroviral-naïve patients
14 living with HIV from northern Poland. *J Int AIDS Soc. The International AIDS*
15 *Society*; 2014;17(1):18929.
- 16 Penrose KJ, Parikh UM, Hamanishi KA, Else L, Back D, Boffito M, et al. Selection of
17 Rilpivirine-Resistant HIV-1 in a Seroconverter From the SSAT 040 Trial Who
18 Received the 300-mg Dose of Long-Acting Rilpivirine (TMC278LA). *Journal of*
19 *Infectious Diseases*. Oxford University Press; 2015 Nov 12;213(6):jiv528–1017.
- 20 Raffi F, Babiker AG, Richert L, Molina J-M, George EC, Antinori A, et al. Ritonavir-
21 boosted darunavir combined with raltegravir or tenofovir–emtricitabine in
22 antiretroviral-naïve adults infected with HIV-1: 96 week results from the
23 NEAT001/ANRS143 randomised non-inferiority trial. *The Lancet*. Elsevier; 2014
24 Nov;384(9958):1942–51.
- 25 Sanford M. Rilpivirine. *Drugs*. 2012 Mar 5;72(4):525–41.
- 26 Santoscoy M, Cahn P, Gonzalez C, Hao W, Pozniak A, Shalit P, et al. TMC278
27 (rilpivirine), a next-generation NNRTI, demonstrates long-term efficacy and
28 tolerability in ARV-naïve patients: 96-week results of study C204. XVIIth
29 International AIDS Conference. Mexico City, MX; 2008.
- 30 Sax PE, Tierney C, Collier AC, Fischl MA, Mollan K, Peeples L, et al. Abacavir-
31 lamivudine versus tenofovir-emtricitabine for initial HIV-1 therapy. *N Engl J*
32 *Med*. Massachusetts Medical Society; 2009 Dec 3;361(23):2230–40.
- 33 Sherer R. Darunavir in the treatment of HIV-1 infection: a viewpoint by Renslow
34 Sherer. *Drugs*. 2007;67(18):2802–3.
- 35 Strategies for Management of Antiretroviral Therapy SMART Study Group, El-Sadr
36 WM, Lundgren JD, Neaton JD, Gordin F, Abrams D, et al. CD4+ count-guided

1 interruption of antiretroviral treatment. *N Engl J Med*. 2006 Nov
2 30;355(22):2283–96.

3 Sugano K, Kansy M, Artursson P, Avdeef A, Bendels S, Di L, et al. Coexistence of
4 passive and carrier-mediated processes in drug transport. *Nature reviews Drug*
5 *discovery*. 2010 Jul 31;9(8):597–614.

6 Taiwo B, Zheng L, Gallien S, Matining RM, Kuritzkes DR, Wilson CC, et al. Efficacy of a
7 nucleoside-sparing regimen of darunavir/ritonavir plus raltegravir in treatment-
8 naive HIV-1-infected patients (ACTG A5262). *AIDS*. 2011 Nov 13;25(17):2113–
9 22.

10 Theys K, Camacho RJ, Gomes P, Vandamme AM, Rhee SY, Portuguese HIV-1
11 Resistance Study Group. Predicted residual activity of rilpivirine in HIV-1
12 infected patients failing therapy including NNRTIs efavirenz or nevirapine. *Clin*
13 *Microbiol Infect*. 2015 Jun;21(6):607.e1–8.

14 Thigpen MC, Kebaabetswe PM, Paxton LA, Smith DK, Rose CE, Segolodi TM, et al.
15 Antiretroviral Preexposure Prophylaxis for Heterosexual HIV Transmission in
16 Botswana. *N Engl J Med*. 2012 Jul 11;:120711140017009.

17 van 't Klooster G, Hoeben E, Borghys H, Looszova A, Bouche M-P, van Velsen F, et al.
18 Pharmacokinetics and Disposition of Rilpivirine (TMC278) Nanosuspension as a
19 Long-Acting Injectable Antiretroviral Formulation. *Antimicrobial Agents and*
20 *Chemotherapy*. 2010 Apr 19;54(5):2042–50.

21 Van Damme L, Corneli A, Ahmed K, Agot K, Lombaard J, Kapiga S, et al. Preexposure
22 prophylaxis for HIV infection among African women. *N Engl J Med*. 2012 Aug
23 2;367(5):411–22.

24 van der Straten A, Van Damme L, Haberer JE, Bangsberg DR. Unraveling the
25 divergent results of pre-exposure prophylaxis trials for HIV prevention. *AIDS*.
26 2012 Apr 23;26(7):F13–9.

27 van't Klooster G, Hoeben E, Borghys H, Looszova A, Bouche M-P, van Velsen F, et al.
28 Nanosuspension of rilpivirine (TMC278) as a long-acting injectable antiretroviral
29 formulation: pharmacokinetics and disposition. *Antimicrobial Agents and*
30 *Chemotherapy*. *Am Soc Microbiol*; 2010.

31 ViiV Healthcare. Ziagen 300 mg Film Coated Tablets - Summary of Product
32 Characteristics (SPC) - (eMC) [Internet]. *medicines.org.uk*. 2000 [cited 2017 Sep
33 27]. Available from: <https://www.medicines.org.uk/emc/medicine/2476>

34 ViiV Healthcare. Triumeq 50mg, 300mg, 600mg Tablets - Summary of Product
35 Characteristics (SPC) - (eMC) [Internet]. *medicines.org.uk*. 2014 [cited 2017 Sep
36 26]. Available from: <https://www.medicines.org.uk/emc/medicine/29178>

- 1 Wensing AM, Calvez V, Günthard HF, Johnson VA, Paredes R, Pillay D, et al. 2014
2 Update of the drug resistance mutations in HIV-1. *Top Antivir Med.* NIH Public
3 Access; 2014 Jun;22(3):642–50.
- 4 World Health Organisation. Consolidated Guidelines on the Use of Antiretroviral
5 Drugs for Treating and Preventing HIV Infection [Internet]. Geneva, Switzerland;
6 2013 [cited 2014 Apr 24]. Available from:
7 [http://apps.who.int/iris/bitstream/10665/85322/1/WHO_HIV_2013.7_eng.pdf?](http://apps.who.int/iris/bitstream/10665/85322/1/WHO_HIV_2013.7_eng.pdf?ua=1)
8 [ua=1](http://apps.who.int/iris/bitstream/10665/85322/1/WHO_HIV_2013.7_eng.pdf?ua=1)
- 9 Zhang H, Wang D, Butler R, Campbell NL, Long J, Tan B, et al. Formation and
10 enhanced biocidal activity of water-dispersable organic nanoparticles. *Nat*
11 *Nanotechnol.* Nature Publishing Group; 2008 Aug;3(8):506–11.
- 12